

Limited impact of on native algal assemblages from rocky intertidal shores

Celia Olabarria, Iván F. Rodil, Mónica Incera, Jesús S. Troncoso

▶ To cite this version:

Celia Olabarria, Iván F. Rodil, Mónica Incera, Jesús S. Troncoso. Limited impact of on native algal assemblages from rocky intertidal shores. Marine Environmental Research, 2009, 67 (3), pp.153. 10.1016/j.marenvres.2008.12.007 . hal-00501991

HAL Id: hal-00501991 https://hal.science/hal-00501991

Submitted on 13 Jul 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

Limited impact of *Sargassum muticum* on native algal assemblages from rocky intertidal shores

Celia Olabarria, Iván F. Rodil, Mónica Incera, Jesús S. Troncoso

PII:	S0141-1136(08)00262-6
DOI:	10.1016/j.marenvres.2008.12.007
Reference:	MERE 3311
To appear in:	Marine Environmental Research
Received Date:	3 October 2008
Revised Date:	18 December 2008
Accepted Date:	22 December 2008



Please cite this article as: Olabarria, C., Rodil, I.F., Incera, M., Troncoso, J.S., Limited impact of *Sargassum muticum* on native algal assemblages from rocky intertidal shores, *Marine Environmental Research* (2008), doi: 10.1016/j.marenvres.2008.12.007

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Limited impact of *Sargassum muticum* on native algal assemblages from rocky intertidal shores

Celia Olabarria^{1*}, Iván F. Rodil¹, Mónica Incera², Jesús S. Troncoso¹

¹Departamento de Ecología e Biología Animal, Facultade de Ciencias del Mar,

Universidade de Vigo, 36200Vigo, Spain.

²Centro Tecnológico del Mar-Fundación CETMAR, C/Eduardo Cabello s/n, 36208

Vigo, Spain

-

*Corresponding author: colabarria@uvigo.es

Color

1 ABSTRACT

2	Invasive species represent a serious threat to natural ecosystems through a
3	range of negative effects on native species in the region invaded. The invasive species
4	Sargassum muticum has invaded several temperate regions worldwide including the
5	Galician rocky shoreline (northwestern Spain) in western Europe. The main aim of
6	this study was to assess if colonization by <i>S. muticum</i> has any effect on native algal
7	assemblages by experimental removal of S. muticum. We predicted that in those plots
8	where S. muticum plants were removed, the structure of native algal assemblages
9	would differ from that in plots where S. muticum plants were untouched. In addition,
10	we predicted that the effect of Sargassum removal would be more important than
11	other causes of variability at the small scale investigated. Results indicated limited
12	impact of S. muticum on native assemblages. The impact was only evident on the total
13	number of native taxa and two understory morpho-functional groups, filamentous and
14	foliose algae, rather than on the entire macroalgal assemblages.
15	

Key words: removal experiment, algal assemblages, biological invasions, rocky

17 intertidal, Sargasum muticum effects

1. Introduction

28	The number of species anthropogenically introduced outside their natural
29	ranges is rapidly increasing (Levin et al., 2002). Although only a very small
30	proportion of transported and introduced species becomes invasive, the ability of
31	these species to restructure and, therefore, change the functioning of a recipient
32	habitat is great (Crooks, 2002). After loss of habitat, invasive species are the second
33	leading cause of biodiversity loss, particularly in freshwater ecosystems (Vitousek et
34	al., 1997; Mack et al., 2000; Ross et al., 2004). Besides imposing huge economic
35	costs, invasive species may cause extinction of native species, reduction of genetic
36	diversity, and biotic homogenization through a variety of mechanisms (Rahel, 2000).
37	Biological invasions are increasing, especially in marine environments where
38	they are linked to the intensification of international shipping, aquaculture and
39	aquarium activity (Verlaque, 1994; Ribera and Bouderesque, 1995). In particular,
40	marine macroalgae are a significant component of marine invasive species with 277
41	introduced species recorded in a recent review (see Williams and Smith, 2007).
42	Displacement of existing flora by introduced algal species has been largely
43	reported (Chambers et al., 1993; Piazzi et al., 2001). Replacement of dominant
44	macroalgae by introduced species may provoke changes of the species composition
45	and their trophic food webs (Piazzi et al., 2001). Several studies have focused on the
46	effects of exotic algae and impacts on native assemblages (e.g. Viejo, 1997; Stæhr et
47	al., 2000; Piazzi et al., 2001; Levin et al., 2002; Britton-Simmons, 2004; Sánchez and
48	Fernández, 2005; Strong et al., 2006). Results indicate that impacts of exotic species
49	may vary greatly depending on the invaded assemblages (e.g. Chapman, 1999). For
50	example, the Japanese seaweed Codium fragile (Sur.) Harriot ssp. tomentosoides (van
51	Goor) Silva has detrimental effects on subtidal kelp communities in the northeast Atlantic
52	Ocean while on the European intertidal shores it occurs in low abundances with no effects on

