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HAL Authorization

**Title:** N-terminal chimeras with signal sequences enhance the functional expression and alter the subcellular localization of heterologous membrane-bound inorganic pyrophosphatases in yeast

**Running Title:** Enhanced expression of H<sup>+</sup>-PPases in yeast

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*Abbreviations used:* ACMA, 9-amino-6-chloro-2-methoxy acridine; CCCP, cyanide *m*-chlorophenylhydrazone; DAPI, 4',6-diamidino-2-phenylindole; FM4-64, N-(3-triethylammoniumpropyl)-4-(*p*-diethylaminophenyl-hexatrienyl) pyridinium dibromide; GFP, green fluorescent protein; H<sup>+</sup>-PPase, proton-translocating inorganic pyrophosphatase; ORF, open reading frame; PP<sub>i</sub>, inorganic pyrophosphate; sPPase, soluble inorganic pyrophosphatase; PCR, polymerase chain reaction; yEGFP, yeast enhanced green fluorescent protein.

## Abstract

Expression of heterologous multispinning membrane proteins in *Saccharomyces cerevisiae* is a difficult task. Quite often, the use of multicopy plasmids where the foreign gene is under the control of a strong promoter does not guarantee efficient production of the corresponding protein. Here, we show that the expression level and/or subcellular localization in *S. cerevisiae* of an heterologous type of multispinning membrane protein, the H<sup>+</sup>-translocating inorganic pyrophosphatase (H<sup>+</sup>-PPase), can be changed by fusing it with various suitable N-terminal signal sequences. Chimeric proteins were constructed by adding the putative N-terminal extra domain of *Trypanosoma cruzi* H<sup>+</sup>-PPase or the *bona fide* signal sequence of *S. cerevisiae* invertase Suc2p to H<sup>+</sup>-PPase polypeptides of different organisms (from bacteria to plants) and expressed in a yeast conditional mutant deficient in its cytosolic P<sub>i</sub> hydrolysis activity when grown on glucose. Chimeric constructs not only substantially enhanced H<sup>+</sup>-PPase expression levels in transformed mutant cells but also allowed functional complementation in those cases in which native H<sup>+</sup>-PPase failed to accomplish it. Activity assays and Western blot analyses further demonstrated the occurrence of most H<sup>+</sup>-PPase in internal membrane fractions of these cells. The addition of N-terminal signal sequences to the vacuolar H<sup>+</sup>-PPase AVP1 from the plant *Arabidopsis thaliana* -a protein efficiently expressed in yeast in its natural form- alters the subcellular distribution of the chimeras suggesting further progression along the secretory sorting pathways, as shown by density gradient ultracentrifugation and *in vivo* fluorescence microscopy of the corresponding GFP-H<sup>+</sup>-PPase fusion proteins.

*Key words:* chimeric proteins, green fluorescent protein, heterologous expression, N-terminal signal sequences, proton-translocating inorganic pyrophosphatase, *Saccharomyces cerevisiae*.

## INTRODUCTION

Heterologous expression may pose important difficulties, especially in the case of multispanning membrane proteins, whose overexpression can be toxic to the host cell. The use of appropriate promoters of known properties allows the control of transcription; however, quite often, high levels of messenger RNA do not imply high amounts of the desired protein. This suggests that, besides transcription, other processes such as translation, protein targeting or degradation may also be crucial in order to obtain high levels of functional foreign proteins in a given system. Eukaryotic cells have an intracellular membrane network that allows them to accommodate additional membrane proteins [1, 2]; therefore, they are frequently utilized in order to express heterologous multispanning membrane proteins.

In eukaryotes, membrane proteins are targeted to specific cell compartments or to the plasma membrane by the endomembrane system, or secretory pathway, that includes the endoplasmic reticulum (ER), Golgi complex, the vacuole and other transport vesicles, and by the endocytic pathway that eventually internalizes them to the endosomes [1]. Native H<sup>+</sup>-ATPases have been used among other membrane proteins to investigate the vesicular transport machinery in the budding yeast *Saccharomyces cerevisiae* [2, 3], in which heterologous membrane proteins are usually delivered by default to the ER.

Nascent proteins in eukaryotic cells must pass stringent quality controls designed to optimize their synthesis, folding and, when necessary, post-translational modifications [4, 5]. It is not unusual that proteins expressed in eukaryotic systems fail to pass these controls being diverted to proteolytic compartments, resulting in low or, indeed, no expression. In the case of *S. cerevisiae*, one of the more advantageous systems for the heterologous expression of membrane proteins [6], these problems have been sometimes overcome by utilizing mutants defective in their vacuolar proteases [7]; however, even the latter may not be sufficient in some cases [8]. Alternatively, construction of chimeric proteins with modified N-terminal domains has also been reported to improve heterologous expression of the integral membrane protein SR Ca<sup>2+</sup>-ATPase in yeast [9]. The latter suggests that the N-terminus might be crucial to determine the fate of membrane proteins in this system.

Here, we show that the expression in *S. cerevisiae* of several bacterial and eukaryotic membrane-bound proton-translocating inorganic pyrophosphatases (H<sup>+</sup>-PPases), a type of membrane-embedded proteins naturally absent in yeast, can be improved by attaching different signal peptides to their N-termini. Naturally occurring H<sup>+</sup>-PPases are located in plasma membrane vesicles and acidocalcisomes of prokaryotes and acidic single-membrane organelles (acidocalcisomes, lysosomes, vacuoles) of certain eukaryotes: protists and plants [10-13], although a plasma membrane localization was also reported for plant-like K<sup>+</sup>-stimulated H<sup>+</sup>-PPase of some parasitic protists [14]. In this work, chimeric H<sup>+</sup>-PPase proteins were expressed in YPC3, a novel version of a yeast conditional mutant in its essential cytosolic pyrophosphatase (Ipp1p), which is unable to grow on glucose [15]. In most cases, expression levels of the chimeras were higher than those of their natural counterparts; this usually improved the ability of the different H<sup>+</sup>-PPases to support growth of YPC3 on glucose by functionally complementing Ipp1p. In the case of *A. thaliana* vacuolar H<sup>+</sup>-PPase AVP1, a protein already expressed efficiently in yeast [15, 16], both expression levels and membrane-associated PPase activity values of the chimeras were similar to those of the native protein, but density gradient ultracentrifugation showed altered subcellular distributions, suggesting different progressions along the secretory pathway. In order to further investigate this effect, GFP was fused to AVP1, and two different N-terminal signal sequences were attached to the

resulting chimeric polypeptide. Fluorescence microscopy analysis of yeast cells expressing these chimeras showed that N-terminal domains substantially modify the subcellular distribution of the heterologously-expressed H<sup>+</sup>-PPase proteins.

## EXPERIMENTAL

### Generation of YPC3 strain

YPC3 mutant strain was obtained from *S. cerevisiae* haploid strain W303-1A (MATa, *ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1, ura3-1*) by inserting the yeast *HIS3* cassette from plasmid pUG34 (<http://mips.helmholtz-muenchen.de/proj/yeast/info/tools/hegemann/gfp.html>) followed by the galactokinase (*GAL1*) promoter between the promoter and the coding sequence of the gene that codes for the cytosolic inorganic pyrophosphatase (*IPP1*) using the single-step transplacement procedure [17]. For this purpose, an appropriate linear DNA fragment was obtained as follows: first, the coding sequence of *IPP1*, along with a 390 bp-long region upstream the putative start codon [18], was amplified from genomic DNA by high fidelity PCR using *TaqPlus* DNA polymerase (Stratagene) and ligated into p-GEM-T plasmid (Promega), following the manufacturers' instructions. Then, two DNA fragments containing the yeast *HIS3* cassette (1.7 kb long) and the *GAL1* promoter (0.75 kb long), respectively, were sequentially introduced at the *EcoRV* site located 21 bases upstream the starting codon of *IPP1* [18] by standard cloning techniques [19]. This yielded plasmid pYPC3 that was digested with *ApaI* and *SacI* and, then, introduced in *S. cerevisiae* haploid strain W303-1A, by the LiOAc-PEG method [20]. Transformants were selected by growing cells on 2% agar plates made in galactose-containing culture medium [21] without histidine. The insertion of the construction was checked by PCR performed with genomic DNA of one of the transformants unable to grow on glucose (YPC3).

