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1	Differences in spatial and seasonal patterns of macrophyte assemblages between a
2	coastal lagoon and the open sea
3	
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15	
16	Abstract
17	
18	Although benthic macrophytes must be considered in monitoring programs to establish
19	the ecological status of transitional and coastal waters in the European Union, the
20	patterns of variability in species composition of macrophyte assemblages in
21	Mediterranean coastal lagoons has scarcely been studied. In this work the spatial (both
22	vertical and horizontal) and seasonal dynamics of macrophyte assemblages in a coastal
23	lagoon (Mar Menor) are compared with those of open coastal assemblages in the SW
24	Mediterranean to analyze any biological variability in lagoon assemblages and the
25	factors that determine such variability. Different assemblages, characterized by well
26	defined groups of species, can be described according to their isolation from the open
27	sea and the type of substratum; at the same time, a vertical zonation pattern, similar to
28	that found in all marine communities but more compressed, exists. This implies that
29	when applying the EU Water Framework Directive or assessing environmental impact,
30	a lagoon should not be considered spatially uniform and unique unit but as a mosaic of
31	assemblages.
32	
33	Keywords: macrophyte assemblages; coastal lagoons; spatio-temporal variability;
34	environmental impact assessment; Mar Menor
35	
36	1. Introduction
37	
38	Coastal lagoons have traditionally been considered as transitional systems between
39	continental and marine domains (Bianchi, 1988 a), a consideration that has gained in
40	importance in the context of the <i>Water Framework Directive (WFD)</i> of the European
41	Union. The WFD requires scientific and biological criteria to establish the basis for

42 typifying coastal ecosystems and transition waters (lagoons and estuaries). In the case of

43 the former, such criteria have long been established, while for transitional waters they

44 do not exist. The proposed criteria are not the same for both categories and, while there

45 is no doubt about the nature of estuaries, there is some controversy as to whether

46 lagoons should be treated as coastal or transitional waters.

47

Furthermore, in accordance with the WFD, the ecological status of a water body must be 48 evaluated using biological indicators, including macroalgae and seagrasses in the 49 context of hydromorphological and physico-chemical conditions (Ballesteros et al., 50 2007). In general, benthic macrophytes are considered good indicators of water quality 51 and different indices and methodologies based on them have been proposed for 52 assessing the ecological status of coastal and transitional waters in the context of the 53 WFD (Pasqualini et al., 2006; Austoni et al., 2007; Ballesteros et al., 2007; Pinedo et al., 54 2007; Wells et al., 2007). However, their application to transitional waters presents 55 difficulties as a consequence of changes in species composition along the natural 56 gradients of environmental conditions in these ecosystems (Wilkinson et al., 2007). 57 58 Some recent proposals look at the problem in estuarine transitional waters but explicitly exclude lagoons (Wilkinson et al., 2007). 59 60 61 Coastal lagoons undergo frequent physical and chemical disturbances and fluctuations (Unesco, 1981) and are consequently naturally stressed habitats (Barnes, 1980). They 62 63 are characterized by particular features, such as shallowness, relative isolation from the 64 open sea, usually as a result of coastal barriers that maintain some communication

65 channels or inlets, and the presence of boundaries with strong physical and ecological

66 gradients (Unesco, 1981). Due to their shallowness, bottoms are usually well irradiated,

67 while currents and hydrodynamics are closely conditioned by bottom topography and

68 wind affects the entire water column, promoting the resuspension of materials and

69 nutrients from the sediment surface layer. Because of these characteristics, coastal

lagoons are usually among the marine habitats that show the greatest biological
productivity (Alongi, 1998).

72

73 Pérès and Picard (1964) considered Mediterranean lagoon systems as a well 74 differentiated and unique homogeneous community, the so-called euryhaline and 75 eurythermal biocenoses. This consideration was maintained by later authors, such as Augier (1982) and Guelorget and Perthuisot (1983), and in the habitat lists of 76 conservation agreements, including OSPAR, Barcelona and Eunis. Although benthic 77 78 macrophytes must be considered in monitoring programs to establish the ecological 79 status of transitional and coastal waters (European Union, 2000), and despite a number 80 of papers that study species composition in transitional waters, the patterns of variability of macrophyte assemblages in coastal lagoons have scarcely been studied in the 81 82 Mediterranean. A few studies have explicitly characterized different communities or phytocoenoses according to the water characteristics in one or several lagoons (Lovric, 83 1979; Zouali, 1979; Chassany de Casabianca, 1980) or described vertical zonation 84 patterns (Cecere et al., 1988, Sfriso, 1987). However, some papers that analyze the 85

differences in species distribution or biomass related to environmental variables such as 86 salinity or trophic conditions (Sfriso et al., 2003; Curiel et al., 2004; Lin and Hung, 87 2004), do not consider the possibility of several communities existing in lagoons, 88 similar to those existing in coastal marine habitats, or, when referring to heterogeneity, 89 assume a single algal community for the whole lagoon (Cecere et al., 1992). In general, 90 and in accordance with the proposal of Péres and Picard (1964), benthic macrophytes 91 are considered to constitute a single assemblage within each lagoon (Chassany de 92 Casabianca, 1979; Nagy, 1979; Skolka and Tiganus, 1985; Cecere et al., 1991; Bachelet 93 et al., 2000; Mouillot et al., 2005). Because of this, such habitats are managed as units, 94 with decisions adopted at lagoon scale, whereas different areas of each lagoon might 95 require different management options (De Biasi et al., 2003). 96 97 98 Inter-lagoon variability has been attributed to many biotic and abiotic factors (Ross and 99 Epperly, 1985; Yañez-Arancibia et al., 1985) and, as regards the functioning of coastal lagoon ecosystems, several proposals have been made using different criteria to classify 100 and typify these environments. Among these proposals, some of the most important use 101 salinity as the main parameter (Anonymous, 1959; Petit, 1953; Por, 1972). Using 102 salinity as a descriptor involves considering autoecological criteria as a classification 103 104 basis, making the use of indicator species feasible (Bianchi 1988a, 1988b). 105 However, intra-lagoon variability also exists and has been related to depth (Sfriso, 106 107 1987; Pasqualini et al., 2006), type of substratum (Pérez-Ruzafa et al., 1989, 2006) and, 108 especially, confinement gradient sensu Guelorguet and Perthuisot (1983) an indicator of the degree of marine influence on lagoon ecosystems (Mariani, 2001). 109 110 Kjerfve (1994) also sub-divided coastal lagoons into three geomorphic types (choked, 111 112 restricted and leaky), which the author considered as three points along a spectrum of 113 water exchange with the coastal sea. The rate and magnitude of oceanic exchange 114 reflects both the dominant forcing functions and the time-scale of hydrological 115 variability. 116 The physical gradients within the lagoon environment arising from these exchange rates 117 have been related to biological gradients in species richness, abundance and 118 productivity. In the early 1980's, Guelorget and Perthuisot (Guelorget and Perthuisot, 119 1983; Guelorget et al., 1983) rejected salinity as an essential parameter for explaining 120

- 121 the observed gradients in density, biomass, species richness and diversity and proposed
- that zonation patterns and species distribution inside the lagoons were determined by
- the degree of confinement, a parameter which represents the turnover time of marine
- 124 water and the impoverishment in some oligo-elements of marine origin. Later, as
- regards the structure of faunal lagoon assemblages, Pérez-Ruzafa and Marcos (1992,
- 126 1993) suggested that, rather than vitamin and oligo-element recycling, the main factor
- 127 explaining them along a confinement gradient would be colonization rates by marine
- species. The species composition in each lagoon site will be the result of equilibria, in
- 129 the context of interspecific competition between marine and lagoon species.

130

131 According to this, lagoon assemblages would be the result of complex environmental

- 132 and biological interactions, and not only a response to the extreme physico-chemical
- 133 conditions or to a single horizontal gradient. The structure and composition of lagoon
- 134 communities would be better described by sinecological criteria and the same structural
- 135 parameters as in open coastal areas. In this way, multifactorial approaches and structural
- indices of lagoon assemblages could be used to describe the ecological status of a
- 137 particular lagoon site and community.
- 138

139 In this work, we study the spatial (both vertical and horizontal) and seasonal dynamics

of macrophyte assemblages in a coastal lagoon, compared with those of open coastal

assemblages in the SW Mediterranean, to analyze biological variability in lagoon

- assemblages, the factors that determine this variability, and the degree of differentiation
- between lagoon and open coastal macrophyte assemblages. The data used for this work
 pre-date the changes that have occurred in the trophic status of the Mar Menor waters in
- the last ten years (Pérez-Ruzafa et al., 2002) so that they are useful for the establishment
- 146 of reference conditions in this lagoon for the implementation of the WFD.
- 147

148 **2. Material and methods**

- 149
- 150 2.1. Study area

The Mar Menor is a hypersaline coastal lagoon, with a surface area of 135 km² located 151 on the southwestern Mediterranean coastline (37°42'00" North – 00°47'00" West) (Fig. 152 1) with a mean depth of 3.6 m and maximum of about 6 m. The salinity of the lagoon 153 154 waters range from 42 to 46. The bed sediment grain size composition is predominantly 155 muddy and sandy, with some areas of natural rocky bottoms around islands and some 156 calcareous and volcanic outcrops. Muddy bottoms, which cover both the whole central 157 area of the lagoon and the shallow zones showing lower hydrodynamism, are covered by a dense meadow of the algae *Caulerpa prolifera* (Forsskål) J.V. Lamouroux or 158 patches of the sea grass Ruppia cirrhosa (Petagna) Grande. Sandy bottoms (with sand 159 160 content up to 89%) are located at the margins of the basin and in the small bays surrounding the islands, in which sparse patches of the phanerogame Cymodocea 161 nodosa (Ucria) Asch. grow. 162

163

164 2.2. Sampling methods

To study the spatial and seasonal patterns of lagoon macrophyte assemblages, three
 localities (two inside the lagoon and another on the Mediterranean coast) were sampled
 monthly from June 1995 to October, 1996. Sampling sites were located where there was

no significant waste input from human activity based on our knowledge of the main

- sources of agricultural runoff inputs (Pérez-Ruzafa et al., 2000), the available
- information on the distribution of urban wastes, coliform bacteria and heavy metals in
- sediments and organisms (Marcos, 1991) and visual survey. The outer locality was
- 172 located at Cala Fría in the Cabo de Palos Marine Reserve, close to the Mar Menor
- 173 lagoon (Fig. 1), in a relatively wind-protected area to avoid excessive differences in

wave exposure compared with the lagoon localities. Of the two inner localities, one was
close to the main channel through which water exchange takes place, and the other one
was located in the southern basin, considered the most isolated and confined area of the
lagoon (Pérez-Ruzafa, 1989; Pérez-Ruzafa et al., 2004, 2005a, 2005b).