53 native assemblages (Buschbaum et al., 2006). The green alga Caulerpa racemosa 54 (Forsskål) J. Agardh has strong effects on composition and structure of native 55 assemblages at different habitats such as rocky bottoms and dead matter of the marine 56 phanerogam Posidonia oceanica (Piazzi et al., 2001). Sargassum muticum (Yendo) 57 Fensholt is native to SE Asia, but its present distribution as an invasive species is 58 widespread, including Europe, the Mediterranean Sea and the west coast of North 59 America. This species becomes established in low intertidal and shallow subtidal 60 habitats where it forms dense stands that may reduce light, dampen flow, increase 61 sedimentation and reduce ambient nutrient concentrations available for native species 62 (Britton-Simmons, 2004). In subtidal habitats S. muticum has been shown to have 63 negative effects on native algae (Sánchez and Fernández, 2005), whereas in intertidal 64 areas some studies suggest that it competes with native algae (e.g. DeWreede, 1983; 65 Viejo, 1997) while other studies show no effects. These contradicting results suggest 66 strong site-specific effects of S. muticum, making generalizations about the impact of 67 this species difficult. 68 S. muticum was first observed on the Galician coast in 1986 (see Pérez-Cirera 69 et al., 1989) and has since quickly colonized most of the Galician rias (sensu Vilas et

 70
 al., 2005) increasing its abundance rapidly. Despite its rapid spread, no studies

assessing consequences of the introduction of this species on native assemblages havebeen done in this area so far.

The objective of this study was to assess the effect of *S. muticum* on native algal assemblages using a removal experiment in which the presence of this species was manipulated. We predicted that if *S. muticum* is removed the remaining assemblages would be distinct, in terms of composition and structure, from those found in plots where *S. muticum* was present. In addition, we predicted that the effect

of removal would be more important than other causes of variability at the smallspatial scale investigated (sites located 400 m apart).

80 2. Material and Methods

81 *2.1. Study area*

82	The study was carried out at Monte Lourido (42° 7' 26'' N; 8° 49' 34'' W) in
83	the Galician coast (NW Spain). Two sites 400 m apart were chosen. Site 1 was on a
84	gently sloping rocky platform (~ 2° in slope) running perpendicular to the sea and
85	slightly more exposed than site 2. The upper intertidal was dominated by Fucus
86	spiralis and the mid and low intertidal was dominated by red, green or some
87	conspicuous brown macroalgae (i.e. Sacchoriza polyschides) depending on the
88	season. Site 2 was a long nearly horizontal platform interrupted at its seaward margin
89	by a series of surge channels that run oblique to the shoreline and are sheltered from
90	the direct wave action. The bottom of these channels was made of cobbles covered
91	with sand. The upper intertidal contained less F. spiralis than Site 1 and mid and low
92	intertidal assemblages were poorer in terms of number of species than Site 1. S.
93	muticum was very abundant at both sites, although plants from Site 2 were bit longer
94	and reached larger sizes than those at Site 1 (personal observation).

95 2.2. Experimental design

96 The experiment was performed in the low intertidal (0.4-0.8 m above the 97 Iowest astronomical tide) from April 2006 to March 2008. Ten permanent 50 x 50 cm 98 plots were randomly establish along 80 m at each site. The corners of the plots were 99 permanently marked using epoxy putty (IVEGOR) for identification. At each site, the 100 experimental plots were arranged in a line parallel to shore separated by ~ 5 m and 101 randomly assigned to control (i.e. with *Sargassum*) or removal (i.e. without

102	Sargassum) treatments with five replicates for each. In plots assigned to the removal
103	treatment, all S. muticum was manually eradicated, both from within the plots and
104	from a 50 cm buffer surrounding the plots. Each plant was removed by carefully
105	prying its holdfast off the rock substrate with a scrapper. The buffer zone was created
106	in order to reduce shading or other effects of adjacent S. muticum. Attention was paid
107	to not damage the understory assemblage or to alter the physical characteristics of the
108	substratum. The removal plots were kept free of S. muticum throughout the course of
109	experiment by removing any new S. muticum recruits from plots and adjacent buffer
110	zones once a month. The control plots were not altered. The experiment was sampled
111	once prior to the beginning of the experiment and then every month, although time
112	and weather constraints prevented sampling on some dates (site 1 in October 2006 and
113	site 2 in November 2006, January and August 2007).
114	Percentage cover of macroalgae was estimated using a 50 x 50 cm metallic
115	quadrat divided into 25 sub-quadrats of 10×10 cm, and a score from 0 to 4% was
116	given to each taxon in each sub-quadrat after visual inspection. Final cover was
117	obtained by summing over the 25 scores obtained in the sub-quadrats (Dethier et al.,
118	1993). Species were identified in the field but when doubt existed, samples were
119	collected for further identification in the laboratory. Macroalgae were grouped into
120	morpho-functional groups based on similarities in morphology and resource use
121	(foliose, filamentous, corticated, leathery, articulated calcareus and encrusting algae;
122	sensu Steneck and Dethier, 1994). In addition, number of native species and diversity
123	(H') were estimated for each plot.