### Multiple sequences alignments and signal peptide predictions

Multiple amino acid sequence alignments of selected eukaryotic and prokaryotic H<sup>+</sup>-PPases were done using the Clustal X v.1.8 program [22] and corrected manually. Sequence data from public databases and unfinished genome projects were obtained by similarity searches using BLAST algorithms against websites of the National Center of Bioinformatics (NCBI), USA (<http://www.ncbi.nih.gov/PMGifs/Genomes/allorg.html>), the Joint Genomic Institute (JGI), USA ([http://spider.jgi-psf.org/JGI\\_microbial/html/](http://spider.jgi-psf.org/JGI_microbial/html/)), the Sanger Institute, UK (<http://www.sanger.ac.uk/Projects/>), or the Institute for Genomic Research (TIGR), USA (<http://www.tigr.org/tdb/mdb/mdb.html>). N-terminal extension analyses of different H<sup>+</sup>-PPases for prediction of signal peptides were performed with the SignalP 3.0 [23] and SIG-Pred programs, available at <http://www.cbs.dtu.dk/services/SignalP/> and [http://bmbpcu36.leeds.ac.uk/prot\\_analysis/Signal.html](http://bmbpcu36.leeds.ac.uk/prot_analysis/Signal.html), respectively.

### Plasmid constructions

Several derivatives of the *URA3*-containing *E. coli/S. cerevisiae* shuttle plasmid pRS699 [24] were obtained in order to express different proteins in YPC3. As a first step, an artificial *SpeI* site was introduced by PCR downstream the *XhoI* site located between the *PMA1* promoter and terminator of pRS699. This allowed directional cloning in the resulting plasmid (named pRS699 b). The coding sequences of the H<sup>+</sup>-PPase (*TcVP*) the parasitic protist (kinetoplastid) *Trypanosoma cruzi* strain Y (accession number AF159881) and the *S. cerevisiae* gene for secreted invertase precursor *SUC2* (accession number AAFW02000123) were introduced into pRS699b following the same strategy: high fidelity PCR amplification (from cDNA or genomic DNA, respectively) with primers containing artificial *SalI* and *SpeI* sites, digestion

of the amplified fragments with these enzymes and ligation to pRS699b cleaved with *XhoI* and *SpeI*. These constructions were subsequently utilized as templates in order to obtain the *PMA1* promoter followed by the sequences coding for the N-terminal domains containing the signal sequences of TcVP and Suc2p by PCR (Table S1). An artificial *EcoRI* site at the 5'-end and two consecutive *SalI* and *SpeI* sites at the 3'-end of each of the amplified DNA fragments were introduced, so that they could be inserted in plasmid pRS699b by utilizing the *EcoRI* and *SpeI* sites of the latter. This yielded plasmids pTC and pINV. Plasmids pHisTC and pHisINV were obtained by the same procedure except that a sequence coding for the epitope MGRGSH<sub>6</sub> was introduced at the 5'-ends of *TgVP* and *SUC2* in their amplification reaction. Finally, the gene coding for yeast-enhanced GFP (yEGFP) was obtained by PCR using plasmid pUG34 (see above) as a template and introducing artificial *XhoI* and *SalI* sites at the 5'-end and 3'-end, respectively. The resulting DNA fragment was ligated in the right sense at the unique *XhoI* site of pRS699b and *SalI* sites of pTC and pINV, respectively. This yielded plasmids pGFP, pTcGFP and pInvGFP. All these plasmids were sequenced to rule out possible errors produced by the PCR reactions.

The coding sequences of different H<sup>+</sup>-PPases were amplified by PCR from genomic DNA preparations or cDNA clones using primers with appropriate artificial restriction sites and inserted between the unique *XhoI/SalI* and *SpeI* sites of plasmids pRS699b, pTC and pINV. *AVP1* (accession number A38230) was also introduced in plasmids pHisTc, pHisINV, pGFP, pTcGFP and pInvGFP. Another version of *AVP1* containing the sequence coding for epitope MGRGSH<sub>6</sub> at the 5'-end, obtained by PCR, was inserted in plasmid pRS699b, yielding pHisAVP1. All plasmids utilized for subsequent experiments are listed in Table S2.

### Transformation and growth of transformed YPC3 cells

*S. cerevisiae* mutant strain YPC3 was transformed with the plasmids shown in Table 2 as described [20]. YPC3 cells were initially grown at 30 °C in galactose-containing synthetic medium devoid of histidine. Transformants were selected by growing cells on 2% agar plates in culture medium without histidine and uracil. Complementation studies of the cytosolic inorganic pyrophosphatase by the H<sup>+</sup>-PPases were performed by pre-inoculating 2 ml of galactose-containing selective medium with transformed cells from the plates and growing overnight at 30 °C with agitation (200 r.p.m.). The following day, 2 ml of glucose-containing selective medium were inoculated with 20 µl of cells grown on galactose and allowed to grow as above. This treatment is necessary to bring down the pyrophosphatase activity associated to Ipp1p [15]. After growth on glucose, serial 10-fold dilutions of the cultures were made in sterile water and 10 µl drops of each dilution were spotted onto YPD and YPGal agar plates. Plates were grown at 30 °C for 5-6 days.

### Isolation of yeast membranes and sucrose gradient fractionation

Preparation of membrane fractions from YPC3 cells transformed with the different plasmids were obtained by a modification of a method previously described [24]: yeast colonies were collected from a plate and liquid-grown up to stationary phase in galactose-containing selective medium; then, 4 ml of stationary cultures were used to inoculate 400 ml of YPD. After overnight growth, cells were sedimented by centrifugation at 700 g for 10 min, washed thoroughly with water, resuspended in 5 ml of ice-cold buffer A (25 mM Tris-HCl, pH 8, 10% glycerol, 4 mM β-mercaptoethanol, 2 mM DTT, 2 mM EDTA, 10 mM MgCl<sub>2</sub>, 1 mM benzamidine, 2 mM ε-aminocaproic acid, 1 mM PMSF), and homogenized by vigorous shaking with glass beads. The homogenate was diluted up to 25 ml with buffer B (10 mM Tris-HCl, pH 7.6, 10% glycerol, 2 mM DTT, 1 mM EDTA) and centrifuged for 10 min at

700 *g* to remove beads and debris. The resulting supernatant was centrifuged for 20 min at 20,000 *g*. The “high density membranes” (HDM) pellet thus obtained was homogenized in stripping buffer [25] and sedimented (20 min at 20,000 *g*), whereas the supernatant was further centrifuged for 30 minutes at 120,000 *g* thus yielding a “low density membranes” (LDM) pellet. This pellet was also taken up and homogenized in stripping buffer and centrifuged for 20 minutes at 120,000 *g*. Finally, both HDM and LDM pellets were washed separately with buffer B and centrifuged (20 min, 120,000 *g*). The final pellets were resuspended and homogenized in 1 ml of buffer B.

HDM preparations obtained from mutant YPC3 expressing the different versions of AVP1 were applied onto continuous 20-50% (w/w) sucrose gradients made by diffusion [24]. After an overnight centrifugation (12 h.) at 100,000 *g*, fractions of approximately 0.6 ml were collected and distributed in microfuge tubes using a peristaltic pump.