178

Macrophyte assemblages were sampled monthly by diving. All macrophyte species 179 were hand-collected in a square of 400 cm^2 (20 x 20 cm), a size considered 180 representative for midlittoral and infralittoral Mediterranean macrophyte communities 181 (Cinelli et al., 1977; Cecere et al., 1988; Ballesteros, 1992, 1993). Two replicates, 182 randomly distributed, were taken for each sample and the species were determined at 183 specific level. One of the samples was used for biomass estimations for this work and 184 the second was used for pigment extraction. Sampled open coastal assemblages include 185 186 midlittoral photophilous algae on rock (MPhR), infralittoral photophilous algae on rock (IPhR), infralittoral sciaphilous assemblage on rock (IScR) and Posidonia oceanica 187 meadow (PM). Inside the lagoon, samples were taken of MPhR a few centimetres under 188 189 the mean water level and at 1.5 m depth, both in well illuminated (IPhR) and in sciaphytic environments (IScR). Furthermore we sampled the Cymodocea nodosa 190 meadow (CyM) on sand and the *Caulerpa prolifera* meadow (CaM) on mud. 191 192 193 Macrophyte samples were kept in a cool-box and taken to the laboratory, where samples 194 were washed using clean sea water to remove sediments and then spread out in a tray 195 containing sea water to estimate the percentage of cover for each species according to 196 Cox (1981). After species level identification, the standing crop biomass for each 197 species was measured in terms of both dry weight (DW) and ash free dry weight 198 (AFDW). DW determination was made by drying to constant weight (24 h) in an oven at 110°C, and subsequently AFDW was determined by burning in a furnace at 450°C for 199 200 6 h. Weight was calculated with a precision of ± 0.0001 g. The classification and 201 nomenclature in this study follows that adopted by Guiry and Guiry (2007). 202 Key hydrographic (hydrodynamism, nutrient concentration in water column, pH, 203 204 temperature and salinity) and sediment characteristics were recorded monthly at the three stations during the study period. 205 206 207 The water column is well mixed and there were no significant differences in temperature or salinity with depth (Pérez-Ruzafa, 1989), so that water samples for 208 environmental variables were taken at an approximate depth of 1 m with a Niskin bottle, 209 or by pumping. Samples for nutrient analysis were kept in the dark at 4°C in the field 210 211 and stored at -28 °C. Nitrate (NO₃-N), nitrite (NO₂-N), ammonium (NH₄-N) and phosphate (PO_4 -P) were determined, following the methods described by Parsons et al. 212 (1984). To determine salinity a Beckman RS 7B salinometer was used, while pH 213 214 measurements were made with a pre-calibrated meter. 215

216 At each site, hydrodynamism (as represented by wave exposure) was estimated

217 according to Keddy (1983) as follows:

218	
219	Wave Exposure = \sum (mean wind velocity _{22.5°} * wind frequency _{22.5°} * effective fetch _{22.5°})
220	
221	where the effective fetch (F_e) is the direct fetch (F) , or distance in kilometres along
222	which the wind blows from each direction, corrected by fetches in directions of less
223	than 45° using the equation,
224	
225	$F_{e} = [F_{(\alpha)} * \cos \alpha + F_{(\alpha + 22.5)} * \cos (\alpha + 22.5) + F_{(\alpha - 22.5)} * \cos (\alpha - 22.5)] / [\cos \alpha + 2 *]$
226	$\cos(\alpha + 22.5)$]
227	
228 229	where α is equal to 0 for each F _e calculated.
230	Sediment samples were taken by diving in the communities on soft substrata, such as
231	sandy bottoms with Cymodocea nodosa or Posidonia oceanica meadows, and muddy
232	bottoms covered by Caulerpa prolifera meadows. Samples were hand taken from the
233	top 10 cm of the substrata using a shovel, stored and transported in darkness and cold in
234	polyurethane bags. At the laboratory they were dried at room temperature. During this
235	process they were stirred to break lumps before being sifted to a size of 2 mm. Grain
236	size distribution was determined in sub-samples by the Bouyoucos hydrometer method
237	(Soil Conservation Service, 1973) after dispersion of clusters by mechanical stirring in a
238	sodic-hexametaphosphate and Na ₂ CO ₃ solution. Previously, salts were eliminated by
239	washing and centrifugation and organic matter was eliminated by hydrogen peroxide
240	treatment. Grain size classification was made according to the International Association
241	for Soil Science (Duchafour, 1975). Organic carbon was determined by the Walkley-
242	Black method (Buchanan, 1984) and total nitrogen by the Kjeldahl method (Bremmer,
243	1965).
244	
245	2.3. Data analyses
246	Macrophyte assemblage structure was specified for each sample according to its species
247	richness and species composition, abundance (percentage of cover and biomass), and
248	Shannon-Wiener's diversity H' . For each species we estimated its relative dominance
249	(% D) in each assemblage and frequency of occurrence (f), classifying them into
250	frequent (F) ($f \ge 40\%$ corresponding to more than / months during the study period),
251	seasonal (S) $(18\% \le f < 40\%)$ and occasional (O) $(f < 18\%)$ species.
252	To study the officity between complex locand's coefficient (Devict al. 1071; Marcelef
255	10 study the arrinity between samples, Jaccard's coefficient (Day et al., 1971; Margaret
254	structure of the vegetation assemblages were assessed by non-metric multi-dimensional
255 256	scaling (MDS) and analyses of similarities (ANOSIM) using the statistical software
250 257	PRIMER 5.0 (Clarke and Warwick 2001) and similarity matrices based on the Bray
251	Curtis similarity coefficient generated with biomass (DW) data Values were square
250	root transformed before the analyses so that each species contributed fairly evenly to
260	each analysis (Clarke and Green, 1988) Pairwise ANOSIM comparisons were made
261	between all groups, using 10000 simulations in each case. Furthermore, site ordination
-51	

was also conducted using detrended correspondence analysis (DCA) on the records of 262 biomass (DW) of each species using the CANOCO 3.15 (ter Braak, 1997). 263 264 To explore the spatial variation of macrophyte assemblages in relation to environmental 265 characteristics, we performed a Canonic Detrended Correspondence Analysis (DCCA) (ter 266 Braak and Prentice 1988) using both square-root transformed cover and biomass (DW 267 and AFDW) data. This analysis was applied because the length of the gradient covered 268 by the first axis, when a Detrended Correspondence Analysis was performed on the 269 species matrix, was higher than 2 in units of standard deviation, and so the response of 270 the species to the environment gradient could not be considered lineal (Jongman et al., 271 1987). The ordination results of are displayed by scaling the axes and adjusting the 272 273 species' scores to the species' variance: the resulting scores are correlations between species and eigenvectors. The relative contribution of each variable to the ordination 274 established by the DCCA was evaluated by Monte Carlo permutation test after 275 performing a forward selection of variables at the 0.1% level of significance. All these 276 277 calculations were made using the CANOCO v. 3.15 package (ter Braak, 1990). 278 To quantify the spatial and temporal variation in macrophyte assemblages, data for 279 280 biomass (total, and per species), species richness, diversity and habitat descriptors were 281 analysed using 3-factor analysis of variance (Underwood, 1997), under the null hypothesis that each variable tested is homogeneous across the different localities, 282 283 assemblages and time. Monthly data were considered as replicates in each season. 284 All factors, community (c=2, photophilous midlittoral and infralittoral), season (s=5, 285 286 from summer 1995 to summer 1996, both included) and locality (1=3, Cabo de Palos, 287 Estacio and El Ciervo island) were fixed orthogonals. For the Caulerpa prolifera 288 meadow, two factor (season and locality) analysis of variance was performed. Prior to 289 analysis, homogeneity of variances was checked using Cochran's test. 290 When the results of the analyses were significant, Tukey's Honestly Significant difference 291 292 (HSD) tests were performed in order to identify the significantly different means. 293 3. Results 294

295

296 *3.1. Variation of environmental parameters*

297

The environmental conditions of the three localities differed (Fig. 2, Table 1). Although

water temperature and the suspended solids content were similar, they differed in

300 salinity, wave exposure and nutrient concentration (Table 2). Temperature showed a

301 marked seasonal pattern, with maximum temperatures in August (28 °C at El Estacio

and El Ciervo island, and 26°C at Cabo de Palos) and minimum in January and February

303 (13 °C at Cabo de Palos and 10 and 9 °C at El Estacio and El Ciervo island,

- 304 respectively). Water transparency was high, Secchi disc visibility reaching the bottom in
- all localities. Suspended solids ranged between 0.01 and 0.18 g/L, with no seasonal

pattern, and did not differ greatly between localities. The greatest difference in salinity 306 was between Cabo de Palos (39-40) and the two lagoon localities (44-47 at El Ciervo 307 island; 42-47 at El Estacio). There was also some time lag in the minimum salinity in 308 the three localities, Cabo de Palos reaching minimum levels in November, El Ciervo 309 island in December-January and El Estacio in February. 310 311 Wave exposure was higher in Cabo de Palos than in the lagoon localities, and the 312 interaction Locality * Season (p < 0.0001) was significant as a consequence of seasonal 313 changes in wind regime and because the three localities did not have the same 314 orientation. 315 316 317 In general, waters were oligotrophic in Cabo de Palos and had a slightly higher nutrient 318 concentration in the lagoon. Nitrate was the main nitrogen compound, ranging between 0.10 and 0.39 μ M in Cabo de Palos, 0.02 and 0.72 μ M in El Ciervo island and, 0.34 and 319 3.18 uM at El Estacio. Maximum values were recorded in summer in the Mar Menor 320 321 localities in the two sampled years and in summer (1995) and autumn (1996) in Cabo de Palos. Minimum values were reached in winter at all sites. The differences between 322 localities were significant (p < 0.0001), and the interaction Locality * Season (p < 0.05) 323 324 indicated that seasonal patterns differed among localities. 325 Phosphate showed highest values at El Ciervo island $(0.67 - 2.38 \,\mu\text{M})$, while the lowest 326 327 values were found in Cabo de Palos ($0.19 - 0.66 \,\mu$ M). El Estacio showed intermediate 328 values of 0.23-0.99 µM. Differences were significant for the factors Locality (p < 0.0001) and Season (p = 0.01) but not for the interaction, indicating that all localities 329 330 showed the same seasonal pattern. 331 332 3.2. Macrophytobenthic species composition at the sampling localities 333 Midlittoral and infralittoral communities on rocky, muddy and sandy substrata from the Mediterranean (Cabo de Palos), and the Mar Menor coastal lagoon (El Estacio and El 334 Ciervo Island) differed both in species composition and dominance. In total, 72 species 335 (23 Chlorophyta, 17 Ochrophyta, 29 Rhodophyta, 1 Cyanophyta and 2 Magnoliophyta) 336 were recorded (Tables 3, 4). The highest floristic richness was found in Cabo de Palos 337 (56 spp), while 25 species were found at El Ciervo island and 21 at El Estacio. A depth-338 related gradient was observed in the assemblage composition of the three sites. The 0 m 339 level at Cabo de Palos possessed the highest number of species (43), while the lowest 340 number (4) was observed in the *Caulerpa* beds from both El Estacio and El Ciervo 341 island. 26 species (37.68 %) were exclusive to the sea, while 13 (18.84%) (Anotrichium 342 343 tenue, Boergeseniella fruticulosa, Caulerpa prolifera, Chaetomorpha linum, *Cladophora albida, Cymodocea nodosa, Cystoseira foeniculacea* f. *tenuiramosa,* 344 Liebmannia leveillei, Polysiphonia subulata, Spyridia filamentosa, Ulva compressa, 345 *Ulva clathrata* and *Valonia aegagropila* were present in the Mar Menor samples only. 346 347 Among these, Boergeseniella fruticulosa, Chaetomorpha linum, Cymodocea nodosa, Cystoseira foeniculacea f. tenuiramosa and Spyridia filamentosa were exclusive to El 348

Ciervo Island and Anotrichium tenue, Polysiphonia subulata, Ulva compressa and Ulva
 clathrata were exclusive to El Estacio channel.

351

In terms of biomass, the *Posidonia* meadow showed the highest mean monthly value (143.80 g DW/400 cm²) while the minimum corresponded to the *Caulerpa prolifera* meadow (11.97 g DW/400 cm²) and the midlittoral photophilous assemblage on rock

 $(12.70 \text{ g DW}/400 \text{ cm}^2)$ at El Ciervo Island.

