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Calmodulin-dependent protein kinase kinase- β activates AMPK without forming a stable complex – synergistic effects of Ca^{2+} and AMP

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Abbreviations: $\alpha 1$ -KD, kinase domain of AMP-activated protein kinase, $\alpha 1$ isoform; ACC, acetyl-CoA carboxylase; AMPK, AMP-activated protein kinase; CaMKK, calmodulin-activated protein kinase kinase; GFP, green fluorescent protein; GST, glutathione-S-transferase

SYNOPSIS

Activation of AMPK by phosphorylation at Thr-172 is catalyzed by at least two distinct upstream kinases, i.e. the tumour suppressor LKB1, and calmodulin-dependent protein kinase kinase- β (CaMKK β). The sequence around Thr-172 is highly conserved between the two catalytic subunit isoforms of AMPK and the twelve AMPK-related kinases, and LKB1 has been shown to act upstream of all of them. We now report that none of the AMPK-related kinases tested could be phosphorylated or activated in intact cells or cell-free assays by CaMKK β , although we did observe a slow phosphorylation and activation of BRSK1 by CaMKK α . Despite recent reports, we could not find any evidence that the α and/or β subunits of AMPK formed a stable complex with CaMKK β . We also showed that increasing AMP concentrations in HeLa cells (which lack LKB1) had no effect on basal AMPK phosphorylation, but enhanced the ability of agents that increase intracellular Ca²⁺ to activate AMPK. This is consistent with the effect of AMP on phosphorylation of Thr-172 being due to inhibition of dephosphorylation, and confirms that the effect of AMP is independent of the upstream kinase utilized.

INTRODUCTION

The AMP-activated protein kinase (AMPK) is an energy sensing system involved in regulating energy balance at both the cellular and the whole body levels [1, 2]. The kinase occurs as heterotrimeric complexes composed of a catalytic α subunit and regulatory β and γ subunits, with each subunit existing in mammals as isoforms encoded by multiple genes ($\alpha 1, \alpha 2; \beta 1, \beta 2; \gamma 1, \gamma 2, \gamma 3$). Metabolic stresses that inhibit ATP synthesis (e.g. hypoxia, hypoglycaemia) or that stimulate ATP consumption (e.g. muscle contraction) cause an increase in the cellular ADP:ATP ratio, which is amplified by adenylate kinase into an even larger increase in the AMP:ATP ratio. AMP and ATP bind antagonistically to two sites formed by the four tandem CBS motifs on the γ subunit [3, 4]. The kinase is only active after phosphorylation of a critical threonine residue within the activation loop of the kinase domain (Thr-172 in rat $\alpha 1/\alpha 2$) by upstream kinases. The major upstream kinase in most cells was identified to be a complex between the tumour suppressor LKB1 and two accessory subunits, STRAD and MO25 [5, 6]. LKB1 appears to be constitutively active [7, 8] and may therefore phosphorylate AMPK continually, but under basal conditions the phosphate appears to be immediately removed by protein phosphatases. However, binding of AMP to the AMPK γ subunits inhibits dephosphorylation of Thr-172, an effect that is antagonized by high concentrations of ATP [9-11]. In addition, binding of AMP (but not ATP) triggers a further allosteric activation of the phosphorylated kinase by up to 10-fold, with the combination of these two effects producing >1000-fold activation [11]. Both stimulatory effects appear to occur because AMP binding relieves the inhibitory effects of an auto-inhibitory domain on the α subunit, which binds to the kinase domain on the opposite side to the substrate-binding site [12]. This represents a sensitive switch mechanism that produces a large activation of AMPK in response to a small increase in the cellular AMP:ATP ratio.