### **Pyrophosphatase activity and H<sup>+</sup>-translocation assays, Western blot and total protein estimation**

Pyrophosphatase activity and proton-translocation assays, and Western blots were performed as described elsewhere [26-28]. Proton-translocation was assayed by recording the quenching of 9-amino-6-chloro-2-methoxy acridine (ACMA). This has been shown to be the probe of choice for the measurement of relatively small  $\Delta$ pH gradients [29]. For immunodetection of H<sup>+</sup>-PPases, two primary antibodies were utilized: one, raised against the synthetic peptide HKAAVIGDTIGDPLKC, a consensus H<sup>+</sup>-PPase sequence [16], was obtained from SIGMA (St. Louis, MI, USA) and routinely utilized at 1:1000 dilution. The other was a polyclonal antibody raised against the purified H<sup>+</sup>-PPase from the marine bacterium *Thermotoga maritima* [30]. This antibody was affinity-purified as described elsewhere [31] and utilized at 1:100 dilution. For studies about the processing of N-terminal ends of chimeric proteins, a monoclonal antibody against the epitope RGSH<sub>4</sub> (QIAGEN, Germantown, MD, USA) was utilized. For localization of different membrane fractions in gradients the following antibodies against marker proteins were used: anti-Pma1p (polyclonal, generously provided by Prof. R. Serrano), anti-Vma1p, anti-Vph1p and anti-Dpm1p (monoclonals, purchased from Molecular Probes, Eugene, OR, USA). Total protein was estimated by the Bradford method [32] with ovoalbumin as a standard.

### ***In situ* fluorescence microscopy methods**

YPC3 cells transformed with plasmids pGFP-VP1, pTcGFP-VP1 and pInvGFP-VP1 and grown in YPD were fixed with 70% ethanol and stained with DAPI as described elsewhere [33]. FM4-64 (Molecular Probes), a vital lipophilic stain that shows red fluorescence when bound to membranes of acidic internal compartments, was used with Texas Red filters for colocalization studies with GFP-tagged H<sup>+</sup>-PPase [34]. Images were taken with a fully automated Leica DM6000B microscope (Leica Microsystems, Wetzlar, Germany) with a 100x objective and equipped with a cooled CCD camera (ORCA-AG, Hamamatsu Photonics, Hamamatsu, Japan) with a Z-optical spacing of 0.2  $\mu$ m.

## RESULTS

### The H<sup>+</sup>-PPase from the kinetoplastid protist *Trypanosoma cruzi* efficiently complements Ipp1p and has a putative N-terminal signal peptide sequence

A yeast mutant bearing an insertion of the *HIS3* cassette and the *GAL1* promoter between the promoter and the coding sequence of gene *IPP1* was generated. The chromosomal insertion was checked by PCR, thus, a fragment of approximately 3.9 kb was amplified using genomic DNA of a selected transformant (YPC3) as template whereas a 1.3 kb long fragment was obtained in the case of strain W303-1A (Fig. 1). Analysis by *Northern blot* and activity assays performed with YPC3 cells grown up to stationary phase on galactose and, then, transferred to glucose for further 12 hours showed that *IPP1* was transcribed only in the presence of galactose (not shown), thereby demonstrating that the only functional copy of this gene in YPC3 was under the control of the *GAL1* promoter.

Transformation of YPC3 with a plasmid bearing the coding sequence of *T. cruzi* H<sup>+</sup>-PPase (TcVP) under the control of constitutive *PMA1* promoter allowed the mutant to recover the capacity to grow efficiently on glucose (Fig. 2A). Activity assays further showed K<sup>+</sup>-stimulated PPase activity in membrane preparations of the former cells and no activity in cytosolic fractions (not shown). A detailed comparative analysis of the deduced amino acid sequence of this protein revealed an extra N-terminal domain with respect to the majority of H<sup>+</sup>-PPases from other sources, such as those well studied ones from the plant *Arabidopsis thaliana* or the proteobacterium *Rhodospirillum rubrum* (RVP). The H<sup>+</sup>-PPases from other parasitic protists like the kinetoplastids *Trypanosoma brucei* (TbVP) and *Leishmania major* (LmVP) or the apicomplexan *Toxoplasma gondii* (TgVP1) also showed extra N-terminal domains of diverse length (Fig. 2B); however, none of these proteins supported YPC3 growth on glucose (not shown). Interestingly, whereas the three analyzed kinetoplastid sequences exhibit definite N-terminal signal sequences in the range of 20 to 24 amino acids including a putative transmembrane region as predicted by the SignalP and SIG-Pred methods, the *T. gondii* H<sup>+</sup>-PPase precursor holds a longer N-terminal extension with a not so well defined bipartite structure (see Fig. 2B). No signal sequences were predicted for the other H<sup>+</sup>-PPases from both prokaryotes and eukaryotes organisms analyzed. It should be noted that the H<sup>+</sup>-PPase of *T. cruzi* has been previously identified by immunochemical methods in the Golgi apparatus and, to a lesser extent, in plasma membrane vesicles of this parasitic protist [14].

### Addition of signal sequences at the N-terminus of several H<sup>+</sup>-PPases increases their expression levels and/or their capacity to functionally complement Ipp1p

Constructs composed of the 5'-end region of TcVP (its predicted 24 amino acids long signal peptide, plus 4 residues after the putative cleavage site) (see Table S1) followed by the coding sequences of different genes for H<sup>+</sup>-PPases were obtained and inserted in the cloning site of a multicopy shuttle plasmid under the control of a constitutive promoter. Similar constructions made with the 5'-region of yeast gene *SUC2*, encoding the 19 amino acids long signal sequence of secreted invertase (see Table S1), were also tested for comparison. YPC3 was transformed with the different plasmids thus obtained (Table S2) and growth of the transformants on glucose was checked. Two main scenarios were observed: (1) several H<sup>+</sup>-PPases could not functionally substitute for Ipp1p originally but gained this ability when an appropriate N-terminal signal domain was added, and (2) *A. thaliana* AVP1 could complement Ipp1p in its original form but this capacity was clearly enhanced when fused with any of the N-terminal domains tested. In one case, that of LmVP, the resulting chimera showed expression levels significantly higher than the natural protein; however, it could not

functionally substitute for Ipp1p (not shown).

Figure 3 shows the results obtained with the  $K^+$ -independent  $H^+$ -PPase from the photosynthetic proteobacterium *R. rubrum* (RVP). Transformation of YPC3 with plasmids pRVP and pTcRVP did not allow the mutant to grow on glucose; however, cells transformed with plasmid pInvRVP efficiently grew on this carbon source. Assays of PPase activity and Western blot analysis further showed the presence of significant amounts of a  $K^+$ -independent  $H^+$ -PPase with an apparent molecular mass (ca. 60 kDa) smaller than that of other  $H^+$ -PPases in membrane fractions of the latter cells. Interestingly, InvRVP was by far the most effectively membrane targeted protein (see Fig. 3C). Similar results were obtained with another prokaryotic  $K^+$ -independent  $H^+$ -PPase, from the actinobacterium *Streptomyces coelicolor* (not shown). In the case of the  $K^+$ -dependent  $H^+$ -PPase TgVP1 of the apicomplexan protist *T. gondii*, only YPC3 cells transformed with plasmid pTcTgVP, encoding a fusion protein with the putative signal sequence of *T. cruzi*  $H^+$ -PPase, could recover the capacity to grow on glucose. PPi hydrolysis activity assays and Western blots consistently demonstrated the presence of a  $K^+$ -dependent PPase in membranes of these cells; however, yeast cells transformed with plasmid pInvTgVP, encoding a fusion protein with the signal sequence of yeast Suc2p, also expressed the protein (Fig. S1). YPC3 cells transformed with plasmid pTcAVP1 could grow on glucose more efficiently than those transformed with plasmids pAVP1 or pInvAVP1; however, activity assays and Western blot analysis showed no significant differences in membrane-associated activity or protein levels (Fig. 4). In all cases,  $H^+$ -translocation activities of membrane preparations were proportional to the PPi hydrolysis activities tested (Figs. 3D, S1D and 4D).