356

357 At Cabo de Palos the two shallower levels were dominated by *Stypocaulon scoparium*

and *Haliptilon virgatum*, the latter becoming less frequent with depth. Neither was
present in the lagoon. At 1.5 m depth, but only in shadow sites, *Peyssonnelia squamaria*was the dominant species. In the deepest water *Posidonia oceanica* was the dominant
seagrass.

362

363 At El Estacio, the 0 m level was dominated by *Cladophora* spp and *Ulva clathrata*,

364 while *Chondrophycus tenerrimus* dominated this level at El Ciervo island. At depth,

365 over hard substrata the community was dominated by *Chondrophycus tenerrimus*, and

366 Cystoseira foeniculacea f. tenuiramosa and Cystoseira compressa. Jania rubens was

the more important epiphyte algae. At El Ciervo island *Cymodocea nodosa* meadows

368 (mixed or not with the green alga *Caulerpa prolifera*, or in patchy outgrowths between

boulders) were dominant at all depths, while at El Estacio, a *Caulerpa prolifera* mono specific meadow was dominant at greater depths.

371

372 *3.3. Vegetation assemblage structure*

The hierarchical clustering of presence/absence data performed on a Jaccard matrix for mean seasonal biomass at each sampling site (assemblage) showed a clear separation between sea and lagoon assemblages, with the midlittoral samples from El Estacio and the 1995 summer sample from El Ciervo island close to them (Fig. 3).

377

At a second level, 10 well defined groups corresponded to the main midlittoral and 378 infralittoral assemblages both in the sea and lagoon environments. Group 1 included all 379 midlittoral Mediterranean samples; group 2 included spring and summer sciaphilous 380 communities on rock at Cabo de Palos; group 3 included Mediterranean infralittoral 381 photophilous samples on rock, except the winter sample, which joined the Posidonia 382 meadow samples, and autumn and winter sciaphilous communities on rock (group 4). 383 Groups 5 to 10 corresponded to lagoon assemblages: El Estacio midlittoral samples 384 385 (G5), Cymodocea nodosa meadows (G6), autumn and winter midlittoral samples at El Ciervo island (G7), infralittoral photophilous assemblages on rock at El Ciervo island, 386 387 including spring midlittoral assemblages at this locality (G8), El Estacio infralittoral photophilous assemblages (G9) and Caulerpa prolifera meadows at both El Estacio and 388 389 El Ciervo island (G10).

390

391 The MDS ordination plot based on biomass (DW) data confirmed these results and

392 clearly separated the samples into localities and assemblages (Fig. 4). This indicated

that the presence/absence of species, rather than their abundance, separates the different 393 groups, and confirms the existence of different assemblages. ANOSIM indicated that 394 these vegetation assemblages differed significantly. Global tests for differences between 395 locality groups showed significant differences between El Ciervo island and Cabo de 396 Palos (R-statistic=0.6, p<0.0001) and between El Estacio and Cabo de Palos (R-397 statistic=0.5, p < 0.0001), but no significant differences were found between El Estacio 398 and El Ciervo island. These differences became significant for all the pairwaise 399 comparisons between localities when only midlittoral and infralittoral photophilous 400 assemblages on rock (present in all the three localities) were considered (R-401 statistic=0.91, p<0.0001; R-statistic=0.996, p<0.0001; R-statistic=0.739, p<0.0001; 402 respectively). When comparisons were made between assemblages, the global test was 403 significant (Global R-statistic= 0.938, p < 0.0001). Pairwise tests were all significant at 404 405 p < 0.01 level (*R*-statistic ≥ 0.5) except between midlittoral and infralittoral photophilous assemblages, on rock at El Ciervo Island, which were significantly different at p < 0.05406 level (*R*-statistic=0.238). 407 408 The diversity values reached by midlittoral assemblages were the highest of all sampled 409 assemblages. Cumulative diversity was 3.26 bits indiv.⁻¹ at Cabo de Palos and 2.53 and 410 2.54 bits indiv.⁻¹ at El Estacio and El Ciervo island, respectively. 411 412 According to this, the different assemblages were characterized by well defined groups 413 414 of species at each locality. 415 416 Cabo de Palos 417 At Cabo de Palos, midlittoral photophilous assemblages (CP_MPhR) were dominated 418 by the frequent species Haliptilon virgatum and Stypocaulon scoparium, which 419 represented 29.5 and 25.5 %, respectively, of the total biomass. Dictyota fasciola, Jania 420 rubens var corniculata and Padina pavonica together constituted an additional 21.9 %. 421 Infralittoral Photophilous assemblages on rocky substrata (IPhR) were dominated by Cladostephus spongiosus (11.7%), Stypocaulon scoparium (60.9%) Haliptilon 422 423 virgatum (10.6%), Padina pavonica (6.0%) and Jania rubens var corniculata (2.8%), all of them frequent species. 424 Infralittoral sciaphilous assemblages on rocky substrata (IScR) was only present at Cabo 425 426 de Palos locality since in the Mar Menor, rocky substrata are hard volcanic crusts with only small holes and crevices (El Ciervo Island) or artificial boulders used to build 427 breakwaters (El Estacio) with too little space and illumination to permit conspicuous 428 429 algal development. The dominant species were Peyssonnelia squamaria (51.6 %, f=100%), Halopteris filicina (19.8%, f=100%), Haliptilon virgatum (3.8%, f=64.7%), 430

- 431 Flabellia petiolata (3.1 %, f=100%), Halimeda tuna (2.1 %, f=76.5%) and Amphiroa
- 432 rigida (1.58 %, f=41.2%). Seasonally or occasionally, Corallina elongata (8.2 %,
- 433 f=5.9%) and *Mesophyllum lichenoides* (2.23 %, f=23.5%) also contributed to the

434 assemblage biomass.

- 435 Posidonia oceanica meadows (PM) was also exclusive to the open coastal
- 436 Mediterranean bottoms. In terms of biomass the community was monospecifically

dominated by *Posidonia oceanica*, which contributed to 99.5% of the total biomass

- 438 (f=100 %). *Halopteris filicina* (0.3 %, f=41.2%) was the only other frequent species in
- the samples from this assemblage.
- 440
- 441 El Estacio
- 442 At El Estacio, midlittoral assemblages (EE_MPhR) were dominated by *Cladophora*
- 443 albida (37.2%), Jania rubens (25.9%) and Ulva clathrata (12.1%). However, except
- the first one, which was frequent, the others can be considered seasonal. *Cladophora*
- 445 *coelothrix* represented only 5.2 % of the total mean biomass but was present during
- 446 most of the year (*f*=52.9 %).
- 447 IPhR assemblages were dominated by a nearly mono-specific cover of Jania rubens
- 448 which represented 84.7 % (f=100 %) of the total assemblage biomass. Other frequent
- species were *Caulerpa prolifera* (6.8 %), *Valonia aegagropila* (2.5 %) and
- 450 Chondrophycus tenerrimus (0.5 %). Seasonally, Ceramium ciliatum (2.1% of the total
- 451 mean biomass), *Cladophoropsis modonensis* (1.6 %), *Padina pavonica* (0.8 %) and
- 452 Acetabularia acetabulum (0.28 %) made significant contributions to the assemblage
- 453 biomass.
- 454
- 455 El Ciervo Island
- 456 At El Ciervo Island, the dominant species in midlittoral assemblages (IC_MPhR) were
- 457 *Chondrophycus tenerrimus* (45.3 %) and *Cladophora albida* (23.5 %), both frequent,
- followed by *Cystoseira compressa* (8.6 %) and *Jania rubens* (6.7 %). *Cystoseira*
- 459 *compressa*, being a perennial species, overcomes the unfavourable season with basal
- 460 parts only and shows a pronounced seasonal cycle at this midlittoral level.
- 461 Infralittoral photophilous assemblages on rocky substrata were dominated by
- 462 Chondrophycus tenerrimus (58.2%), Cystoseira compressa (15.6%) and Cystoseira
- foeniculacea f. tenuiramosa (6.7%), all them with high frequencies of occurrence
- 464 (f=100 %). Also frequent but with a lower contribution to the total biomass were
- 465 Caulerpa prolifera (3.3 %, f=88.2%), Padina pavonica (4.0, f=94.1%) Cymodocea
- 466 nodosa (3.1%, f=76.5%) and Jania rubens (4.0%, f=64.7%).
- 467
- 468 *Cymodocea nodosa* (CyM) and *Caulerpa prolifera* (CaM) meadows were only sampled
- in the Mar Menor lagoon where the former constituted the most conspicuous
- community on sandy bottoms and the latter on muddy bottoms with a high organic
- 471 matter content. In terms of biomass, the former was monospecifically dominated by
- 472 Cymodocea nodosa, which contributed 98.9% to the total biomass (f=100 %). Any other
- 473 species, such as *Chaetomorpha linum*, *Acetabularia acetabulum* or *Acetabularia*
- *calyculus* were occasional and, in the last case, linked to the existence of small stones orshells.
- 476 On the other hand the *Caulerpa prolifera* meadow was dominated by this alga which
- 477 contributes 99.6 % and 97.2 % to the total biomass at El Estacio and El Ciervo island,
- 478 respectively (*f*=100 % in both cases). At El Ciervo island there were some seasonal
- 479 contributions from *Chaetomorpha linum*.
- 480

481 *3.4. Environmental factors explaining the assemblage composition and structure*

482 When performing the redundancy analyses (DCCA) on the macrophyte species matrix

using environmental data as explanatory variables, the cumulative percentage of

484 variance of the species-environment relation for the first two axes reached 59.1 % (Fig.

485 5). The first axis had an eigenvalue of 0.82 and explained 41.1 % of total variance.

486

The stepwise forward selection of environmental variables pointed to salinity, 487 temperature, organic matter, wave exposure, depth, phosphate, nitrate, and salinity and 488 temperature stress as having a significant influence on the assemblage structure (Fig. 6). 489 All the samples were distributed in the space defined by the positive parts of axes I and 490 II. The first axis represents a gradient from higher wave exposure and lower salinity on 491 the left, to little exposed assemblages and highly saline waters on the right. Other 492 descriptors related to natural environmental stress (range of salinity and temperature) 493 were also related to the positive part of the first axis. The second axis was related with 494 depth and nutrient content in the water column, with greater depth and phosphate in the 495 positive part and nitrate in the negative part. This axis explained only 18 % of the total 496 497 variance.

498

499 According to this, samples were distributed along a gradient from the Mediterranean

assemblages on the left to the lagoon assemblages on the right. In the lagoon part,

samples corresponding to El Estacio and to El Ciervo island were well differentiated.

502 Both in the Mediterranean and lagoon samples, two gradients corresponding to the two

axes were well defined within each locality, with deeper assemblages in the upper zone
and shallower (and environmentally more stressed) assemblages on the right (Fig. 5a).

505

506 Thus, the macrophytobenthic species correlated with the positive portion of axis 1 are

507 typical lagoon species such us *Cladophora albida*, *Anotrichium tenue*, *Valonia*

508 *aegagropila*, *Ulva clathrata* and *Liebmannia leveillei*. The negative part of axis 1 would

509 be determined by those species which do not inhabit the lagoon. The other species,

510 located in the centre of the plot, can be considered intermediate relative to their salinity

and temperature preferences. The second axis appears to be related to a gradient of

512 decreasing depth and increasing environmental stress. Its negative part was determined

513 by Ulva compressa, Ulva clathrata, Scytosiphon lomentaria, Liebmannia leveillei,

514 Polysiphonia subulata and Anotrichium tenue. The positive part was determined by the

515 phanerogams *Posidonia oceanica* and *Cymodocea nodosa*, in Cabo de Palos and El

516 Ciervo island, respectively, and the algae Cystoseira compressa, Cystoseira

517 foeniculacea f. tenuiramosa, Chondrophycus tenerrimus, Dictyota linearis, Taonia

518 *atomaria*, *Palmophyllum crassum* and *Codium bursa*, in both sites (Fig. 5b).