Some human tumour cells (e.g. HeLa cells) do not express LKB1, but AMPK can still be phosphorylated at Thr-172 and activated in such cells using Ca²⁺ ionophores. This led to the discovery that the Ca²⁺- and calmodulin dependent protein kinases, especially CaMKK β , could act as alternate upstream kinases that can phosphorylate Thr-172 on AMPK [13-15]. CaMKK α and CaMKK β [16] were originally discovered as Ca²⁺- and calmodulin-dependent protein kinases that acted upstream of calmodulin-dependent protein kinases I and IV. The Ca²⁺ \rightarrow CaMKK \rightarrow AMPK pathway is triggered by a rise in cytosolic Ca²⁺ without any requirement for an increase in AMP, and is responsible

for AMPK activation in response to K^+ -induced depolarization in neurones [13], muscarinic activation in neuroblastoma cells [17], thrombin activation of endothelial cells [18], treatment of smooth muscle cells with vasoconstrictors [19] and stimulation of antigen receptors in T cells [20].

The sequence around Thr-172 on the AMPK α subunits is highly conserved (see Fig. 1) in the kinase domain sequences of kinases (termed AMPK-related protein kinases), that lie on the same branch of the human kinome [21]. By studying phosphorylation in cell-free assays and in cells lacking LKB1, including LKB1^{-/-} mouse embryo fibroblasts and HeLa cells, it was shown that at least twelve of these were also dependent on LKB1 for basal phosphorylation of the threonine residue equivalent to Thr-172, and hence for basal activity. These include the brain-specific kinases BRSK1 and BRSK2 (also known as SAD-B and SAD-A), SIK1, SIK2 and SIK3 (also known as SIK, QIK and QSK), NUA1 and NUA2 (also known as ARK5 and SNARK), MARK1, MARK2, MARK3 and MARK4, and the testis-specific kinase SNRK [7, 22].

In this paper we have addressed three questions regarding the regulation of AMPK and the ARKs by CaMKKs. Firstly, it had not been established whether the ARKs are substrates for CaMKKs, as is AMPK itself. Secondly, it has recently been claimed that CaMKK β can form a complex with the α and β subunits of AMPK in the absence of a γ subunit [23], a hypothesis that we wished to confirm. Thirdly, if the effect of AMP on phosphorylation of Thr-172 on AMPK is due to inhibition of dephosphorylation we were puzzled as to why, in cells such as HeLa cells that express CaMKK β but lack LKB1, AMPK is not activated by treatments that increase intracellular AMP [5].

EXPERIMENTAL

Materials and proteins

STO-609 was from Tocris Bioscience, Bristol, UK, and calmodulin from CN Biosciences, Nottingham, UK. A23187, phenformin and 2-deoxyglucose were from Sigma, Poole, UK. Zeocin and blasticidin were from Invitrogen, Renfrew, UK. Expression and purification of GST-AMPK- α 1, GST-BRSK1, GST-BRSK2, GST-LKB1:STRAD:MO25 and GST-CaMKK β were as described previously [24]. For GST-CaMKK α , a pGEX plasmid encoding CaMKK α (residues 1-505) was used to transform *E. coli* (BL21 (DE3) pLys), which were grown in LB ampicillin at 37°C until A_{600} was 0.4-0.6. Expression was induced with 50 μ M IPTG and cells harvested after 18 hr at 15°C. The harvested cells were lysed and GST-CaMKK α affinity purified using glutathione-Sepharose chromatography.

Plasmids

The GFP-AMPK- α 1, AMPK- β 1 and AMPK- γ 1 plasmids were described previously [25]. To make FLAG-tagged CaMKK β , a pGEX construct encoding human CaMKK β was digested with BamHI and XhoI and the insert cloned into the vector pCMV-Tag2B using standard methods.

Antibodies

The antibody against the phosphorylated form of Thr-172 on the AMPK α subunits was from Cell Signaling Technology. Antibodies against actin and the FLAG epitope, and sheep immunoglobulin were from Sigma, antibodies against green fluorescent protein (GFP) from Roche, against MARK3 (C-TAK1) from Millipore, and against AMPK- γ 1 from Abcam. Antibodies against AMPK- α 1 and - α 2 [26], LKB1 (N-terminal dephospho-Ser31 [24]), CaMKK β [13], AMPK-pan β [25], glutathione-S-transferase (GST) [27], and SIK1, SIK2, SIK3, NUA2, MARK1 and MARK4 [7] were as described previously.