#### **The N-terminal signal sequences from TcVP and Suc2p are processed when the chimeras with AVP1 are expressed in YPC3**

In order to check whether the N-terminal signal sequences of TcVP and Suc2p were effectively processed during delivery and membrane sorting of the chimeric proteins, YPC3 cells were transformed with plasmids pHisAVP1, pHisTcAVP1 and pHisInvAVP1 (Table S2). Immunoblots of membrane preparations obtained from the different transformants using an antibody against the epitope RGS<sub>H4</sub> recognised a polypeptide only in the case of cells transformed with pHisAVP1; however, the antibody against a highly conserved  $H^+$ -PPase sequence identified polypeptides of the right size (ca. 70 kDa) in all cases (Fig. S2). Activity assays data were similar to those shown in Fig. 4. These results clearly indicate that the N-terminal domain of TcVP holds a *bona fide* signal sequence that, like the homologous signal peptide of Suc2p, is correctly processed in yeast. To our knowledge, this is the first direct evidence reported for protein processing and sorting of a  $H^+$ -PPase, in a heterologous system in this case. An N-terminal fusion of TcVP with the RGS<sub>H4</sub> domain that is functionally expressed in YPC3 seems to be processed as well, but unfortunately the chimeric protein is not properly recognised by the  $H^+$ -PPase antibodies used in this study (data not shown).

#### **Natural AVP1 and its derived N-terminal chimeras TcAVP1 and InvAVP1 exhibit different subcellular distributions in yeast cells**

Fractionation by sucrose gradient ultracentrifugation of high density membranes obtained from yeast cells transformed with plasmids pAVP1, encoding natural plant vacuolar AVP1, and pTcAVP1 and pInvAVP1, encoding derived N-terminal chimeras with signal sequences, showed significant differences in the subcellular distributions of  $H^+$ -PPase protein and PPi hydrolytic activity. In the case of pAVP1, activity was preferentially associated to internal membranes (Golgi complex/vacuoles), with some activity associated to the plasma

membrane. In cells transformed with pInvAVP1, an analogous distribution was found, although plasma membrane-associated activity was clearly higher. In contrast to that, membrane preparations from cells transformed with TcAVP1 showed virtually all the PPase activity to be associated to ER/Golgi/vacuolar membranes (Fig. 5). Western blot analyses showed AVP1 to be evenly distributed between gradient fractions 3 and 12 (being fraction 1 the bottom of the gradient), whereas TcAVP1 appeared preferentially in fractions 8-12 (Golgi complex/vacuole). Strikingly, in the case of InvAVP1 most of the protein appeared associated to the plasma membrane (fractions 3-5) (Fig. 5).

### ***In situ* fluorescence microscopy confirms subcellular distribution results obtained with sucrose density gradients**

The question of the subcellular distribution of the different versions of AVP1 was further investigated by fluorescence microscopy of YPC3 cells. Transformants with plasmids pGFP-VP1 and pInvGFP-VP1, encoding respectively a GFP-VP1 fusion protein without and with the Suc2p signal sequence, showed a very characteristic pattern of green fluorescence around the nucleus (cisternae vesicles of the ER) and in the plasma membrane. On the contrary, green fluorescence in cells transformed with plasmid pTcGFP-VP1 appeared exclusively associated to the internal membrane network, showing a spread spotted distribution throughout the YPC3 cells (Fig. 6A). Labelling of the different transformed yeast cells with FM4-64 clearly shows that GFP-VP1 and InvGFP-VP1 do not co-localize with the vacuolar membrane; by contrast, TcGFP-VP1 seems to partially co-localize with a high number of small acidic vesicles (Fig. 6B).

## **DISCUSSION**

A *S. cerevisiae* haploid strain (YPC3), whose gene coding for the essential cytosolic soluble PPase (*IPP1*) is under the control of the glucose-repressible galactokinase (*GAL1*) promoter, has been generated by the single-step transplacement procedure. This approach resulted in the insertion of the yeast *HIS3* marker gene followed by the *GAL1* promoter between the promoter and the coding sequence of *IPP1*. Consequently, the conditional mutant strain generated YPC3 cannot grow in the presence of glucose because Ipp1p, essential for growth, is not expressed under these conditions. YPC3 is more convenient than YPC1, a mutant with the same phenotype previously described [15], to check functional complementation of Ipp1p due to its higher genomic stability.

The H<sup>+</sup>-PPase of the kinetoplastid protist *T. cruzi* (TcVP) has a putative N-terminal extension with the hallmarks of type I membrane protein signal sequences, as determined by *in silico* studies: a hydrophobic core, which presumably comprises a transmembrane (TM) segment, with two flanking groups of charged residues and a putative peptidase cleavage site at the C-terminus of the signal peptide [35]. The observation that the growth of YPC3 on glucose was efficiently supported by TcVP prompted us to investigate the possible effects that this domain (absent in many other proteins of this class) might have on the heterologous expression in yeast of other H<sup>+</sup>-PPases available in our laboratory. Comparative studies of these effects with those obtained with chimeras made with the N-terminal domain of a *S. cerevisiae* protein were also performed. Suc2p N-terminal signal peptide was chosen for two main reasons: (a) the initial steps of multispinning membrane proteins translation are analogous to those of secreted proteins synthesis [36], (b) this signal peptide had been studied in significant detail and utilized to efficiently translocate polypeptides across the ER membrane [37-39].

Plasmids bearing sequences coding for either of these N-terminal peptides followed by those of RVP and TgVP1 significantly increased the expression levels of the corresponding proteins in yeast. This improvement was high enough to functionally complement Ipp1p. Isolation of membrane fractions, assaying of membrane-associated PPI-hydrolysis activity as well as Western blot analyses revealed that expression levels of the chimeric proteins were higher than those of their original counterparts. It should be noted, however, that the achieved targeting efficiencies varied significantly between chimeric H<sup>+</sup>-PPases from different sources, e.g., N-terminal fusions with the Suc2p signal sequence dramatically enhanced protein and activity levels of prokaryotic H<sup>+</sup>-PPases, whereas both Tc- and Inv- fusions showed similar effectiveness with their eukaryotic counterparts. On the other hand, it was previously reported that the full-length TgVP1 gene cannot be expressed in vacuolar protease-deficient *S. cerevisiae* strains unless its N-terminal domain is removed [8]. This could be explained by assuming that the latter is not recognized by the yeast signal recognition particle (SRP) and, consequently, the polypeptide synthesis would be stopped at the initial steps of translation. Addition of the TcVP N-terminal signal sequence to TgVP1 would allow this recognition and, therefore, the expression of the chimeric protein in our non-deficient vacuolar protease yeast strain. The fact that a signal peptide from a trypanosomatid protist such as *T. cruzi* may be efficiently recognised by *S. cerevisiae* SRP supports the idea that components of the SRP pathway and salient features of the molecular mechanism of SRP-dependent protein targeting are conserved among different organisms [40]. Altogether, these results show that addition of appropriate N-terminal signal domains to multispinning membrane proteins can enhance their heterologous expression in yeast, presumably by increasing the targeting of the proteins to the ER, a proposal that is consistent with the results obtained with fluorescence microscopy of GFP-AVP1 fusions (see below). However, it is necessary to try different combinations of N-termini and coding sequences in order to optimize expression, as it is not possible to know before hand whether a given chimeric polypeptide will be effectively expressed in this system; that is, a trial-and-error approach is still needed. In any case, we have developed a useful set of plasmids where the desired ORF can be inserted downstream the sequences of several N-terminal signal peptides yielding different chimeras whose expression levels can be readily tested in *S. cerevisiae*.