519

520

521 3.5. Spatio-temporal variation in macrophytobenthic assemblage structure

522 3.5.1. Changes in biomass

523 The seasonal variation in vegetation composition and structure of the assemblages and

524 in the biomass of individual species showed a different pattern, depending on the

assemblage and locality (Fig. 7 and 8). The two-way analysis of variance performed 525 considering the factors Season and Local_Assemblage found significant differences 526 between local assemblages but did not show significant differences in biomass between 527 seasons (Table 5). However, the three-way analysis performed for photophilous 528 assemblages on rock, which were the only ones present in the three localities and so 529 permitted a well balanced analysis, showed significant differences between Localities 530 (p<0.001), Assemblages (p<0.001) and the interaction Season*Locality (p<0.05) (Table 531 6) but, again, no differences were found between seasons either for the interaction 532 Season*Assemblage or Season*Locality*Assemblage. This suggests that the 533 assemblages show low seasonal changes in their total biomass and that these small 534 changes are dependent on differences in the environmental conditions in the three 535 localities that condition the biomass assemblage (Fig. 7 a-c). On the other hand, the 536 537 Caulerpa prolifera meadow showed significant differences in biomass between localities but they showed the same seasonal pattern (Table 7, Fig. 7d). 538 539 540 The behaviour of individual species was heterogeneous (Fig. 8). Of the 36 species 541 analysed, 15 showed seasonal changes at some significant level (p < 0.05) (Table 8). Cladostephus spongiosus, Cymodocea nodosa, Halopteris filicina and Peyssonnelia 542 squamaria showed significant differences in biomass between assemblages but not 543 544 significant seasonal variations (at p < 0.05). 545 546 Ceramium ciliatum and Ceramium tenerrimum (G. Martens) Okamura only showed 547 significant temporal patterns, but no differences were found between assemblages. Cystoseira compressa, Cystoseira foeniculacea f. tenuiramosa and Stypocaulon 548 549 scoparium showed significant differences in biomass between assemblages and between 550 seasons, but the temporal patterns were the same in all the communities and localities in 551 which they were present. 552 553 On the other hand, although Acetabularia acetabulum, Caulerpa prolifera, Chondrophycus tenerrimus, Dictyota fasciola, Dictyota linearis, Haliptilon virgatum, 554 555 Jania rubens, Jania rubens var corniculata, Padina pavonica and Ulva intestinalis also 556 showed significant seasonal variations, the interaction Season*Local Assemblage was also significant, indicating that those patterns were different in each local assemblage. 557 558 559 Cladophora vagabunda, Cladophoropsis modonensis, Corallina elongata, Dasya corymbifera, Laurencia obtusa, Liagora viscida, Lyngbia sordida and Valonia 560 aegagropila showed no spatial or temporal variations at the studied scales. 561 562 563 3.5.2. Changes in community structure 564 The spatial and seasonal variability in species richness and diversity were higher than in 565 total biomass. Species richness and diversity showed strongly significant differences between local assemblages, with the Cabo de Palos midlittoral and the El Ciervo island 566

- infralittoral photophilous communities being the richest and most diverse and the
- 568 Caulerpa prolifera meadows, both at El Estacio and El Ciervo island, the least rich and

least diverse. However, seasonal variations showed different patterns for species 569 richness and for diversity. Species richness showed significant seasonal variations at 570 p < 0.05 level but the Tukey test identified no significant groups. In the case of diversity, 571 seasonal variations were highly significant (p < 0.001), with the highest diversity in 572 spring and summer and lowest in winter and autum. However, the fact that the 573 interaction Season* Local Assemblage was also significant (p < 0.001) indicates that 574 seasonal patterns in diversity were not the same at all the Local assemblages (Fig. 7i-l). 575 576 The ANOVA three-way analysis performed on photophilous assemblages on rock gave 577 similar results (Table 6). In the case of species richness, differences were significant for 578 Season (p < 0.005) and for Locality (p < 0.001), and for the interactions between 579 Locality*Assemblage (p < 0.001). In the case of diversity, differences were also 580 581 significant for Season (p<0.001) and for Locality (p<0.001), and the interactions between Locality*Assemblage (p<0.001) and Season* Locality* Assemblage 582 (p<0.005). In both cases, the highest values were reached at Cabo de Palos and the 583 lowest at El Estacio. 584 585 In the *Caulerpa prolifera* meadow (Table 7), differences were only significant for 586 diversity in the case of the factor Locality (IC>EE, p<0.05) and for the interaction 587 588 Season x Locality (p < 0.05). 589 4. Discussion and conclusions 590 591 Lagoon macrophyte benthic assemblages are diverse, heterogeneous and well 592 593 characterized according to the hydrological conditions pertaining to a particular depth 594 range or substrate or in given wave exposure conditions. This work contributes to 595 solving the long-standing discrepancies, whether coastal lagoons show similar vertical 596 zonation patterns to those observed in open coastal areas (as assumed by Mars, 1966; 597 Bianchi, 1988a, 1988b; Occhipinti et al., 1988) or whether, on the contrary, the lagoon environment constitutes a uniform system, controlled by extreme physico-chemical 598 599 conditions which disguise any small scale pattern. According to our results, the eurihaline and eurithermal biocenosis proposed by Pérès and Picard (1964) and Augier 600 (1982) to define coastal lagoon assemblages applies to just one particular assemblage 601 characteristic from lagoon soft (muddy) bottoms in more or less confined areas. 602 603 Our results showed that both marine and lagoon assemblages are clearly differentiated 604 as regards species composition, and that different lagoon assemblages are similarly well 605 differentiated according to vertical and horizontal zonation patterns. Inside the lagoon, 606 607 the macrophyte assemblages showed the same vertical zonation patterns and patch distribution (according the nature of the substrata and physico-chemical environmental 608 609 conditions) as the equivalent assemblages in open coastal areas. 610 However, the boundaries between vertical zonation levels were narrower in the inner 611 lagoon and some species that are characteristic from the midlittoral in open coastal 612

cons were common in the infralittoral part in the lagoon. While the midlittoral and

- 614 infralittoral photophilous assemblages on rock shared 34.0 % and 33.3% of the algae
- species at Cabo de Palos and El Estacio, respectively, at El Ciervo island these two
- assemblages shared up to 79.2 % of the species. This would explain the structural
- 617 similarities and the proximity in the ordination diagrams observed between midlittoral
- 618 photophilous assemblages on rock from Cabo de Palos and the infralittoral photophilous
- 619 assemblage in El Ciervo island.
- 620
- 621 Nineteen species (27.5 %) are shared by the lagoon and open coast marine assemblages
- studied in this work and most of the others characterize (Pérès and Picard, 1964;
- Augier, 1982) or are common in the different communities in the south-western
- 624 Mediterranean (Pérez-Ruzafa, I. and Honrubia, 1984; Pérez-Ruzafa et al., 1991; Flores-
- 625 Moya et al., 1995a, 1995b; Conde et al., 1996).
- 626
- The main differences observed between assemblages are due to a few exclusive species from distinct levels and environments and to differences in the dominance relationships. In the lagoon environment, especially in the most confined areas, the term "paralic" (accept Frigori et al. 1082; Cualoreat et al. 1082; Cualoreat and Parthuiset 1082, 1002)
- 630 (*sensu* Frisoni et al., 1983; Guelorget et al., 1983; Guelorget and Perthuisot, 1983, 1992)
- referring to typically lagoon species could still be useful.
- 632

Despite the confinement theory (Guelorget et al., 1983; Guelorget and Perthuisot, 1983, 633 634 1992), according to which there would be a decreasing gradient in species richness from 635 the inlets communicating with the open sea to the inner parts (more confined) of the lagoon, the assemblages at El Estacio showed the lowest number of species and lowest 636 637 diversity. In fact, El Estacio only shared 13 species with the open coastal assemblages. Pérez-Ruzafa (1989) and Pérez-Ruzafa and Marcos (1992, 1993), on the basis of 638 639 evidence from faunal assemblage gradients in the Mar Menor, reformulated the 640 confinement theory in different terms so that, instead of the recycling of "vitamins or 641 vital elements" and the influence of seawater, the main factor explaining the structure of lagoon assemblages in a confinement gradient would be the rates of colonization by 642 643 Mediterranean species. So, for the fauna, the species composition at each lagoon site will be the result of equilibria in the context of interspecific competition between sea 644 and lagoon species, taking into account that lower competition coefficients of 645 646 Mediterranean species in the lagoon environment or the partial or complete lack of reproduction inside the lagoon could be compensated by high immigration rates of 647 juveniles or adults from outer habitats. However, this mechanism does not seem to be 648 649 valid for vegetation. It could apply to some vegetal species – for example, some mats of *Posidonia oceanica* have occasionally been observed in the inlet-influenced areas of the 650 Mar Menor and the invasive algae *Caulerpa prolifera* were seen to progressively 651 colonize the lagoon from the less confined areas to the inner parts of the lagoon (Pérez-652 Ruzafa et al., 1989, 2005b) and is now colonizing rocky substrates (unpublished data). 653 However, according to the results of this work, this does not seem to be applicable to 654 most macrophyte species and the dispersion and colonization mechanisms of the 655 vegetation need to be analysed in greater depth. 656

657

El Estacio assemblages can be considered distinct assemblages (well differentiated and 658 characterized by some exclusive species). The site is not an ecotone between open 659 coastal and lagoon communities, but a lagoon location with environmental instability 660 tolerated only by some species. This would explain its lower species richness and 661 diversity. This instability could not be related to the maximum and minimum 662 temperature or salinity, or to the medium and long term variation in these parameters, 663 but with the short term (hours to days) scale determined by the changes in direction of 664 the current at the inlet (with a mean periodicity of 6 hours) (Arevalo, 1988) and the 665 alternating influence of Mediterranean and lagoon conditions. 666 667 The different assemblages described in this work could be considered as real biocenoses 668 669 sensu Margalef (1974), as communities that do not depend on organisms external to

- 670 themselves and in which the internal relationships are always stronger than the
- 671 interchanges with adjacent ones. In the case of both the vertical and horizontal zonation
- of macrophyte assemblages, the different communities can not be understood without
- the concepts of *continuum* and *nodum*, *sensu* Boudouresque (1970, 1971), and of
- 674 biocenosis (see Ballesteros, 1992 for a general review).
- 675
- 676 Many characteristic species showed significant differences in biomass for the factor
- 677 Local_assemblage and for the interaction Locality x Assemblage in the photophilous
- rocky assemblages. This would confirm the differences obtained in the ordination
- analyses. However, precisely because of the differences existing between the three
- 680 localities, a more complete bionomic study would need to replicate each one of the three
- 681 environmental situations.
- 682

683 As regards the total assemblages, biomass is higher in Cabo de Palos than in the lagoon and in the infralittoral assemblages than in the midlittoral. No significant differences 684 685 were found between the same assemblage in the two localities inside the lagoon except for the Caulerpa prolifera meadow, which showed higher biomass in El Estacio than in 686 687 El Ciervo island. De Biasi et al. (2003) also found that macroalgae were generally more 688 abundant in the outer part of the Orbetello lagoon but did not consider vertical zonation patterns or the influence of depth in their study. These authors rejected the theory of 689 690 "confinement" to describe spatial pattern in assemblages in every lagoon because of the spatial variability that they found over scales of tens to hundreds of metres. However, as 691 they did not consider the direction of currents and did not analyze any environmental 692 693 variable, such a rejection can not be supported.