124 2.3. Statistical analysis

Since experimental plots were repeatedly sampled over time (i.e. 20 sampling
dates), a technique that could account for temporal autocorrelation was necessary.

127 Data (percentage cover of main morpho-functional groups, number of native taxa and 128 diversity) were analysed using Population-Averaged Generalized Estimating 129 Equations (PA-GEEs), an extension of GLMs for correlated data (Liang and Zeger, 130 1986). PA-GEEs were run using the "geeglm" procedure in the R program, version 131 R2.2.1 (R Development Core Team 2005), specifying a link function and an error 132 structure for the residuals. We specified a first order autoregressive model AR (1) in 133 all the analyses assuming time dependence for each plot. For data expressed as 134 percentage cover and diversity, a Gaussian distribution of the error terms was 135 assumed and we used an identity link function. For data expressed as number of algal 136 species, a Poisson distribution of the error terms was assumed, and we used a log-link 137 function.

138 Two-way crossed analysis of similarity (ANOSIM) (Clarke, 1993) on fourth 139 root transformed data of percentage cover for each species was used to test for 140 differences in assemblages between treatments and over time at each site separately. 141 Treatment with two levels (control and removal) and time with 10 levels (April, July, 142 September, December 2006 and 2007, and February 2007 and 2008) were the factors 143 in the analysis. Pair-wise comparisons were also calculated. In addition, responses of 144 assemblages to treatments were visualised by non-metric multidimensional scaling 145 (nMDS) on the basis of Bray-Curtis similarity matrices on fourth-root transformed 146 percentage cover data. For all analyses, S. muticum was omitted from the data set to 147 check if the possible differences between treatments were caused not by the presence 148 of this species, but by changes in the remaining assemblages. Species that mostly 149 contributed to the dissimilarity/similarity between the two experimental treatments 150 were identified using SIMPER analysis (Clarke, 1993). All the calculations were 151 performed using PRIMER 6 for Windows (Plymouth, UK).

152 **3. Results**

153	The percentage cover of S. muticum varied significantly between treatments
154	(PA-GEEs; treatment: estimate: -15.36, standard error: 2.58, $p < 0.001$). Thus in
155	control plots the mean percentage cover of S. muticum was 29.3 ± 2.7 and 42.7 ± 3 % at
156	site 1 and site 2, respectively, whereas in removal treatments the mean percentage
157	cover of the alga was 5±1.3 and 7.2±1.7 % at site 1 and site 2, respectively (Fig. 1).
158	3.1. Effects of S. muticum on morpho-functional groups
159	Removal of S. muticum resulted in a significant increase of the mean
160	percentage cover of filamentous and foliose algae ($p < 0.05$; Table 1, Fig. 2 a,b). Mean
161	percentage cover of encrusting algae also increased in those plots where S. muticum
162	was removed, but this increase was marginally non-significant ($p=0.056$; Table 1,
163	Fig. 2c). The removal also affected the mean number of taxa (used as a surrogate
164	measure of native species diversity), with significant lower values occurring in control
165	plots, especially at site 2 ($p < 0.05$; Table 1, Fig. 2 d). The effect of S. muticum on
166	these morpho-functional groups was also consistent across sites (i.e. non-significant
167	interaction treatment x site, $p > 0.05$; Table 1). Diversity and percentage cover of
168	corticated, leathery and calcareous algae were not affected significantly by treatments
169	(neither as main effect nor in interaction with site).
170	3.2. Changes in assemblages
171	Removal of <i>S. muticum</i> did not affect the composition of macroalgal
172	assemblages. Two-way crossed ANOSIM results showed signifant differences
173	between sites over time (Table 2, Fig. 3), but not between treatments (differences

- between treatments: site 1, R=0.04, p>0.05; site 2, R=0.08, p>0.05 averaged across
- all sampling times). There were only significant differences between treatments in