AVP1 turned out to be a different case: the ability of this protein to functionally complement Ipp1p was significantly improved by attaching TcVP N-terminal domain to its N-terminus, although only a slight improvement was obtained with Suc2p; however, similar membrane-associated PPI hydrolytic activity and protein levels were observed for both chimeras. By utilizing an antibody against the epitope RGSH<sub>4</sub>, it was shown that both TcVP and Suc2p N-terminal peptides added were processed when attached to AVP1. This may not be significant in the case of Suc2p; however, it strongly suggests that the N-terminus of TcVP is fully recognisable by *S. cerevisiae* type I signal peptidases [41]. This idea was further supported by the fact that identical results were obtained with other yeast strains, such as W303-1A and the vacuolar protease-deficient mutant BJ5457 [7] (data not shown). N-terminal signal peptides have been shown to occur in all organisms studied to date and share some conserved general features (see Fig. 2), consequently, they have been extensively used fused with foreign soluble proteins in order to improve expression in yeast although with varied results, revealing that not all signal sequences are functionally equivalent. Thus, species-specificity of signal peptide utilization has been demonstrated not only between divergent organisms. i. e., mammalian and yeast [42], but also between closely related species [43]. This seems to be also the case for heterologous membrane proteins (mostly transporters and receptors) expressed in yeast, although the information available is much scantier. Here,

we report the first direct evidence obtained so far for protein precursor processing of a proton pump, in this case, a member of the H<sup>+</sup>-PPase family. On the other hand, it was important to check this point as we aim at optimizing production of these membrane proteins in yeast with minimal changes with respect to the natural polypeptide.

Results with TgVP I and AVP1 demonstrate that, although a threshold of PPase activity is required to complement Ipp1p, this may not be enough and some other biologically relevant features, such as subcellular distribution, are important. The possibility that AVP1 and its derived chimerae, TcAVP1 and InvAVP1, might have different locations within the cell was studied by sucrose density gradient ultracentrifugation. Previous separation of the yeast membrane preparation in "high density" (HDM) and "low density" (LDM) fractions was essential to achieve the desired resolution in the gradients. Fractionation of HDM indicated that AVP1 was rather evenly distributed at different membranes, although activity was maximal at internal membranes (ER/Golgi complex/vacuole). The chimera TcAVP1 and its associated activity seemed to be preferentially located in internal membranes, showing a rather homogeneous intracellular distribution; however, in the case of InvAVP1 a striking situation was observed: although most of the polypeptide was found at the plasma membrane, membrane-bound K<sup>+</sup>-stimulated PPi hydrolytic activity was higher in internal membranes. This result suggests two important points: (a) the N-terminus of Suc2p can preferentially target membrane-bound proteins to the plasma membrane, and (b) the lipid composition of the membrane where InvAVP1 is located may significantly modulate its activity. Thus, according to the results obtained with AVP1 and InvAVP1, the plasma membrane does not seem to provide an optimal lipid environment for this protein, naturally occurring in plant vacuolar membrane (tonoplast) [11, 12]. It is worthwhile pointing out that modulation of activity by lipids has been reported for other proton pumps, such as plant plasma membrane H<sup>+</sup>-ATPases [44], a mammalian vacuolar H<sup>+</sup>-ATPase [27] and the H<sup>+</sup>-PPase from *Trypanosoma cruzi* [14].

Density gradient ultracentrifugation presents certain limitations at resolving subcellular fractions in yeast [24]; therefore, these studies were completed by *in situ* fluorescence microscopy studies using protein fusions with the green fluorescent protein. Initially, immunocytochemical localization with antibodies currently available in our laboratory was attempted without success. Then, fusion of GFP at the C-terminus of AVP1 [45] was carried out, but this yielded a polypeptide with no activity. This is in agreement with previous studies indicating that the C-terminus is essential for both the oligomerization and functionality of the H<sup>+</sup>-PPase proteins in the membrane [46]. Consequently, a fusion protein in which the yEGFP was attached at the N-terminus was constructed. The resulting polypeptide was efficiently expressed in YPC3 as demonstrated by its capacity to complement Ipp1p and the high levels of K<sup>+</sup>-dependent membrane-associated PPi hydrolytic activity found; actually, GFP-AVP1 showed higher expression levels than AVP1 (not shown). The presence of GFP at the N-terminus of some membrane proteins has been previously reported to enhance their expression levels in heterologous systems [47]. This effect is currently being investigated for other H<sup>+</sup>-PPases available in our laboratory.

Fluorescence microscopy showed a pattern of GFP-AVP1 distribution strikingly similar to that previously obtained for AVP1 by immunocytochemical localization: the protein is mainly associated to ER-derived perinuclear membranes (cisternae vesicles) with a significant fraction located at the plasma membrane and small prevacuolar vesicles [15]. This similarity was further confirmed by sucrose gradient fractionation (not shown). Therefore, the presence of GFP at the N-terminus does not seem to alter the targeting of AVP1 in yeast. Further addition of TcVP N-terminal signal domain resulted in major changes in green

fluorescence distribution, which remains associated to internal membranes. On the other hand, addition of Suc2p signal peptide resulted in a subcellular distribution similar to that of GFP-AVP1. Co-localization of TcGFP-AVP1 with the acidic vesicles membranes further shows that this chimera is effectively targeted in a functional active form to specific internal, possibly vacuolar, compartments. Altogether, fluorescence microscopy results confirmed those obtained by sucrose density gradient fractionation and strengthen the idea that the presence of an appropriate N-terminal signal peptide dramatically alters the intracellular targeting and allows further trafficking along the biosynthetic secretory pathway of H<sup>+</sup>-PPases heterologously expressed in *S. cerevisiae*.

As a final conclusion, it is important to point out that the strategy followed in this work can be utilized in concert to others found in the literature aimed at optimizing heterologous expression of membrane proteins in yeast. Actually, the expression levels of several of the chimeric proteins described here can be further increased if they are expressed in vacuolar-deficient yeast mutants [7]. All the strategies described here are already under way in our laboratory in order to get the high amounts of protein required for structural studies, such as crystallization and 2D/3D structure determination. On the other hand, results obtained with AVP1 further support that the N-terminal signal sequences alter the subcellular localization of a membrane-embedded protein. This capacity could also be utilized to preferentially target proteins to specific organelles and/or membranes within the cell, which may be crucial when performing studies of functional complementation.

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## AUTHOR CONTRIBUTION

R. Drake carried out most of the experimental work, except generation of YPC3 and fluorescence microscopy. J. R. Pérez-Castiñeira supervised R. Drake's work, designed the study, supervised data collection, generated YPC3 mutant, performed microscopy work and drafted the manuscript. A. Serrano initially conceived the study, did the *in silico* analyses, co-supervised the experimental work and data collection and contributed to the manuscript draft.

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## LEGENDS OF FIGURES

**Figure 1. Generation of YPC3, a new yeast mutant whose essential cytosolic inorganic pyrophosphatase *Ipp1p* is under the control of the galactokinase (*GAL1*) promoter.**

(A) The gene coding for the cytosolic pyrophosphatase of yeast haploid strain W303-1A was disrupted by insertion of a linear DNA fragment comprising the yeast *HIS3* cassette as a selection marker followed by the galactokinase (*GAL1*) gene promoter. Arrows represent the oligonucleotides (20 bp and 22 bp-long for the forward and the reverse, respectively) utilized as PCR primers. (B) Analysis by PCR of the insertion, genomic DNA obtained from strains YPC3 and W303-1A were utilized as templates and oligonucleotides corresponding to the promoter region and the end of the coding sequence of *IPP1* as primers. (C) Phenotypic characterization of YPC3 by checking its growth on glucose as compared with the yeast strain utilized for its generation. See main text for details.