694

695 Substrate type, nutrient concentration and the influence of sea vs fresh water are among

- the main factors affecting the species composition and abundance in macrophyte
- 697 assemblages in coastal lagoons (Pasqualini et al., 2006). According to our results, two
- additional factors seem to overlap in determining macrophyte biomass: open sea *vs*.
- lagoon environment, and infralittoral *vs.* midlittoral. Both could be defined as
- roo environmental stress gradients operating at the same time on two different spatial

scales: from hundreds of metres to kilometres in the horizontal plane, thus defining the 701 702 degree of isolation from the sea, and from centimetres to metres in the vertical zonation. In this way, infralittoral photophilous assemblages and *Posidonia* meadows at Cabo de 703 Palos showed the highest biomass followed by midlittoral photophilous assemblages in 704 the same locality, while the latter show the same biomass as the infralittoral 705 assemblages of the lagoon. Finally, midlittoral lagoon assemblages show the lowest 706 707 biomass. 708 This parallel effect of confinement and vertical zonation also occurs in species 709 composition so that, as stated above, some species that are characteristic of the 710 midlittoral in open coastal zones, are common in the infralittoral in the lagoon. There 711 are also structural similarities, as shown by the proximity in the ordination diagrams, 712 between midlittoral photophilous assemblages on rock from Cabo de Palos and the 713 714 infralittoral photophilous assemblage in El Ciervo island. 715 716 In addition, many species showed seasonal variations, depending on the locality. Seasonal changes in algal abundance are common in the Mediterranean where algal 717 abundance drops off sharply in summer (Coma et al., 2000). However, as with light and 718 719 nutrients, the main factors determining the production of algal communities, display 720 different temporal dynamics. The dynamics of algae from different groups and depths will be conditioned by the seasonal cycle of the main limiting factor affecting them. In 721 722 shallow assemblages that reach a production peak in spring, nutrients are usually the 723 main limiting factors, while light availability becomes the limiting factor at increasing 724 depth so that deeper assemblages reach their maximum production in summer 725 (Ballesteros, 1991). In the study area, light, water temperature and salinity showed similar seasonal patterns in the three localities, while nutrient concentration in the water 726 727 column showed different dynamics. Thus, the difficulty of finding significant seasonal 728 differences in biomass may be related to the fact that each factor acts in a different way on the biomass each season. Furthermore, many of the observed changes in vegetation 729 would be due to the natural seasonal life cycle of the species present. There are 730 731 perennial species that are present in a given season in the form of erect thalli and in other periods as basal parts or encrusting thalli (e.g. Cystoseira spp., Chondrophycus 732 tenerrimus) and there are annual species which have a life cycle lasting only a restricted 733 734 period of the year or species with several generations in a year (Ulva spp., Cladophora spp.) (Feldmann, 1951). 735 736 737 Maximum development of the assemblages on rock was observed in summer and autumn in Cabo de Palos but in spring at El Ciervo island, a lag that could be related 738

with the negative effect of extreme summer conditions in the inner lagoon. A spring

- maximum in macrophyte biomass has been observed in other Mediterranean lagoons
- 741 (De Biasi et al., 2003).
- 742

The pelagic primary productivity cycle shows different patterns in different parts in the

Mar Menor (Pérez-Ruzafa et al., 2005a). In the internal areas of the lagoon, including El

Ciervo island, the concentrations of chlorophyll *a* in the water column were also 745 maximal in spring and minimal in winter. At El Estacio, and in the central part of the 746 lagoon, under Mediterranean influence, the maximum was reached in summer. This 747 pattern seems to be similar to that observed in infralittoral photophilous assemblages. 748 749 Of particular note were the differences in biomass observed between the summer of 750 1995 and 1996, particularly in the Cabo de Palos assemblages, where nearly all the 751 dominant species decreased substantially during the second year. Only Haliptilon 752 virgatum, in the infralittoral photophilous community, showed the inverse pattern 753 754 Between 1995 and 1996 the total biomass dropped by one third in the midlittoral 755 photophilous community, by two thirds in the infralittoral photophilous assemblage and 756 757 by half in the sciaphilous assemblage. However, the *Posidonia oceanica* assemblage underwent a much smaller decrease. 758 759 Despite the interannual decrease in biomass observed in the open coast assemblages, the 760 lagoon assemblages at El Estacio and El Ciervo island did not show the same general 761 pattern. However, the behaviour of some species, such as *Cladophora albida* and 762 *Padina pavonica*, whose biomass decreased, or the opposite response shown by 763 764 Chondrophycus tenerrimus and Cystoseira compressa, whose biomass increased, suggests that the observed pattern was not a consequence of a local impact but of a 765 general process affecting the whole area. Such environmental stress would not affect the 766 767 species already adapted to extreme environments (Caulerpa prolifera, Chondrophycus tenerrimus or Cystoseira compressa at El Ciervo island) and would mainly have a 768 769 negative effect on the species in more stable environments. 770 771 These differences also suggest that supra-annual temporal scales of variability are 772 important in macrophyte lagoon assemblage dynamics. In this work, Caulerpa prolifera was present as a frequent and characteristic species on the infralittoral rocky bottoms of 773 both El Estacio and El Ciervo island. This species is characteristic of muddy substrata 774 and invaded the Mar Menor lagoon in the early 1970's after widening of El Estacio inlet 775 (Pérez-Ruzafa et al., 1989, 1991). The colonization of the infralittoral photophilous 776 777 assemblages on rock by this invasive algae, which is characteristic of muddy bottoms, constitutes a progressive process (Pérez-Ruzafa et al., 1991) which is endangering the 778 biological diversity of the Mar Menor hard substrata lagoon assemblages, in the same 779 way that it has already affected the communities on sandy bottoms (Pérez-Ruzafa et al., 780 781 2006). Accordingly, long term studies will be necessary to analyze the inter-annual 782 patterns of variation. 783 784 In conclusion, macrophytic benthic assemblages in the Mar Menor lagoon are not 785 homogeneous and it is possible to identify distinct assemblages with the same vertical

786 zonation that can be found in all marine communities, but characterized by well defined

groups of species, according to their isolation from the open sea and the type of

substratum. This implies that when applying the EU Water Framework Directive a 788

- lagoon should not be considered as a homogeneous unit but as a mosaic of assemblages. 789
- 790
- However, before the structure and composition of the assemblages described for the 791
- Mar Menor can be generalised and applied to other lagoon communities in the 792
- Mediterranean, more extensive studies of other lagoons are necessary. 793
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1027 Figure captions

1028 Figure 1. Geographical location of the Mar Menor lagoon and the Cabo de Palos marine 1029 protected area, and location of sampling stations. 1030 1031 Figure 2. Seasonal dynamic of salinity, temperature, nutrient concentration and 1032 suspended solids in the water column in the three localities studied. CP: Cape of Palos; 1033 1034 EE: El Estacio; IC: El Ciervo Island. 1035 Figure 3. Hierarchical clustering of presence/absence data performed on a Jaccard 1036 matrix for macrophyte mean seasonal biomass at each sampling site (assemblage) and 1037 locality. Sample codes (Locality Assemblage Season Year): CP Cabo de Palos: EE El 1038 Estacio; IC El Ciervo island_ MPh: midlittoral photophilous assemblages on rock; IPh: 1039 infralittoral photophilous assemblages on rock; ISc: infralittoral sciaphilous 1040 1041 assemblages on rock; PM: Posidonia oceanica meadow; CvM: Cvmodocea nodosa meadow; CaM: Caulerpa prolifera meadow_ Sm Summer; A Autumn; W Winter; Sp 1042 Spring_5 1995; 6 1996. See text for the description of clusters 1 to 10. 1043 1044 1045 Figure 4. MDS ordination plot based on macrophyte biomass (DW) data. 1046 Figure 5. Ordination of samples (a) and species (b) in the representations of the first 1047 axes of the redundancy analysis (DCCA) performed on the macrophyte species matrix 1048 using environmental data as explanatory variables. Key of abbreviations for samples 1049 and species are showed in Fig. 3 and Tables 3 and 4, respectively. 1050 1051 Figure 6. Environmental variables explaining the species and samples distribution in the 1052 space defined by the two first axes of the redundancy analysis (DCCA) performed on 1053 the macrophyte species matrix represented in Fig. 5. 1054 1055 Figure 7. Seasonal variation in biomass (a-d), species richness (e-h) and Shannon 1056 diversity (i-1) in the macrophyte assemblages of the Mar Menor lagoon and Cabo de 1057 Palos. Sample codes (locality assemblage) correspond to that Fig. 3. Error bars 1058

- 1059 correspond to mean standard error.
- 1060
- 1061 Figure 8. Seasonal variation in biomass of dominant macrophyte species at the different
- assemblages and localities studied. Sample codes (locality_assemblage) correspond to
- 1063 that Fig. 3. Error bars correspond to mean standard error.

1064 Figure 1





1066 Figure 2







Figure 4 1070



- IPh: infralittoral photophilous assemblages on rock
 O ISc: infralittoral sciaphilous assemblages on rock
 White symbols IC: El Ciervo island
 SoB: enring 1996
 SoB: enring 1996
- PM: Posidonia oceanica meadow
- CyM: Cymodocea nodosa meadow
- CaM: Caulerpa prolifera meadow

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Sp6: spring 1996

Sm6: summer 1996

1072 Figure 5





1076 Figure 7



Figure 8 1078



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Table 1. Mean values (± standard error of the mean, SEM) of environmental variables at the three localities studied. Salinity stress and Temperature stress are the difference between maximum and minimum values of these variables through the studied period.