177macroalgae assemblage structure (Fig. 3) were also detected by ANOSIM at both si178(differences among time factor: site 1, R=0.519, $p<0.01$; site 2, R=0.465, $p<0.01$;179Table 2; Figure 3). Pair-wise comparisons detected differences in macroalgal180assemblages in most cases (Table 2).181The foliose alga <i>Ulva rigida</i> (7.07%), the encrusting alga <i>Lithophyllum</i> 182 <i>incrustans</i> (6.62%), the leathery alga <i>Bifurcaria bifurcata</i> (6.17%) and the articulate183calcareus alga <i>Corallina officinalis</i> (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.</i> 186 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188 <i>S. muticum</i> was very abundant in control plots during the whole experiment,190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of the196native species (PA-GEEs: $p> 0.05$) except	176	April and September 2007 (Table 2). Time (monthly and interannually) differences in
178(differences among time factor: site 1, R=0.519, $p<0.01$; site 2, R=0.465, $p<0.01$;179Table 2; Figure 3). Pair-wise comparisons detected differences in macroalgal180assemblages in most cases (Table 2).181The foliose alga <i>Ulva rigida</i> (7.07%), the encrusting alga <i>Lithophyllum</i> 182 <i>incrustans</i> (6.62%), the leathery alga <i>Bifurcaria bifurcata</i> (6.17%) and the articulate183calcareus alga <i>Corallina officinalis</i> (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.</i> 186 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188 <i>S. muticum</i> was very abundant in control plots during the whole experiment,199a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4c). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p> 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p< 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).<	177	macroalgae assemblage structure (Fig. 3) were also detected by ANOSIM at both sites
179Table 2; Figure 3). Pair-wise comparisons detected differences in macroalgal180assemblages in most cases (Table 2).181The foliose alga <i>Ulva rigida</i> (7.07%), the encrusting alga <i>Lithophyllum</i> 182 <i>incrustans</i> (6.62%), the leathery alga <i>Bifurcaria bifurcata</i> (6.17%) and the articulate183calcareus alga <i>Corallina officinalis</i> (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.</i> 186 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188 <i>S. muticum</i> was very abundant in control plots during the whole experiment,190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	178	(differences among time factor: site 1, R=0.519, <i>p</i> <0.01; site 2, R=0.465, <i>p</i> <0.01;
180assemblages in most cases (Table 2).181The foliose alga Ulva rigida (7.07%), the encrusting alga Lithophyllum182incrustans (6.62%), the leathery alga Bifurcaria bifurcata (6.17%) and the articulate183calcareus alga Corallina officinalis (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, B, bifurcata (8.62%), U.186rigida (7.6%), L. incrustans (7.28%) and C. officinalis (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188S. muticum was very abundant in control plots during the whole experiment,189although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4190a). The following months after removal, Ulva rigida and Corallina officinalis191decrease of the former during autumn and winter was followed by an increase in192percentage cover of L. incrustans (illustrated by site 1 in Fig. 4 d). In contrast, B.194bifurcata195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for U. rigida (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	179	Table 2; Figure 3). Pair-wise comparisons detected differences in macroalgal
181The foliose alga Ulva rigida (7.07%), the encrusting alga Lithophyllum182incrustans (6.62%), the leathery alga Bifurcaria bifurcata (6.17%) and the articulate183calcareus alga Corallina officinalis (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, B, bifurcata (8.62%), U.186rigida (7.6%), L. incrustans (7.28%) and C. officinalis (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188S. muticum was very abundant in control plots during the whole experiment,189although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4190a). The following months after removal, Ulva rigida and Corallina officinalis191decrease of the former during autumn and winter was followed by an increase in192percentage cover of L. incrustans (illustrated by site 1 in Fig. 4 d). In contrast, B.194bifurcata was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for U. rigida (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	180	assemblages in most cases (Table 2).
182 <i>incrustans</i> (6.62%), the leathery alga <i>Bifurcaria bifurcata</i> (6.17%) and the articulate183calcareus alga <i>Corallina officinalis</i> (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.186<i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188<i>S. muticum</i> was very abundant in control plots during the whole experiment,189although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i>191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i>194<i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).</i>	181	The foliose alga Ulva rigida (7.07%), the encrusting alga Lithophyllum
183calcareus alga <i>Corallina officinalis</i> (4.84%) were the species that mostly contributed184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.</i> 186 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188 <i>S. muticum</i> was very abundant in control plots during the whole experiment,189although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	182	incrustans (6.62%), the leathery alga Bifurcaria bifurcata (6.17%) and the articulated
184to the dissimilarity between assemblages in both control and removal plots over tim185In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U.</i> 186 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the187highest contribution to the observed dissimilarity among assemblages.188 <i>S. muticum</i> was very abundant in control plots during the whole experiment,189although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	183	calcareus alga Corallina officinalis (4.84%) were the species that mostly contributed
In March 2008, 2 years after the start of the experiment, <i>B. bifurcata</i> (8.62%), <i>U. rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the highest contribution to the observed dissimilarity among assemblages. <i>S. muticum</i> was very abundant in control plots during the whole experiment, although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4 a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B. bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$; Estimate: 2.86, S.E: 1.06; Fig. 4 b).	184	to the dissimilarity between assemblages in both control and removal plots over time.
 <i>rigida</i> (7.6%), <i>L. incrustans</i> (7.28%) and <i>C. officinalis</i> (6.06%) still explained the highest contribution to the observed dissimilarity among assemblages. <i>S. muticum</i> was very abundant in control plots during the whole experiment, although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4 a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	185	In March 2008, 2 years after the start of the experiment, B. bifurcata (8.62%), U.
 highest contribution to the observed dissimilarity among assemblages. <i>S. muticum</i> was very abundant in control plots during the whole experiment, although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4 a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	186	rigida (7.6%), L. incrustans (7.28%) and C. officinalis (6.06%) still explained the
<i>S. muticum</i> was very abundant in control plots during the whole experiment, although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4 a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$; Estimate: 2.86, S.E: 1.06; Fig. 4 b).	187	highest contribution to the observed dissimilarity among assemblages.
although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4 a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$; Estimate: 2.86, S.E: 1.06; Fig. 4 b).	188	S. muticum was very abundant in control plots during the whole experiment,
190a). The following months after removal, <i>Ulva rigida</i> and <i>Corallina officinalis</i> 191dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The192decrease of the former during autumn and winter was followed by an increase in193percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> 194 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4 e). No195significant differences between treatments were detected in percentage cover of thes196native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$;197Estimate: 2.86, S.E: 1.06; Fig. 4 b).	189	although the percentage cover declined in late summer (illustrated by site 1 in Fig. 4
dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: $p > 0.05$) except for <i>U. rigida</i> (PA-GEEs: $p < 0.01$; Estimate: 2.86, S.E: 1.06; Fig. 4 b).	190	a). The following months after removal, Ulva rigida and Corallina officinalis
 decrease of the former during autumn and winter was followed by an increase in percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	191	dominated the primary substratum (illustrated by site 1 in Fig. 4 b, c; SIMPER). The
 percentage cover of <i>L. incrustans</i> (illustrated by site 1 in Fig. 4 d). In contrast, <i>B.</i> <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	192	decrease of the former during autumn and winter was followed by an increase in
 <i>bifurcata</i> was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	193	percentage cover of L. incrustans (illustrated by site 1 in Fig. 4 d). In contrast, B.
 significant differences between treatments were detected in percentage cover of thes native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	194	bifurcata was not abundant in the removal plots (illustrated by site 1 in Fig. 4e). No
 native species (PA-GEEs: <i>p</i>> 0.05) except for <i>U. rigida</i> (PA-GEEs: <i>p</i>< 0.01; Estimate: 2.86, S.E: 1.06; Fig. 4 b). 	195	significant differences between treatments were detected in percentage cover of these
197 Estimate: 2.86, S.E: 1.06; Fig. 4 b).	196	native species (PA-GEEs: p> 0.05) except for U. rigida (PA-GEEs: p< 0.01;
	197	Estimate: 2.86, S.E: 1.06; Fig. 4 b).
1984. Discussion	198	4. Discussion