**Figure 2. Functional complementation of the cytosolic pyrophosphatase *Ipp1p* of *S. cerevisiae* by the H<sup>+</sup>-PPase from the kinetoplastid protist *Trypanosoma cruzi* (A) and multiple sequences alignment of the N-terminal regions of H<sup>+</sup>-PPase orthologs from selected eukaryotic (bold) and prokaryotic (plain) organisms (B).**

N-terminal extensions having the general features of signal peptides, namely, a hydrophobic core and a positively charged N-terminal domain, and predicted as such by the SignalP and SIG-Pred programs are shown boxed and the corresponding putative cleavage sites indicated

by vertical arrowheads. Positively charged (K,R) and hydrophobic (A,V,L,I,W,F,M) amino acid residues in the putative signal sequences are represented in italics and in boldface with grey background, respectively. Note the bipartite signal peptide with two putative cleavage sites predicted for the H<sup>+</sup>-PPase of the apicomplexan protist *Toxoplasma gondii*, for which the N-terminal residues of truncated constructs used by Drozdowicz et al. [8] for expression in yeast are indicated by folded arrows. Approximate locations of the transmembrane segment (TMS) characteristic of signal leader peptides and the first one (TMS1) in all H<sup>+</sup>-PPases are also shown [48]. The protein sequences used are (accession numbers in parentheses): *Trypanosoma cruzi* (AAF80381); *Trypanosoma brucei* (AAK95376); *Leshmania major* (CAJ08309); *T. gondii* TgVP1 (AAK38077); *Plasmodium falciparum* PfVP1 (AAD17215); *Arabidopsis thaliana* AVP1 (A38230); *Oryza sativa* (BAA08232); *Thermotoga maritima* (D72049); *Rhodospirillum rubrum* (AAC38615); *Methanosarcina mazei* (AAM22543); *Pyrobaculum aerophilum* (AAF01029); *Streptomyces coelicolor* (T36668), and *Chlorobium tepidum* (contig g12, NC\_002932). The unpublished H<sup>+</sup>-PPase sequences from the microalga *Chlorella* sp. NC64A were identified by their JGI genome project code numbers.

**Figure 3. Drop tests (A), membrane-associated PPI hydrolysis activity assays (B), Western blot analysis (C) and traces of ACMA fluorescence quenching by membrane preparations (D) of yeast mutant strain YPC3 transformed with plasmids pRS699b, pRVP, pTcRVP and pInvRVP.**

Experiments were performed with three independent clones taken from each of the transformant plates. (A) Typical drop test. Activity values shown in (B) are means  $\pm$  S.E. (n=3) using LDM preparations obtained from YPC3 cells transformed with pRS699b as reference. White and black bars represent activities in the absence and presence of potassium chloride, respectively. (C) Immunodetection of low density membrane (LDM) preparations as described in Experimental Procedures. Approx. 70  $\mu$ g of protein were loaded per lane. (D) Typical trace of ACMA fluorescence quenching obtained with LDM preparations of YPC3 transformed with plasmid pInvRVP. No quenching was observed in the case of RVP whereas, in the case of TcRVP, an intermediate trace between those of C- and InvRVP was obtained (not shown).

**Figure 4. Drop tests (A), membrane-associated PPI hydrolysis activity assays (B), Western blot analysis (C) and traces of ACMA fluorescence quenching by membrane preparations (D) of yeast mutant strain YPC3 transformed with plasmids pRS699b, pAVP1, pTcAVP1 and pInvAVP1.**

Same conditions as in Fig. 3 except that similar traces to that of AVP1 were routinely obtained with LDM preparation of YPC3 cells transformed with plasmid pInvAVP1.

**Figure 5. Distribution of membrane-bound PPI hydrolysis activity on a continuous 20%-50% (w/w) sucrose gradient.**

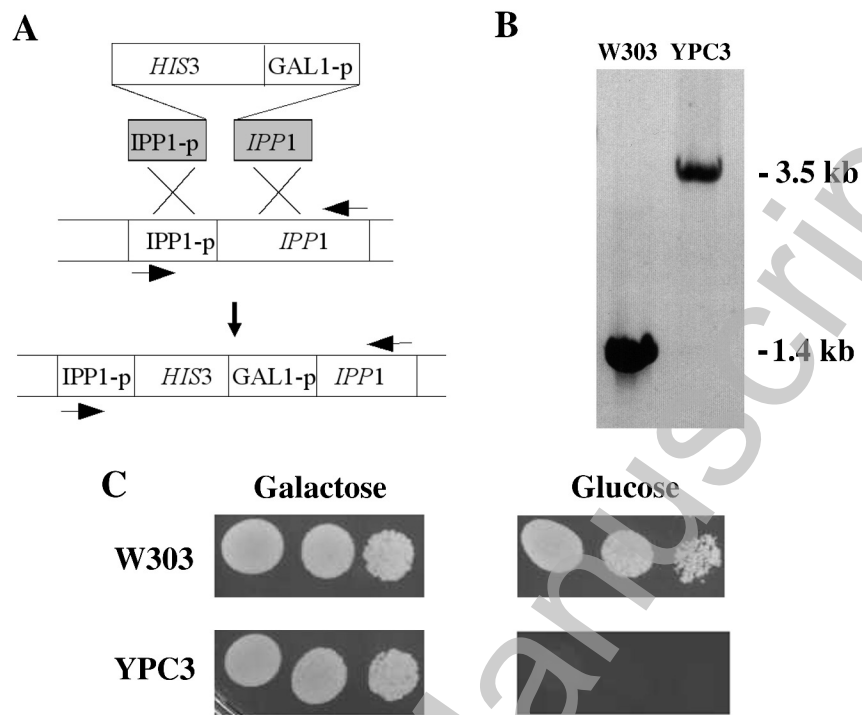
High density membrane preparations obtained from YPC3 yeast cells transformed with pAVP1 ( $\blacklozenge$ ), pTcAVP1 ( $\blacksquare$ ) and pInvAVP1 ( $\blacktriangle$ ) were loaded onto sucrose gradients as described in Experimental Procedures. After overnight centrifugation, 0.6 ml samples were collected using a peristaltic pump and activity was measured. Activity data are average of

three experiments performed with independent clones. Total PPase activity recovered from the gradients were of the same order of magnitude. Sample 1 corresponds to the bottom of the gradient. Immunoblots performed using samples 3 to 14 from the gradients and an affinity-purified antibody against the *T. maritima* H<sup>+</sup>-PPase [30, 31] are shown. Gradient samples (aprox. 100  $\mu$ l) were concentrated by precipitation with 10% TCA before loading. Antibodies against several marker proteins were also utilized to localize different yeast membranes: Vph1p, vacuolar membrane; Dpm1, ER membrane; Vma1p, vacuolar and Golgi membranes; Pma1p, plasma membrane.

**Figure 6. *In situ* fluorescence microscopy of YPC3 cells transformed with plasmids pGFP AVP1, pInvGFP AVP1 and pTcGFP AVP1.**

(A) Open field and details of a single cell are shown. Panels show clockwise from bottom left: Nomarski, GFP-tagged proteins, DAPI staining of nuclei and the overlap of the three images. (B) Visualization of GFP-tagged proteins and membranes of internal acidic compartments with red fluorescent vital dye FM4-64. Panel shows clockwise from bottom left: Nomarski, GFP-tagged proteins, FM4-64 staining and the overlap. Cells were processed and visualized as described in Experimental Procedures. Images were deconvolved using the Leica LAS-AF software. Maximum projection images are shown. White arrowheads indicate green fluorescence associated to perinuclear and plasma membranes in (A) and red fluorescence associated to internal acidic membranes in (B).