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		Cabo de P	alos	El Ciervo	island	El Estacio	
Wave exposure $76518.58 \pm 8212.63 \ 1411.17 \pm 139.51 \ 3568.90 \pm 442.81$ water columnNitrite (μ M) $0.071 \pm 0.007 \ 0.100 \pm 0.008 \ 0.126 \pm 0.025$ Nitrate (μ M) $0.196 \pm 0.022 \ 0.214 \pm 0.056 \ 1.216 \pm 0.208$ Ammonia (μ M) $0.152 \pm 0.008 \ 0.217 \pm 0.015 \ 0.165 \pm 0.009$ Phosphate (μ M) $0.383 \pm 0.038 \ 1.055 \pm 0.129 \ 0.488 \pm 0.054$ Salinity $39.43 \pm 0.14 \ 45.90 \pm 0.31 \ 44.80 \ \pm 0.38$ Temperature (°C) $20.07 \pm 1.20 \ 21.23 \pm 1.66 \ 21.27 \ \pm 1.61$ pH $8.10 \ \pm 0.04 \ 8.22 \ \pm 0.06 \ 8.12 \ \pm 0.04$ Suspended solids (gL ⁻¹) $0.074 \ \pm 0.010 \ 0.087 \ \pm 0.007 \ 0.080 \ \pm 0.010$ Salinity stress $2 \ 3 \ 5$ Temperature stress (°C) $13 \ 19 \ 18$ sedimentOrganic carbon (%) $0.61 \ \pm 0.02 \ 1.49 \ \pm 0.24 \ 3.19 \ \pm 0.34$ Clay (%)(<2µ) $7.28 \ \pm 0.11 \ 10.32 \ \pm 1.04 \ 17.55 \ \pm 0.71$ Fine Silt (%)(20-50µ) $0.58 \ \pm 0.07 \ 1.00 \ \pm 0.30 \ 3.42 \ \pm 0.78$ Fine Sand (%)(50µ-0.2mm) $4.06 \ \pm 0.34 \ 17.84 \ \pm 1.70 \ 39.74 \ \pm 2.98 \ Coarse Sand (%)(0.2-2mm) \ 86.62 \ \pm 0.41 \ 68.31 \ \pm 3.49 \ 30.69 \ \pm 3.34$		mean	± SEM	mean	± SEM	mean	± SEM
water columnwater column0.071 ± 0.007 0.100 ± 0.008 0.126 ± 0.025 Nitrate (μ M)0.196 ± 0.022 0.214 ± 0.056 1.216 ± 0.208 Ammonia (μ M)0.152 ± 0.008 0.217 ± 0.015 0.165 ± 0.009 Phosphate (μ M)0.383 ± 0.038 1.055 ± 0.129 0.488 ± 0.054 Salinity39.43 ± 0.14 45.90 ± 0.31 44.80 ± 0.38 Temperature (°C)20.07 ± 1.20 21.23 ± 1.66 21.27 ± 1.61 pH8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹)0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress2355555Temperature stress (°C)13191818sedimentOrganic carbon (%)0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ)	Wave exposure	76518.58	±8212.63	1411.17	±139.51	3568.90	±442.81
water columnNitrite (μ M)0.071 \pm 0.0070.100 \pm 0.0080.126 \pm 0.025Nitrate (μ M)0.196 \pm 0.0220.214 \pm 0.0561.216 \pm 0.208Ammonia (μ M)0.152 \pm 0.0080.217 \pm 0.0150.165 \pm 0.009Phosphate (μ M)0.383 \pm 0.0381.055 \pm 0.1290.488 \pm 0.054Salinity39.43 \pm 0.1445.90 \pm 0.3144.80 \pm 0.38Temperature (°C)20.07 \pm 1.2021.23 \pm 1.6621.27 \pm 1.61pH8.10 \pm 0.048.22 \pm 0.068.12 \pm 0.04Suspended solids (gL ⁻¹)0.074 \pm 0.0100.087 \pm 0.0070.080 \pm 0.010Salinity stress2355Temperature stress (°C)131918sedimentOrganic carbon (%)0.61 \pm 0.021.49 \pm 0.243.19 \pm 0.34Clay (%)(<2µ)		,	0212.00	,	107.01	2000.20	
Ntrrte (μ M) 0.071 ±0.007 0.100 ±0.008 0.126 ±0.025 Nitrate (μ M) 0.196 ±0.022 0.214 ±0.056 1.216 ±0.208 Ammonia (μ M) 0.152 ±0.008 0.217 ±0.015 0.165 ±0.009 Phosphate (μ M) 0.383 ±0.038 1.055 ±0.129 0.488 ±0.054 Salinity 39.43 ±0.14 45.90 ±0.31 44.80 ±0.38 Temperature (°C) 20.07 ±1.20 21.23 ±1.66 21.27 ±1.61 pH 8.10 ±0.04 8.22 ±0.06 8.12 ±0.04 Suspended solids (gL ⁻¹) 0.074 ±0.010 0.087 ±0.007 0.080 ±0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ±0.02 1.49 ±0.24 3.19 ±0.34 Clay (%)(<2 μ) 7.28 ±0.11 10.32 ±1.04 17.55 ±0.71 Fine Silt (%)(2-20 μ) 1.46 ±0.18 2.52 ±0.76 8.59 ±1.96 Coarse Silt (%)(20-50 μ) 0.58 ±0.07 1.00 ±0.30 3.42 ±0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ±0.34 17.84 ±1.70 39.74 ±2.98 Coarse Sand (%)(0.2-2mm) 86.62 ±0.41 68.31 ±3.49 30.69 ±3.34	water column						
Ntrate (μ M) 0.196 ±0.022 0.214 ±0.056 1.216 ±0.208 Ammonia (μ M) 0.152 ±0.008 0.217 ±0.015 0.165 ±0.009 Phosphate (μ M) 0.383 ±0.038 1.055 ±0.129 0.488 ±0.054 Salinity 39.43 ±0.14 45.90 ±0.31 44.80 ±0.38 Temperature (°C) 20.07 ±1.20 21.23 ±1.66 21.27 ±1.61 pH 8.10 ±0.04 8.22 ±0.06 8.12 ±0.04 Suspended solids (gL ⁻¹) 0.074 ±0.010 0.087 ±0.007 0.080 ±0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ±0.02 1.49 ±0.24 3.19 ±0.34 Clay (%)(<2 μ) 7.28 ±0.11 10.32 ±1.04 17.55 ±0.71 Fine Silt (%)(2-20 μ) 1.46 ±0.18 2.52 ±0.76 8.59 ±1.96 Coarse Silt (%)(20-50 μ) 0.58 ±0.07 1.00 ±0.30 3.42 ±0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ±0.34 17.84 ±1.70 39.74 ±2.98 Coarse Sand (%)(0.2-2mm) 86.62 ±0.41 68.31 ±3.49 30.69 ±3.34	Nitrite (µM)	0.071	± 0.007	0.100	± 0.008	0.126	± 0.025
Ammonia (μ M) 0.152 ± 0.008 0.217 ± 0.015 0.165 ± 0.009 Phosphate (μ M) 0.383 ± 0.038 1.055 ± 0.129 0.488 ± 0.054 Salinity 39.43 ± 0.14 45.90 ± 0.31 44.80 ± 0.38 Temperature (°C) 20.07 ± 1.20 21.23 ± 1.66 21.27 ± 1.61 pH 8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2 μ) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Nitrate (μ M)	0.196	±0.022	0.214	± 0.056	1.216	± 0.208
Phosphate (μ M) 0.383 ± 0.038 1.055 ± 0.129 0.488 ± 0.054 Salinity 39.43 ± 0.14 45.90 ± 0.31 44.80 ± 0.38 Temperature (°C) 20.07 ± 1.20 21.23 ± 1.66 21.27 ± 1.61 pH 8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2 μ) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Ammonia (µM)	0.152	± 0.008	0.217	± 0.015	0.165	± 0.009
Salinity 39.43 ± 0.14 45.90 ± 0.31 44.80 ± 0.38 Temperature (°C) 20.07 ± 1.20 21.23 ± 1.66 21.27 ± 1.61 pH 8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2 μ) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Phosphate (μ M)	0.383	± 0.038	1.055	± 0.129	0.488	± 0.054
Temperature (°C) 20.07 ± 1.20 21.23 ± 1.66 21.27 ± 1.61 pH 8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress235Temperature stress (°C)131918sedimentOrganic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ)	Salinity	39.43	± 0.14	45.90	± 0.31	44.80	±0.38
pH 8.10 ± 0.04 8.22 ± 0.06 8.12 ± 0.04 Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress235Temperature stress (°C)131918sedimentOrganic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ)	Temperature (°C)	20.07	± 1.20	21.23	±1.66	21.27	±1.61
Suspended solids (gL ⁻¹) 0.074 ± 0.010 0.087 ± 0.007 0.080 ± 0.010 Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20µ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50µ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50µ-0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	pH	8.10	± 0.04	8.22	±0.06	8.12	± 0.04
Salinity stress 2 3 5 Temperature stress (°C) 13 19 18 sediment Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)($<2\mu$) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Suspended solids (gL^{-1})	0.074	±0.010	0.087	±0.007	0.080	±0.010
Temperature stress (°C)131918sediment10 ± 0.24 3.19 ± 0.34 Organic carbon (%)0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ)7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20µ)1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50µ)0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50µ-0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Salinity stress	2		3		5	
sedimentOrganic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(<2µ)	Temperature stress (°C)	13		19	$\langle \rangle$	18	
sedimentOrganic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)(< 2μ) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34				-			
Organic carbon (%) 0.61 ± 0.02 1.49 ± 0.24 3.19 ± 0.34 Clay (%)($<2\mu$) 7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)($2-20\mu$) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)($20-50\mu$) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50μ -0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)($0.2-2mm$) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	sediment	0.61	10.02	1 40	10.24	2 10	10.24
Clay (%)(<2 μ)7.28 ± 0.11 10.32 ± 1.04 17.55 ± 0.71 Fine Silt (%)(2-20 μ)1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50 μ)0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50 μ -0.2mm)4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm)86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Organic carbon (%)	0.61	± 0.02	1.49	± 0.24	3.19	± 0.34
Fine Silt (%)(2-20µ) 1.46 ± 0.18 2.52 ± 0.76 8.59 ± 1.96 Coarse Silt (%)(20-50µ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50µ-0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Clay (%)($< 2\mu$)	1.28	±0.11	10.32	±1.04	17.55	± 0.71
Coarse Silt (%)(20-50µ) 0.58 ± 0.07 1.00 ± 0.30 3.42 ± 0.78 Fine Sand (%)(50µ-0.2mm) 4.06 ± 0.34 17.84 ± 1.70 39.74 ± 2.98 Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Fine Silt (%)(2-20 μ)	1.46	±0.18	2.52	± 0.76	8.59	±1.96
Fine Sand (%)(50 μ -0.2mm) 4.06 \pm 0.34 17.84 \pm 1.70 39.74 \pm 2.98 Coarse Sand (%)(0.2-2mm) 86.62 \pm 0.41 68.31 \pm 3.49 30.69 \pm 3.34	Coarse Silt (%)($20-50\mu$)	0.58	±0.07	1.00	± 0.30	3.42	± 0.78
Coarse Sand (%)(0.2-2mm) 86.62 ± 0.41 68.31 ± 3.49 30.69 ± 3.34	Fine Sand (%)(50μ -0.2mm)	4.06	±0.34	17.84	± 1.70	39.74	± 2.98
	Coarse Sand $(\%)(0.2-2mm)$	86.62	± 0.41	68.31	± 3.49	30.69	± 3.34
	and the second se						

		Nitrite		Nitrate		Amonia		Phosfate		Salinity		Temperature	Particular (Suspended	solids	рН		Wave Exposure	
Source	df	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р	F-ratio	Р
LOCALITY	2	16.435	***	51.609	***	9.207	***	25.409	***	406.684	***	0.674	ns	0.92	ns	16.159	***	292.57	***
SEASON	6	3.317	*	4.292	**	2.771	*	3.468	**	12.504	***	22.171	***	0.77	ns	9.883	***	6.887	***
LOCALITY*SEASON	12	2.528	*	3.008	**	0.71	ns	1.171	ns	2.351	*	0.302	ns	0.502	ns	2.626	*	8.401	***
Error	30																		
							a ser and												

Table 2. Results of mixed analysis of variance performed on environmental variables showing the effects of the factors Locality, Season and their interaction (Locality x Season) indicating the factors at which significant variation exists (*** P < 0.001; ** P < 0.01; * P < 0.05).

and the

Table 3. Mean values of biomass (DW) per 400 cm², relative dominance (% D) and class (F) of frequency of occurrence of the macrophyte species sampled in this study at Cabo de Palos (CP). Abbreviations of species names used in Fig. 4b, total species richness (Sr), mean total biomass (B) and Shannon diversity (H', in bits*indiv⁻¹) at each assemblage, are also indicated (F frequent; S seasonal; O occasional). MPhR: Midlittoral Photophilous assemblage on Rock; IPhR: Infralittoral Photophilous assemblage on Rock; IScR: Infralittoral Sciaphilous assemblage on Rock; PM: *Posidonia oceanica* meadow.