199 Results indicated limited impact of *S. muticum* on native assemblages. The200 impact was evident on the total number of native taxa and two understory morpho-

201 functional groups, filamentous and foliose algae, rather than on the entire macroalgal 202 assemblages. In particular, the total number of taxa as well as the percentage cover of 203 filamentous and foliose algae was smaller in control than removal plots. In addition, 204 these effects were consistent at small spatial scale as predicted. 205 Studies of S. muticum have varied widely in their conclusions about its effect 206 on native assemblages. In general, studies done on intertidal shores have found little 207 (Buschbaum et al., 2006; Harries et al., 2007) or no impact of S. muticum (e.g. 208 DeWreede, 1983; Viejo, 1997; Sánchez and Fernández, 2005), but studies in subtidal 209 habitats have indicated relatively strong effects (Ambrose and Nelson, 1982; Stæhr et 210 al., 2000; Brittom-Simmons, 2004). Nevertheless, the impact of S. muticum on native 211 assemblages not only depends on the type of habitat, but also on the abundance of S. 212 muticum (Viejo, 1997, Britton-Simmons, 2004; Stæhr et al., 2000). According to this, 213 effect of *Sargassum* should be more evident in densely colonized areas, where this 214 species should be more likely to provoke changes in native macroalgal assemblages 215 through competitive interactions (Ambrose and Nelson, 1982; Viejo, 1997; Britton-216 Simmons, 2004). However, at low densities effects of invader on native macroalgal 217 assemblages might be very weak (see Viejo, 1997; Britton-Simmons, 2004). This 218 experiment was conducted at sites where the abundance of S. muticum was 219 comparatively low because the low intertidal corresponds to the upper end of its 220 distribution. Therefore, one may expect that the impact of S. muticum on native 221 assemblages is weaker because it is less abundant in this tidal zone. 222 Our results indicated a larger number of native taxa in those plots where 223 Sargassum was removed. Results from studies in marine benthic assemblages are 224 scarce and conflicting regarding relationships between diversity and potential for 225 invasion. For example, Dunstan and Johnson (2004) found positive relationships