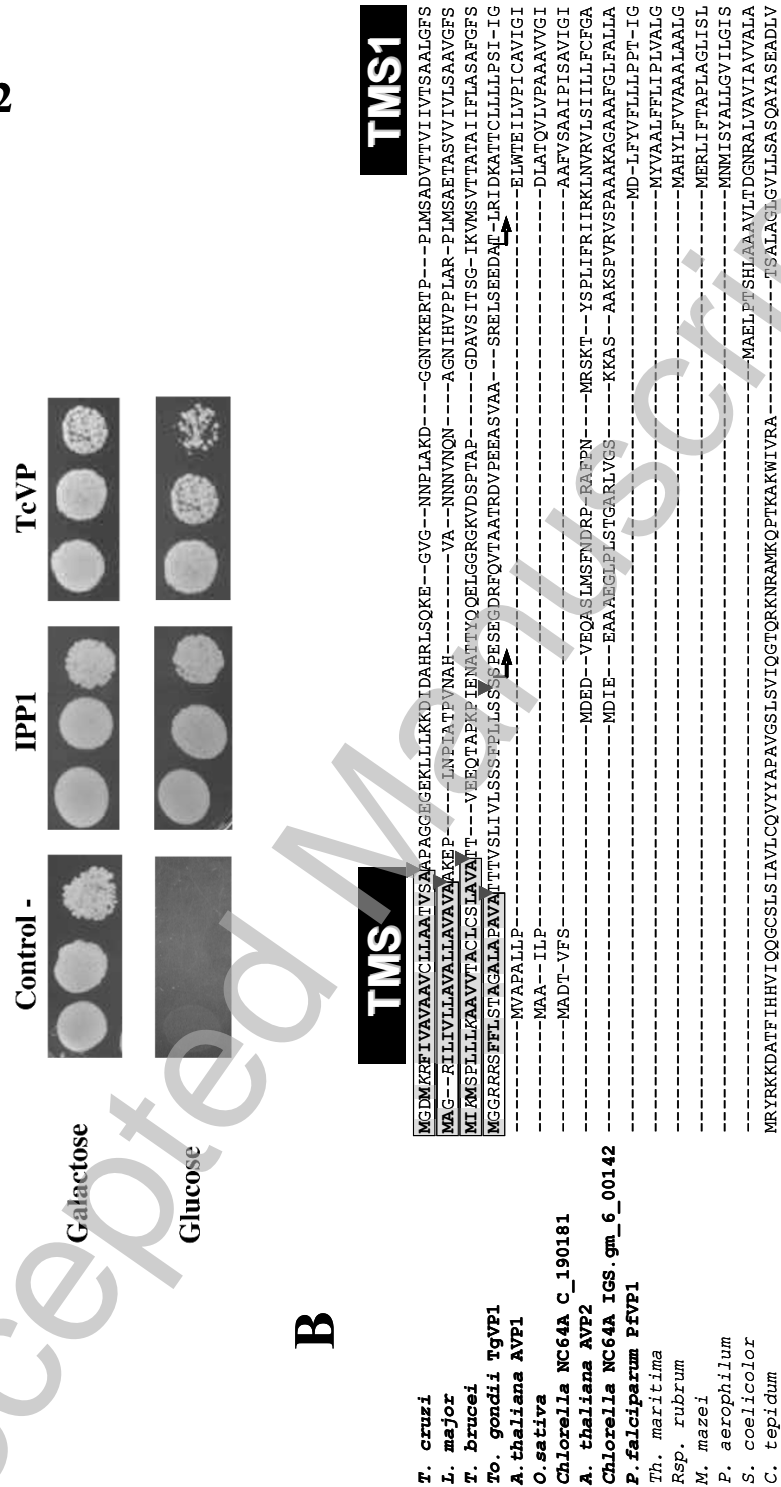
**Fig. 1**



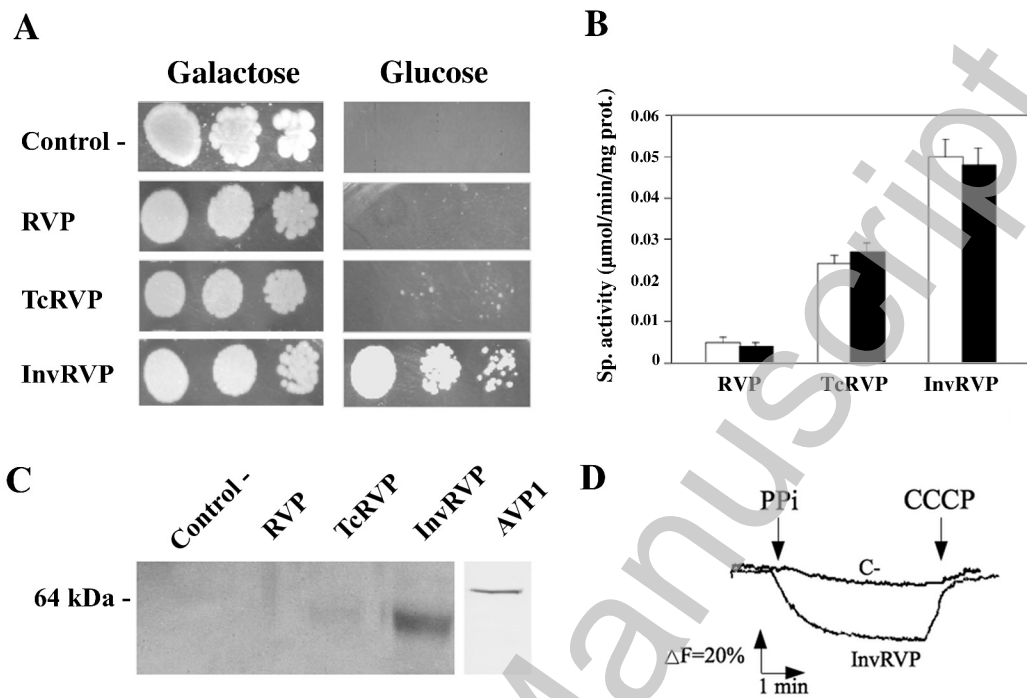
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Fig. 2