		an										1	
		СР								7		P	
		MPhR			IPhR			IScR	diam.	чү ^г 1111	PM		
	Abbrev.	mean	%D	f		%D	f		%D	f		%D	f
Cvanonhyta										Å	1		
Lynghia sordida Gomont	Lsor	0.005	0.0	0					L -	and the second			
Chloronhyta	2501	0.000	0.0	0									
Acetabularia acetabulum (Linnoous) P.C. Silvo	1000	0.007	0.0	Ο	0.002	0.0	0	0.000	0.1	ç			
Anadyomana stallata (Wulfon) C. Agordh	Aato	0.007	0.0	ç	0.002	0.0	ě	0.005	0.1	0	0.002	0.0	0
Chasternership genera (Dillegen) Vätning	Aste	0.037	0.1	0	0.021	0.0	2	0.055	0.2	0	0.002	0.0	0
Chaledomorpha derea (Diliwyn) Kutzing	Caer	0.007	0.0	ů.			P						
Claaophora coelothrix Kutzing	Ccoe	0.644	2.1	5		de la compañía de la							
Cladophora pellucida (Hudson) Kutzing	Cpel	0.001	0.0	0		M	b.	0.000	0.0	~	0.004	0.0	~
Cladophora vagabunda (Linnaeus) Hoek	Cvag	0.007	0.0	Ö		illine '	₽	0.006	0.0	0	0.004	0.0	0
Cladophoropsis modonensis (Kützing) Reinbold	Cmod	0.039	0.1	0			_						
Codium bursa (Linnaeus) C. Agardh	Cbur			<u></u>	0.034	0.1	0						
Codium vermilara (Olivi) Delle Chiaje	Cver			1.4	0.449	1.1	S	0.288	1.7	S	0.028	0.0	0
Dasycladus vermicularis (Scopoli) Krasser	Dver	0.127	0.4	0	0.336	0.8	0						
Flabellia petiolata (Turra) Nizamuddin	Fpet	0.002	0.0	0	0.017	0.0	0	0.520	3.1	F	0.030	0.0	S
Halimeda tuna (J. Ellis & Solander) J.V. Lamouroux	Htun	0.238	0.8	F	0.081	0.2	S	0.360	2.1	F			
Palmophyllum crassum (Naccari) Rabenhorst	Pcra							0.024	0.1	0			
Ulva intestinalis Linnaeus	Uint	1.310	4.2	F									
<i>Ulva rigida</i> C. Agardh	Urig 🥖	0.018	0.1	0									
Valonia utricularis (Roth) C. Agardh	Vutr 🥌	0.052	0.2	F				0.042	0.3	0			
Magnolionhyta													
Posidonia oceánica (L.) Delile	Poce	$\mathbf{\Psi}$									143.0	99.5	F
Ochronhyta		÷											-
Cladostenhus spongiosus (Hudson) C Agardh	Cspo	0.131	04	S	4 835	117	F	0.131	0.8	F	0.055	0.0	0
Colpomenia sinuosa (Mertens ex Roth) Derbès & Solier	Csin	0.047	0.1	ŏ	0.122	03	$\hat{0}$	0.011	0.0	ò	0.000	0.0	0
Custosaira brachucarna var balaarica (Sauvageau) Giaccone	Chra	1 1 1 0	3.6	Ē	0.038	0.1	ŏ	0.011	0.1	0			
Cystoseira compressa (Esper) Gerloff & Nizemuddin	Com	0.301	1.2	S	0.050	0.1	0						
Districts farsciola (Poth) IV Lamouroux	Dfac	1 781	57	E				0.012	0.1	0			
Dictional Jusciola (Rolli) J. V. Lamouroux	Dias	1./01	5.7	I.	0.620	15	ç	0.012	0.1	E	0.007	0.0	0
Halentavia filiaina (Crateloun) Kützina	Dilli Lifi				0.020	1.5	3	2 2 4 2	10.9	Г	0.007	0.0	E
Dadius a sussian (Linnocuo) Thinu	ПШ Dmari	2616	07	Б	2 475	60	Б	0.206	19.0	Г С	0.3/1	0.5	Г
Paalha pavonica (Linnaeus) Thivy	Ppav	2.010	8.3	г	2.4/3	0.0	Г	0.296	1.8	3	0.027	0.0	0
Sargassum vulgare C. Agardh	Svui	0.071	0.0	0	0.093	0.2	0						
Scytosiphon lomentaria (Lyngbye) Link	Slom	0.0/1	0.2	0									
Sphacelaria cirrosa (Roth) C. Agardh	Scir	0.009	0.0	S			Б						
Stypocaulon scoparium (Linnaeus) Kützing	Ssco	8.022	25.5	F	25.27	60.9	F						
Taonia atomaria (Woodward) J. Agardh	Tato				0.007	0.0	0						
Zonaria tournefortii (J.V. Lamouroux) Montagne	Ztou										0.114	0.1	0
Rhodophyta													
Alsidium corallinum C. Agardh	Acor	0.089	0.3	0	0.002	0.0	0						
Amphiroa rigida J.V. Lamouroux	Arig	0.105	0.3	S	0.405	1.0	S	0.267	1.6	F			

Ceramium ciliatum (J. Ellis) Ducluzeau	Ceil	0.351	1.1	0									
Ceramium dianhanum (Lightfoot) Roth	Cdia	0.016	0.1	ŏ								đ	•
Ceramium tenerrimum (G. Martens) Okamura	Cten	0.064	0.2	ŏ								A	Mar.
Champia parvula (C. Agardh) Harvey	Cnar	0.003	0.0	õ							4	6/	er m
Chondria canillaris (Hudson) M J. Wynne	Ccan	0 141	04	Ĕ				0.011	0.1	0		- M	
Chondrophycus tenerrimus (Cremades) G Furnari Boisset Cormaci et Serio	Chten	0.555	1.8	F				0.011	0.1	0	and the second	_ ₹	▶
Coralling elongata I Ellis & Solander	Celo	0 141	0.4	ò	0 177	04	0	1 391	82	0	1		
Dasva corymbifera I Agardh	Dcor	0.111	0.1	0	0.010	0.0	ŏ	0.032	0.2	ŏ		1	
Digenea simplex (Wulfen) C. Agardh	Dsim	0 188	0.6	0	0.010	0.0	0	0.052	0.2	÷.		<i>y</i>	
Falkenbergia rufolanosa Stadium	Fruf	0.009	0.0	ŏ					d.	A	0.004	0.0	0
Gelidium nusillum (Stackhouse) Le Iolis	Gnus	0.037	0.0	ŏ						Γ.	0.001	0.0	0
Gelidium spinosum (S.G. Gmelin) P.C. Silva	Gspi	0.047	0.2	ŏ				0.009	01	0	7		
Gratelounia filicina (IV Lamouroux) C. Agardh	Gfil	0.005	0.0	ŏ				0.005	0	U			
Halintilon virgatum (Zanardini) Garbary & H.W. Johansen	Hvir	9 2 5 4	29.4	Ĕ	4 384	10.6	F	0.638	3.8	F	0.027	0.0	0
Hypnea musciformis (Wulfen) IV Lamouroux	Hmus	0.065	0.2	ŝ		1010	<u>.</u>	0.050	0.0	•	0.027	0.0	0
Jania Rubens (Linnaeus) J.V. Lamouroux	Jrub	0.617	2.0	F	0.292	0.7	F	0.043	0.3	F	0.002	0.0	0
Jania rubens var corniculata (Linnaeus) Yendo	Icor	2.468	7.9	F	1 169	2.8	F	0.170	1.0	F	0.007	0.0	ŏ
Laurencia obtusa (Hudson) IV Lamouroux	Lobt	200		•	0.001	0.0	0	0.020	0.1	ŝ	0.007	0.0	0
Liagora viscida (Forsskål) C. Agardh	Lvis	0 1 7 4	0.6	0	0.625	1.5	Š	0.020	0.1	0			
Mesonhyllum lichenoides (L Ellis) M. Lemoine	Mlic	0.171	0.0	0	0.025	1.5		0 378	22	S			
Peyssonnelia sayamaria (S.G. Gmelin) Decaisne	Psou					hh.		8 716	51.6	F	0 103	0.1	S
Tricleocarna fragilis(Linnaeus) Huisman & Townsend	Tfra	0415	13	S	, W			0.002	0.0	Ô	0.105	0.1	5
Thereocul pu fluging (Emmadus) Thirsman & Townsend	1 II u	0.115	1.5	N.	▶ ₽			0.002	0.0	0			
	Sr	43	din.	- N	24			26			15		
	B	31 42		m)	41 47			16 90			143.8		
	H'(loga)	3 26		,	2.08			2 45			0.06		
	$H'(log_2)$	3.26	the second secon	Þ	2.08			2.45			0.06		

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Table

Table 4. Mean values of biomass (DW) per 400 cm², relative dominance (% D) and class (F) of frequency of occurrence of the macrophyte species sampled in this study at the Mar Menor localities. Abbreviations of species names used in Fig. 4b, total species richness (Sr), mean total biomass (B) and Shannon diversity (H', in bits*indiv⁻¹) at each assemblage, are also indicated (F frequent; S seasonal; O occasional). EE: El Estacio; IC: El Ciervo Island. MPhR: Midlittoral Photophilous assemblage on Rock; IPhR: Infralittoral Photophilous assemblage on Rock; CaM: *Caulerpa prolifera* meadow; CyM: *Cymodocea nodosa* meadow. + indicate species observed in the assemblage but not collected in the sample.

		FF									10											
		EE			IDI D			C-M			IC MDL D			IDL D			C-M			C-M		
	A h	MPhK	0/ D	c	IPNK	0/ D	£	CaM	0/D	-	WPNK	0/D	c	IPhK	0/ D	c	Сум	0/ D	£	CalM	0/D	£
Currentente	Ablev	mean	%D	1	mean	%D	1	mean	%D	1	mean	%0D	1	mean	%D	1	mean	%D	1	mean	%0D	1
Cyanophyta Lunchis condition Compart	Laam	0.004	0.0	c					. 💓	pr-	0.217	17	0	0.215	0.0	0	0.006	0.0	0			
Chlevenbute	LSOI	0.994	9.0	3				4			0.217	1./	0	0.215	0.8	0	0.000	0.0	0			
Chiorophyta	A	0.062	0.6	0	0.057	0.24	C	0.001	0.0	0	0.029	0.2	0	0.000	0.4	Б	0.006	0.0	0	0.001	0.0	0
Acetabularia acetabulum (Linnaeus) P.C. Silva	Aace	0.062	0.0	0	0.037	0.5	3	0.001	0.0	0	0.038	0.5	0	0.099	0.4	г	0.000	0.0	0	0.001	0.0	0
Caulouna nuclifana (Eoroskål) IV. Lomourouv	Coro				1 2 7 9	60	Е	22.22	00.6	Б	0 000	07	ç	0.850	2.2	Б	0.022	0.2	0	11.62	07.2	Б
Chartomorpha garag (Dillwarn) Kützing	Coor	<u>т</u>			1.370	0.0	г	LL.LL	99.0	г	0.088	0.7	3	0.839	5.5	г	0.033	0.2	0	11.05	91.2	Г
Chaetomorpha linum (O.F. Müller) Kützing	Clin	1									0.004	0.0	0	0.246	0.0	F	0.006	0.0	0	0 3 1 5	26	S
Cladophora albida (Nees) Kutzing	Calb	3 860	37.2	Ē		- alle					2 984	23.5	F	0.035	0.7	0	0.000	0.0	0	0.515	2.0	5
Cladophora coelothrir Kützing	Ccoe	0.543	52	F							0.093	0.7	0	0.055	0.1	s						
Cladophora vagabunda (Linnaeus) Hoek	Cvag	0.545	5.4	1.4	0.008	0.0	0				0.022	0.7	0	0.005	0.0	0						
Cladophorapsis modonensis (Kützing) Reinhold	Cmod				0.319	1.6	ŏ				0.009	0.1	0	0.002	0.0	Ŭ						
Ulva clathrata (Roth) C. Agardh	Ucla	1 257	12.1	S	0.017	1.0	0				0.007	0.1	Ŭ									
Ulva compressa Linnaeus	Ucom	0.285	2.7	S																		
Ulva intestinalis Linnaeus	Uint										0.040	0.3	0	0.016	0.1	0						
Ulva rigida C. Agardh	Urig	J																				
Valonia aegagropila C. Agardh	Vaeg	Dara and			0.510	2.5	F							0.011	0.0	0						
Valonia utricularis (Roth) C. Agardh	Vutr																					
Magnoliophyta	A CONTRACT																					
Cymodocea nodosa (Ucria) Asch.	Cnod										0.321	2.5	S	0.811	3.1	F	14.60	98.9	F	0.024	0.2	0
Ochrophyta																						
Cystoseira compressa (Esper) Gerloff & Nizamuddin	Ccom										1.097	8.6	S	4.095	15.6	F	0.063	0.4	0			
Cystoseira foeniculacea f. tenuiramosa (Ercegovic) A. Gómez Garreta, M.C.																						
Barceló, M.A. Ribera & J. Rull Lluch	Cfoe										0.466	3.7	S	1.755	6.7	F	0.016	0.1	0			
Dictyota linearis (C. Agardh) Greville	Dlin			_							0.054	0.4	0	0.063	0.2	S						
Liebmannia leveillei J. Agardh	Llev	0.022	0.2	0			~						~	+								
Padina pavonica (Linnaeus) Thivy	Ppav	0.100	1.0	~	0.166	0.8	S				0.201	1.6	0	1.048	4.0	F						
Scytosiphon lomentaria (Lyngbye) Link	Slom	0.132	1.3	0								+										
Sphacelaria tribuloides Meneghini	Stri				+										+							
Alsidium corallinum C. Agardh	Acor	0.007	0.1	0	0.014	0.1	0							+			+					
Anotrichium tenue (C. Agardin) Nagen	Aten	0.007	0.1	0	0.014	0.1	0							0.021	0.1	0						
<i>Boergeseniella fruticulosa</i> (Wulfen) Kylin	Biru	0.276	26	0	0 425	2.1	0				0.271	2.1	0	0.031	0.1	0						
Ceramium citatum (J. Ellis) Duciuzeau	Cdia	0.370	3.0 0.2	0	0.433	2.1	0				0.271	2.1	0	0.003	0.2	0						
Ceramium auaphanum (Light1001) Koth	Ctan	0.022	0.2	0	0.008	0.0	0				0.005	0.0	0	0.025	0.1	0						
Ceramium ienerrimum (G. Mattens) Okamura	Cten	0.082	0.8	0	0.110	0.5	0				0.005	0.0	U	0.025	0.1	0						