226 between species richness and abundance of invaders using assemblages of sessile 227 invertebrates. Conversely, other experimental and observational studies have reported 228 negative relationships between the numbers of invasive and native taxa (Stachowicz et 229 al., 1999; 2002; White and Shurin, 2007). In fact, White and Shurin (2007) showed 230 that S. muticum was less abundant in areas with high diversity of native species. In our 231 study, the observed negative relationship may have different explanations. Firstly, 232 Sargassum may outcompete native species and therefore reduce native diversity, or 233 secondly, its environmental niche at low intertidal may coincide with areas of low 234 native diversity. The latter explanation, however, seems very unlikely because the low 235 intertidal area on the northwestern Iberian rocky shores is not a species-poor zone. In 236 fact, low intertidal assemblages on this coast are highly diverse and show important 237 spatial variation in species composition at scale of a few metres (see Araujo et al., 238 2005). Nevertheless, this experiment did not allow us to disentangle such possibilities. 239 Previous studies on S. muticum have indicated the important role of functional 240 groups in the invasion process (Viejo, 1997; Britton-Simmons, 2006). Recent findings 241 of Arenas et al. (2006) have pointed out that identity of functional groups is more 242 decisive than functional richness in determining the resistance of marine macroalgal 243 assemblages (Arenas et al., 2006). Functional groups play an important role in the 244 invasion process due largely to their effect on the primary resource requirements of 245 space and light (Arenas et al., 2006; Britton-Simmons, 2004; 2006). Alternatively, 246 different functional groups that present different survival strategies may be affected 247 differently by invasive species (see Viejo, 1997). In this study we found a negative 248 effect of S. muticum on the percentage cover of foliose and filamentous algae. These 249 results partially agree with those reported by Viejo (1997) that found an impact of 250 Sargassum on the leathery and foliose algae. Although several mechanisms

251	underlying the invasion process have been proposed to explain changes in native
252	macroalgal assemblages (Ambrose and Nelson, 1982; Critchley et al., 1990; Vadas et
253	al., 1992; Viejo, 1997; Britton-Simmons 2004) we only can speculate on the causative
254	mechanisms. For example, foliose algae, mainly composed of the opportunistic
255	species Ulva sp, might be inhibited by Sargassum by pre-emption of space (see Viejo,
256	1997). Foliose and filamentous algae might be also negatively influenced by
257	Sargassum due to competition for light. In fact, Britton-Simmons (2004) has
258	demonstrated experimentally the negative effect of Sargassum canopy on growth of
259	understory species and Arenas et al. (2006) have found that filamentous species such
260	as Ceramiales are mainly affected by light.
261	In summary, results suggest that there was limited effect of Sargassum on
262	macroalgal assemblages. It might be possible that the spatial (i.e. 50 x 50 cm plots)
263	and temporal (i.e. 2 years) scales used in this study were not appropriate to detect a
264	stronger impact. In fact, previous studies focused on effects of Sargassum on
265	macroalgal assemblages have used larger temporal scales, i.e. \geq 3 years (e.g. Viejo,
266	1997; Britton-Simmons, 2004, Sánchez and Fernández, 2005). The fact that two
267	morpho-functional groups were affected by Sargassum also highlights the importance
268	of functional groups in the invasion process, and the need to do manipulative
269	experiments in which functional groups are altered (see Arenas et al., 2006; Britton-
270	Simmons, 2006).
271	

272 Acknowledgments

This research has been supported by the Spanish Government through the Ministry of Education and Science (PROJECT CGL2005-02269). Funds to I.F. Rodil were provided in the course of a Maria Barbeito grant (Xunta de Galicia).

276 **References**

277	Ambrose, R.F., Nelson, B.V., 1982. Inhibition of giant kelp recruitment by an
278	introduced brown alga. Botanica Marina 25, 265-267.
279	Araujo, R., Bárbara, I., Sousa-Pinto, I., Quintino, V., 2005. Spatial variability of
280	intertidal rocky shore assemblages in the northwest coast of Portugal.
281	Estuarine, Coastal and Shelf Science 64, 658-670.
282	Arenas, F., Sánchez, I., Hawkins, S.J., Jenkins, S.R., 2006. The invasibility of marine
283	algal assemblages: role of functional diversity and identity. Ecology 87 (11),
284	2851-2861
285	Britton-Simmons, K.H., 2004. Direct and indirect effects of the introduced alga
286	Sargassum muticum on benthic, subtidal communities of Washington State,
287	USA. Marine Ecology Progress Series 277, 61-78.
288	Britton-Simmons, K.H., 2006. Functional group diversity, resource preemption and
289	the genesis of invasion resistance in a community of marine algae. Oikos 113,
290	395-401.
291	Buschbaum, C., Chapman, A.S., Saier, B., 2006. How an introduced seaweed can
292	affect epibiota diversity in different coastal systems. Marine Biology 148, 743-
293	754.
294	Chambers, P.A., Barko, J.W., Smith, C.S., 1993. Evaluation of invasions and declines
295	of submersed aquatic macrophytes. Journal of Aquatic Plant Management 31,
296	218-220.
297	Chapman, A.S., 1999. From introduced species to invader: what determines variation
298	in the success of Codium fragile ssp. tomentosoides (Chlorophyta) in the
299	North Atlantic Ocean?. Helgolander Meeresunters 52, 277-289.
300	Clarke, K.R., 1993. Nonparametric mutivariate analises of changes in community
301	structure. Australian Journal of Ecology 18, 117-143.
302	Crooks, J.A., 2002. Characterizing ecosystem-level consequences of biological
303	invasions: The role of ecosystem engineers. Oikos 97, 153-166.
304	Critchley, A.T., De Visscher, P.R.M., Nienhuis, P.H., 1990. Canopy characteristics of
305	the brown alga Sargassum muticum (Fucales, Phaeophyta) in Lake
306	Grevelingen, southwest Netherlands. Hydrobiologia 204/205, 211-217.