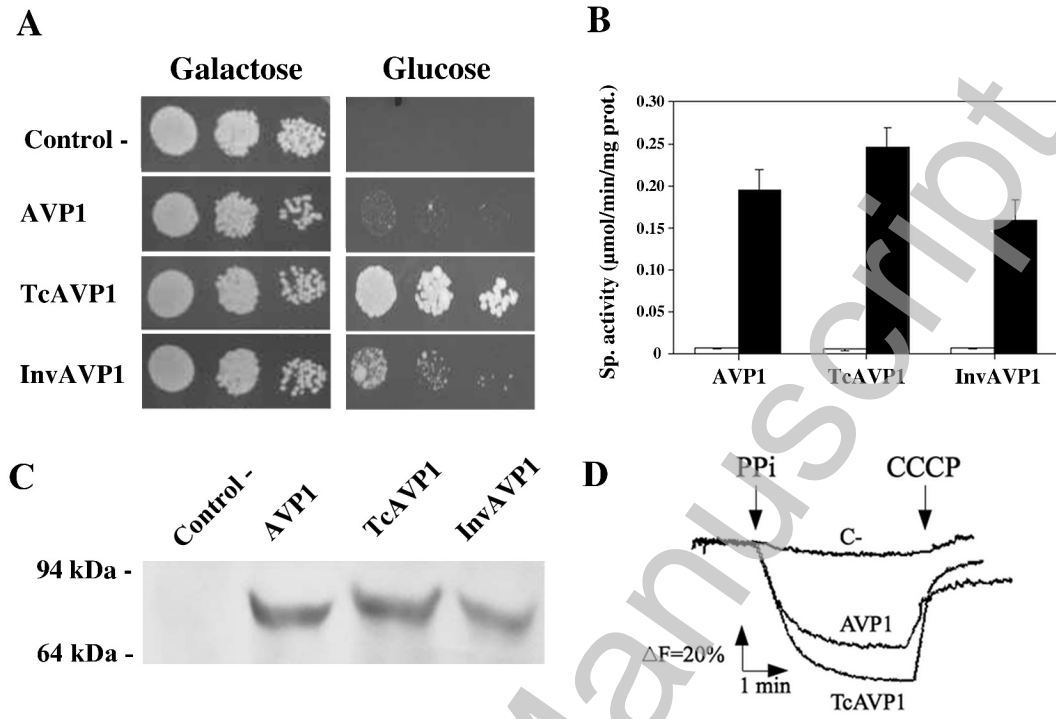
**Fig. 3**



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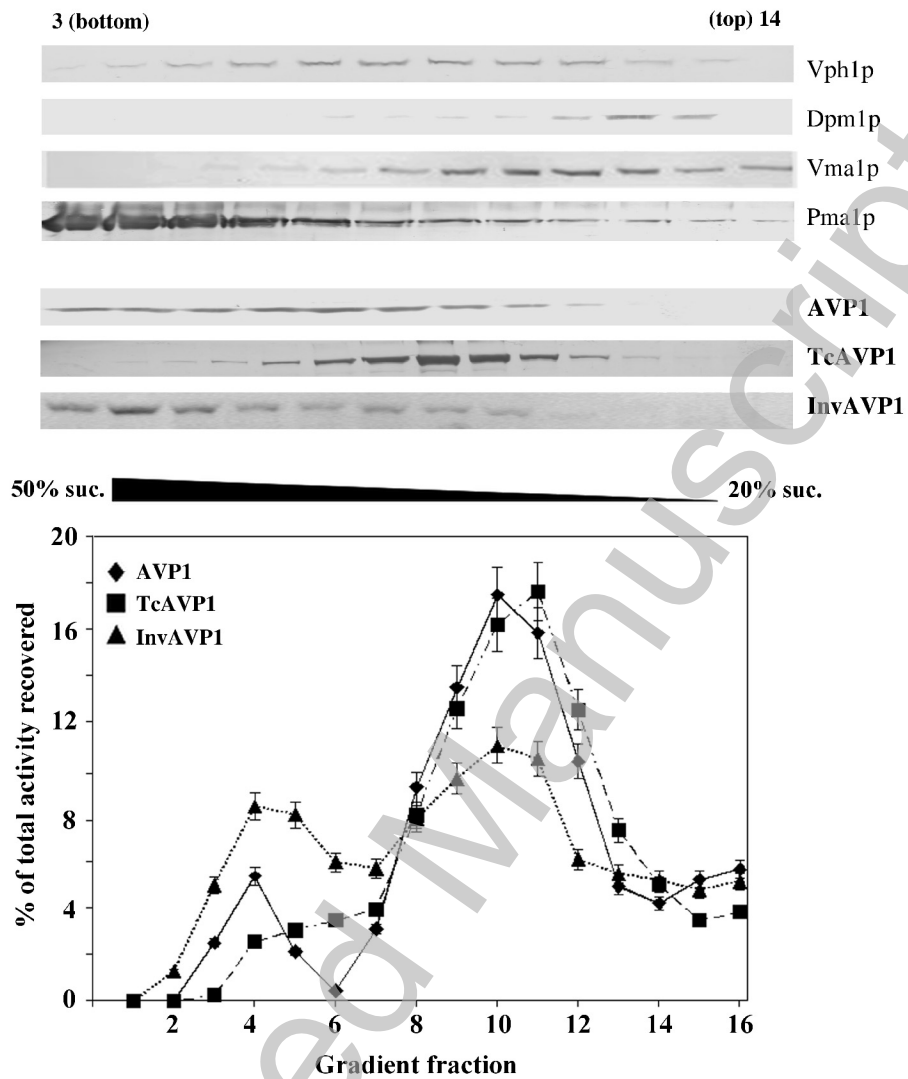
**Fig. 4**



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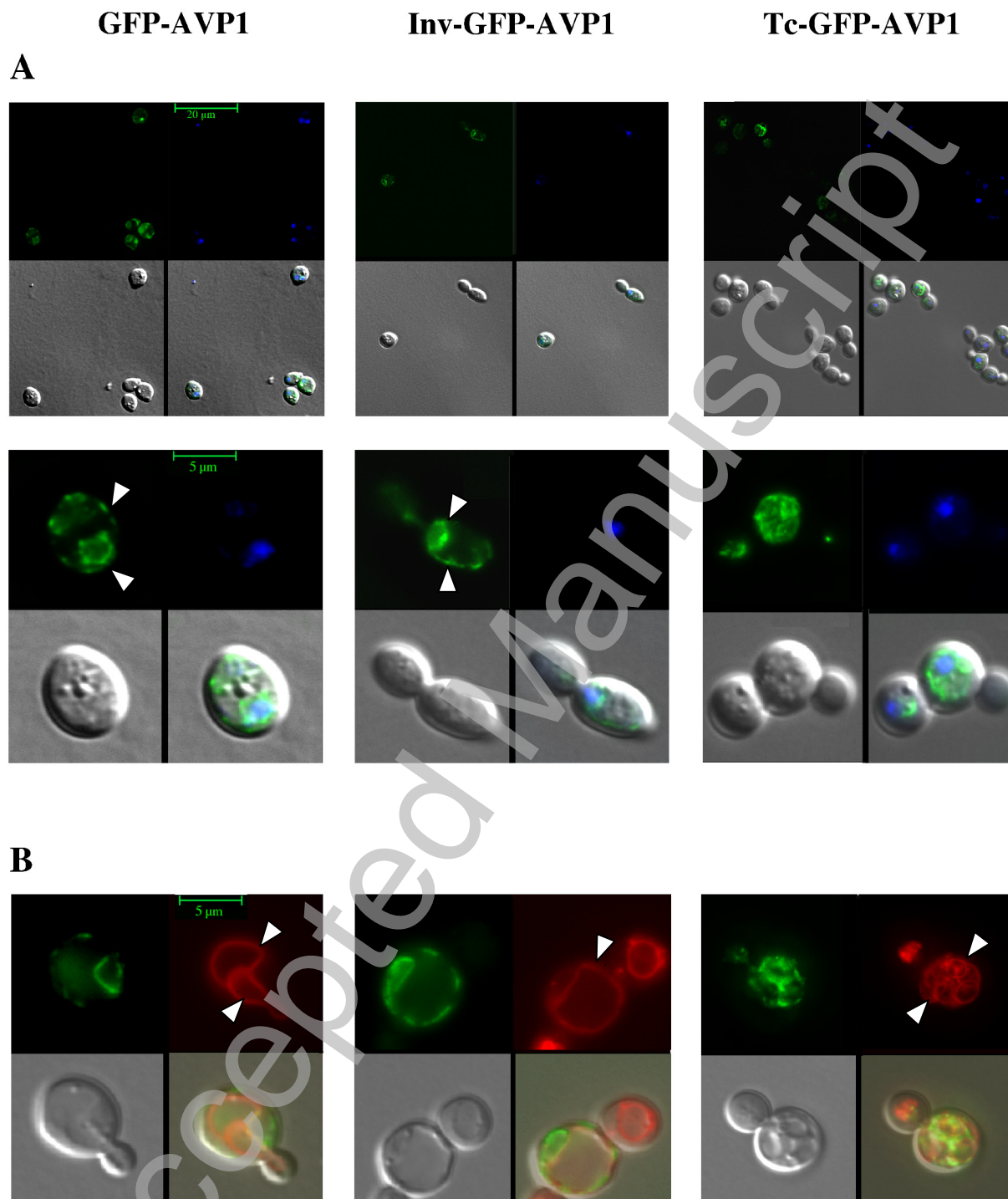
**Fig. 5**



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**Fig. 6**



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