Champia parvula (C. Agardh) Harvey Chondria capillaris (Hudson) M.J. Wynne Chondrophycus tenerrimus (Cremades) G. Furnari, Boisset, Cormaci et Serio Dasya corymbifera J. Agardh Falkenbergia rufolanosa Stadium Herposiphonia secunda (C. Agardh) Ambronn	Cpar Ccap Chten Dcor Fruf Hsec	0.051	0.5	0	0.093 0.007 +	0.5 0.0	F O	0.056	0.3	0.0 0.0 O 5.7	005 048 749	0.0 0.4 45.3	O O F	0.178 15.31 0.013	0.7 58.2 0.1	F F O	0.004 0.014 0.006	0.0 0.1 0.0	0 0 0	
Hypnea musciformis (Wulfen) J.V. Lamouroux Jania Rubens (Linnaeus) J.V. Lamouroux Polysiphonia subulata (Ducluzeau) P.L. Crouan & H.M. Crouan Spyridia filamentosa (Wulfen) Harvey	Hmus Jrub Psub Sfil	2.690	25.9	S	17.20 0.001	84.7 0.0	F O	0.025	0.1	O 0.3	851 137	6.7 1.1	s o	+ 1.042 0.287	4.0 1.1	F S	0.012	0.1	0	
	Sr B H'	14 10.38 2.53			14 20.31 0.99		4	4 22.30 0.04		21 12 2.:	2.70 52			22 26.31 2.21			11 14.76 0.12			4 11.97 0.20
									7											
		4		5	- Ander															
																				
	And a																			

Table

Table 5. Results of mixed analysis of variance showing the effects of the factors season, local assemblage and their interaction (Season*Local_assemblage on the biomass, species richness and H' Shannon index of diversity of macrophyte assemblages at the three localities studied, indicating the factors at which significant variation exists. Results of Tukey (HSD) test are also shown indicating the homogeneous groups identified. See Tab. 3 and 4 for sample codes.

		Biomass			Species			H' diversity		
		n=165			n=165			n=165		
Source	df	F-ratio	р	Tukey (HSD)	F-ratio	р	Tukey (HSD)	F-ratio	р	-
SEASON	4	0.776	0.543	-	3.009	0.021	nd	8.295	0.0001	Spring_96=Summer_96=Summer_95>
LOCAL_ASSEMBLAGE	10	206.712	0.0001	CP_PM>CP_IPhR> CP_MPhR=IC_CyM= IC_IPhR= EE_CaM= EE_IPhR=CP_IScR> IC_MPhR= IC_CaM= EE_MPhR	34.289	0.0001	CP_MPhR= IC_IPhR> CP_IScR =CP_IPhR> IC_MPhR> CP_PM= EE_IPhR = IC_CyM= EE_MPhR > IC_CaM = EE_CaM	41.271	0.0001	Winter 96=Autumn 95 CP_MPhR=IC_IPhR> CP_IPhR=CP_IScR> IC_CyM > CP_PM =IC_MPhR > EE_MPhR > EE_IPhR > IC_CaM = EE_CaM
SEASON*	40	1.296	0.147		1.275	0.163		2.454	0.0001	
Error	110									
				A company of the second s						

Table 6. Results of mixed analysis of variance showing the effects of the factors season, locality, assemblage and their interaction (Season*Locality; Season* Assemblage; Locality* Assemblage and Season* Locality*Assemblage) on the biomass, species richness and H' Shannon index of diversity of the midlittoral and infralittoral macrophyte photophilous assemblages on rock, indicating the factors at which significant variation exists (bold figures). Results of Tukey (HSD) test for individual factors are also shown indicating the homogeneous groups identified. (nd: non differentiated groups). Sample codes as in Table 3 and 4.

		Biomass	s n =90		Species richness			н		
							4		/	
Source	df	F-ratio	р	Tukey (HSD)	F-ratio	р	Tukey (HSD)	F-ratio	р	Tukey (HSD)
SEASON	4	1.722	0.157		2.928	0.028	nd	8.369	0.0001	Spring_96=Summer_95>Winter_96=Autumn_95
LOCALITY	2	17.381	0.0001	CP>IC=EE	41.75	0.0001	CP>IC>EE	31.813	0.0001	CP=IC>EE
ASSEMBLAGE	1	15.392	0.0001	IPhR>MPhR	0.612	0.437		1.8	0.185	
SEASON*LOCALITY	8	2.402	0.026		1.035	0.42	N N	0.682	0.705	
SEASON*ASSEMBLAGE	4	1.218	0.313		0.479	0.751	- P	0.764	0.553	
LOCALITY*ASSEMBLAGE	2	0.635	0.534		24.399	0.0001		17.648	0.0001	
SEASON*LOCALITY*ASSEMBLAGE	8	0.987	0.455		1.786	0.098		3.629	0.002	
Error	60									
			A CONTRACT OF A							

Table

Table 7. Results of mixed analysis of variance showing the effects of the factors season, locality, assemblage and their interaction (Season*Locality; Season*Assemblage; Locality*Assemblage and Season*Locality*Assemblage) on the biomass, species richness and H' Shannon index of diversity in the *Caulerpa prolifera meadow*, indicating the factors at which significant variation exists .. Results of Tukey (HSD) test for individual factors are also shown indicating the homogeneous groups identified. (>= is used to indicate that only the extremes differ significantly).

		Biomass			Species richness	4		H' diversity		
Source	df	F-ratio	р	Tukey (HSD)	F-ratio	р	Tukey (HSD)	F-ratio	р	Tukey (HSD)
SEASON	4	4.039	0.015	Winter_96>=Spring_96=Autumn_95=Summer_95>=Summer_96	1.125	0.373		2.36	0.088	
LOCALITY	1	55.473	0.0001	EE>IC	0.333	0.57		7.819	0.011	IC>EE
SEASON*LOCALITY	4	2.69	0.061	÷	2.208	0.105		3.146	0.037	
				A Company of the second						

Table

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Table 8. Results of mixed analysis of variance showing the effects of the factors season, local_assemblage and their interaction (Season*Local_Assemblage) on the biomass, of frequent species, indicating *F*_va lues and significance levels. *p<0.05; **p<0.01; ***p<0.005; ****p<0.001; ns: not significant.

Species	Abrev.	SEASON	LOCAL_ASSEMBLAGE	SEASON*LOCAL_ASSEMBLAGE
Acetabularia acetabulum	Aace	6.365 ****	3.621 ***	3.302 ****
Caulerpa prolifera	Cpro	2.745 *	114.808 ****	2.933 *
Ceramium ciliatum	Ccil	2.863 *	0.274 ns	0.86 ns
Ceramium diaphanum	Cdia	1.202 ns	0.234 ns	0.949 ns
Ceramium tenerrimum	Cten	5.432 ***	0.345 ns	0.345 ns
Champia parvula	Cpar	0.79 ns	0.162 ns	1.21 ns
Chondria capillaris	Ccap	0.613 ns	4.402 ***	2.063 *
Chondrophycus tenerrimus	Chte	2.78 *	41.414 ****	1.271 ns
Cladophora albida	Calb	2.373 ns	4.977 *	2.935 *
Cladophora coelothrix	Ccoe	1.008 ns	1.066 ns	0.973 ns
Cladophora vagabunda	Cvag	0.794 ns	0.408 ns	1.041 ns
Cladophoropsis modonensis	Cmod	2.023 ns	1.197 ns	1.291 ns
Cladostephus spongiosus	Cspo	0.679 ns	15.886 ****	0.643 ns
Corallina elongata	Celo	0.878 ns	0.774 ns	1.057 ns
Cymodocea nodosa	Cnod	1.48 ns	119.785 ****	1.713 ns
Cystoseira compressa	Ccom	9.488 ****	21.087 ****	1.438 ns
Cystoseira foeniculacea f. tenuiramosa	Cfoe	7.77 ****	24.82 ****	1.804 ns
Dasya corymbifera	Dcor	1.144 ns	0.798 ns	0.9 ns
Dictyota fasciola	Dfas	3.496 *	3.779 ns	3.531 *
Dictyota linearis	Dlin	3.781 **	2.537 ns	2.28 *
Falkenbergia rufolanosa	Fruf	2.042 ns	0.605 ns	2.298 *
Haliptilon virgatum	Hvir	2.896 *	29.688 ****	2.896 **
Halopteris filicina	Hfil	0.593 ns	18.794 ****	0.272 ns
Jania rubens	Jrub	3.438 *	24.07 ****	2.241 ***
Jania rubens var corniculata	Jcor	2.896 *	29.688 ****	2.896 **
Laurencia obtusa	Lobt	0.996 ns	1.16 ns	1.099 ns
Liagora viscida	Lvis	1.926 ns	1.03 ns	0.628 ns
Lyngbia sordida	Lsor	1.162 ns	1.671 ns	1.318 ns
Padina pavonica	Ppav	5.77 ****	6.865 ****	2.112 **
Peyssonnelia squamaria	Psqu	0.142 ns	70.693 ****	0.119 ns
Posidonia oceanica	Poce	1.088 ns	-	-
Scytosiphon lomentaria	Slom	1.927 ns	-	-
Stypocaulon scoparium	Ssco	4.545 ***	26.023 ****	0.979 ns
Ulva clathrata	Ucla	1.762 ns	-	-
Ulva intestinalis	Uint	4.219 **	10.083 ****	4.511 ***
Valonia aegagropila	Vaeg	1.651 ns	0.182 ns	0.311 ns