307	Dethier M.N. Graham F.S. Cohen S. Tear, I.M. 1993 Visual versus random-
308	point percent cover estimations: 'objective' is not always better. Marine Ecology
309	Progress Series 110 9–18
310	DeWreede R E 1983 Saraassum muticum (Eucales Phaeophyta): regrowth and
311	interaction with Phodomela larix (Ceramiales, Phodonhyta). Phycologia 22
312	153-160
313	Dunstan PK Johnson C R 2004 Invasion rates increase with species richness in a
314	marine epibenthic community by two mechanisms. Oecologia 138, 285-292.
315	Harries, D.B., Harrow, S., Wilson, J.R., Mair, J.M., Donnan, D.W., 2007. The
316	establishment of the invasive alga Sargassum muticum on the west coast of
317	Scotland: a preliminary assessment of community effects Journal of Marine
318	Biological Association of United Kingdom 87, 1057-1067.
319	Levin P.S. Cover I.A. Petrik R. Good T.P. 2002 Community-wide effects of
320	nonindigenous species on temperate rocky reefs. Ecology 83 (11), 3182-3193.
321	Liang, K.Y., Zeger, S.L., 1986. Longitudinal data analysis using generalized linear
322	models. Biometrika 73 (19), 13-22.
323	Mack, R.N., Simberloff, D., Lonsdale, W.M., Evans, H., Clout, M., Bazzaz, F.A.,
324	2000. Biotic invasions: causes, epidemiologoy, global consequences, and
325	control. Ecological Applications 10, 689-710.
326	Pérez-Cirera, J.L., Cremades, J., Bárbara, I., 1989. Precisiones sistemáticas y
327	sinecológicas sobre algunas algas nuevas para Galicia o para las costas
328	atlánticas de la Península Ibérica. Anales del Jardín Botánico de Madrid 46,
329	35-45.
330	Piazzi, L., Ceccherelli, G., Cinelli, F., 2001. Threat to macroalgal diversity: effects of
331	the introduced green alga Caulerpa racemosa in the Mediterranean. Marine
332	Ecology Progress Series 210, 140-159.
333	Rahel, F.J., 2000. Homogenization of fish faunas across the United States. Science
334	288, 854-856.
335	R Development Core Team, 2005. R: A language and environment for statistical
336	computing, reference index version 2.1.1. R Foundation for Statistical
337	Computing, Vienna, Austria. Available from: http://www.R-project.org
338	Ribera, M.A., Bouderesque, C.F., 1995. Introduced marine plants, with special
339	referente to macroalgae: mechanisms and impact, in: Round, F.E., Chapman,
340	D.J., Progress in Phycological Research, Biopress LTD Publ., pp. 187-268.

341	Ross, D.J., Johnson, C.R., Hewitt, C.L., Ruiz G.M., 2004. Interaction and impacts of
342	two introduced species on a soft-sediment marine assemblage in SE Tasmania.
343	Marine Biology 144, 747-756.
344	Sánchez, I., Fernández, C., 2005. Impact of the invasive seaweed Sargassum muticum
345	(Phaepohyta) on an intertidal macroalgal assemblage. Journal of Phycology
346	41, 923-930.
347	Stachowicz, J.J., Whitlatch, R.B., Osman, R.W., 1999. Species diversity and invasion
348	resistance in a marine ecosystem. Science 286, 1577-1579.
349	Stachowicz, J.J., Fried, H., Osman, R.W., Whitlatch, R.B., 2002. Biodiversity,
350	invasion resistance, and marine ecosystem function: reconciling pattern and
351	process. Ecology 83, 2575-2590.
352	Steneck, R.S., Dethier, M.N., 1994. A functional group approach to the structure of
353	algal-dominated communities. Oikos 69 (3), 476-497.
354	Strong, J.A., Dring, M.J., Maggs, C.A., 2006. Colonisation and modification of soft
355	substratum habitats by the invasive macroalga Sargassum muticum. Marine
356	Ecology Progress Series 321, 87-97.
357	Stæhr, P.A., Pedersen, M.F., Thomsen, M.S., Wernberg, T., Krause-Jensen, D., 2000.
358	The invasion of Sargassum muticum in Limfjorden (Denmark) and its possible
359	impact on the indigenous macroalgal community. Marine Ecology Progress
360	Series 207, 79-88.
361	Vadas R.L. Sr., Johnson, S., Norton, T.A., 1992. Recruitment and mortality of early
362	post-settlement stages of benthic algae. British Phycological Journal 27, 331-
363	351.
364	Verlaque, M., 1994. Checklist of introduced plants in the Mediterranean: origins and
365	impact on the environment and human activities. Oceanologica Acta 17, 1-23.
366	Viejo, R.M., 1997. The effects of colonization by Sargassum muticum on tidepool
367	macroalgal assemblages. Journal of Marine Biological Association of the
368	United Kingdom 77, 325-340.
369	Vilas, F., Bernabeu A, Méndez G., 2005. Sediment distribution pattern in the Rias
370	Baixas (NW Spain): main facies and hydrodynamic dependence. Journal of
371	Marine Systems. 54, 261-276.
372	Vitousek, P.M., Mooney, H.A., Lubchenco, J., Melillo, J.M., 1997. Human
373	domination of Earth's Ecosystems. Science 277, 494-499.

- 374 White, L.F., Shurin, J.B., 2007. Diversity effects on invasion vary with life history
- 375 stage in marine macroalgae. Oikos 116 (7), 1193-1203.
- 376 Williams, S.L., Smith, J.E., 2007. A global review of the distribution, taxonomy and
- 377 ecological impacts of introduced seaweeds. Annual Review of Ecology
- 378 ACCEPTED Evolution and Systematics 38, 327-359.
 - 379

	Number of		Diversity		Filamentous		Foliose		Corticated L		Leathery	Leathery algae		Calcareous		ng
	native taxa				algae		algae		algae		<i>y</i>		algae		algae	
	Е	SE	E	SE	Е	SE	Е	SE	Е	SE	Е	SE	E	SE	E	SE
Intercept	2.09***	0.04	1.16***	0.03	3.14***	0.38	17.15***	1.22	12.54***	1.31	7.50***	1.45	7.81***	0.99	7.79***	1.54
Treatment (T)	0.07*	0.04	-0.02	0.03	0.81*	0.38	3.06*	1.22	0.04	1.31	-2.17	1.45	0.27	0.99	2.98†	1.54
Site (S)	-0.02	0.04	0.05	0.03	-0.7	0.38	-3.49 **	1.22	-2.47	1.31	-3.33*	1.45	0.83	0.99	3.03*	1.54
T x S	0.05	0.04	0.00	0.03	0.04	0.38	-0.10	1.22	2.34	1.31	1.87	1.45	-1.37	0.99	-0.45	1.54
Correlation parameter	0.63	0.04	0.37	0.05	0.18	0.05	0.37	0.04	0.46	0.10	0.89	0.03	0.68	0.06	0.89	0.02
Scale parameter	0.79	0.09	0.08	0.01	47.3	13.68	310	48.8	129.1	22.8	79.8	26.8	58.9	8.97	79	21

Table 1. Results of Population-Averaged Generalized Estimating Equations examining the effects of Sargassum muticum on native algal assemblages

E = estimate; SE = standard error. p<0.05; p<0.01; p<0.01; p<0.001; p=0.056)

Correction

Years	2006		2007		2008	
Months	Site	Treatment	Site	Treatment	Site	Treatment
April	0.424**	-0.074	0.384**	0.226*	+	+
July	0.414**	0.074	0.48**	0.144	+	+
September	0.521**	0.044	0.232*	0.228*	+	+
December	0.356**	0.02	0.48**	0.134	+	+
February	+	+	0.476**	-0.074	0.214*	-0.022
ACCEPTERMANUSCAN						

Table 2. Results of the ANOSIM and pair-wise tests for differences on macroalgal assemblages between treatments (control and removal) and sites (1 and 2) over time (dates as in Figure 3)

Figure captions

Corre

Fig. 1. Percentage cover (mean \pm SE; n= 100) of *Sargassum muticum* in control and removal plots over the course of the experiment.

Fig. 2. Percentage cover (mean \pm SE; n= 5) of filamentous (a), foliose (b), encrusting algae(c), and number of native taxa (d) at site 1 and site 2.

Fig. 3. Non-metric multidimensional scaling (nMDS) for differences in assemblages showing temporal changes between treatments at site 1 and site 2 (n= 5). The first data (April 2006) for each site belongs to the pre-removal samples. Circle: site 1, triangle: site 2.

Fig. 4. Percentage cover (mean \pm SE; n=5) of Sargassum muticum (a), Ulva rigida (b), Corallina officinalis (c), Lithophyllum incrustans(d), and Bifurcaria bifurcata (e), over time at site 1. The first data point (April 2006) represents the pre-removal samples. S. muticum was excluded from the ANOSIM and SIMPER analyses.



Fig. 1 Olabarria et al.



ACERT



A





A Colored and a second and a se