Cell culture

HEK293 cells and the HeLa cells used in Figs. 6 and 7 were cultured in Dulbecco's modified Eagles's medium (DMEM) containing 10% (v/v) foetal bovine serum (FBS), 100 IU/ml penicillin and 100 µg/ml streptomycin. HeLa cells stably expressing wild type LKB1 and derived from T-REX™ HeLa cells (Invitrogen) were previously described [28]. The parental T-REX™ line was used in all experiments involving HeLa cells except Figs. 6 and 7, and was cultured in Eagle's Minimal Essential medium, 10% (v/v) FBS, 5 µg/ml blasticidin, but without tetracycline. Cells stably expressing LKB1 were grown in this medium with the addition of 100 µg/ml zeocin. The parental cell line was transfected with 10 µg (per 100 mm dish) FLAG-CaMKKβ using the polyethylenimine method [29]. Cells were transfected with combinations (3 µg each) of GFP-AMPK-α1, β1, γ1 and FLAG-CaMKKβ as indicated in the text using the polyethylenimine method [29]. The cells were cultured for a further 36 hr and were left untreated, or were treated with 10 µM A23187 for 1 hr. Cells were lysed in 0.5 ml of ice-cold lysis buffer (50 mM Tris/HCl, pH 7.2, 1 mM EGTA, 1 mM EDTA, 50 mM NaF, 1 mM sodium pyrophosphate, 1% (w/v) Triton X-100, 0.1 mM phenylmethane sulphonyl fluoride (PMSF), 1 mM DTT, 0.1 mM benzamidine and 5 µg/ml soybean trypsin inhibitor) after rapid rinsing in ice-cold PBS. The lysates were centrifuged (4°C; 10 min; 21000xg) and the supernatants frozen for later analysis.

Immunoprecipitation of AMPK and CaMKKβ

HEK-293 cell lysates (600 µg protein) were incubated at 4°C for 2 hr on a roller mixer with 5 µl of Protein G-Sepharose non-covalently conjugated to 5 µg of AMPK-α1/α2 or CaMKKβ antibodies. After extensive washing, the immunoprecipitates were heated in SDS sample buffer, resolved by SDS/PAGE and blots were probed with the indicated antibodies.

Immunoprecipitate kinase assays

Endogenous AMPK and the AMPK-related kinases from HeLa cells were assayed in immunoprecipitates as described in [24, 30] using the AMARA peptide [31] as substrate. GST-AMPK-α1, GST-BRSK1 and GST-BRSK2 were assayed as described previously [24]. For CaMKKα/β, assays were performed with or without 1 µM calmodulin and 1 mM CaCl₂ as indicated in the text.

Estimation of cellular ADP:ATP ratio

Culture medium was quickly aspirated from cells grown on 10 cm culture dishes and cells were washed with 1 ml of ice-cold PBS. After rapid aspiration of PBS, a minimal volume of ice cold 5% perchloric acid was added and the samples vortexed to ensure complete lysis. After centrifugation (14,000xg; 4°C; 3 min) to remove acid-insoluble material, the supernatant was extracted with two washes of an equal volume of 1:1 tri-n-octylamine and 1,1,2-trichlorotrifluoroethane. The nucleotides remaining in the aqueous phase were then separated by capillary electrophoresis with on-column isotachopheric concentration, using run buffers consisting of 50 mM Na phosphate, and 50 mM NaCl (pH 5.2; leading buffer) and 100 mM MES/Tris pH 5.2 (tailing buffer). To each buffer was added 0.2% hydroxyethylcellulose to decrease electro-osmotic flow. Nucleotide peaks were detected by UV absorbance at 260 nm, and integrated using System Gold software (Beckman). Nucleotide ratios were calculated from peak areas after correction for retention times. Identification of peaks as ATP, ADP and AMP were confirmed by additional runs spiked with internal standards, and analysis of absorbance spectra of individual peaks.

Statistics

Where multiple treatments were compared, 1- or 2-way analysis of variance (ANOVA) was used as indicated, with Bonferroni's post test of selected treatments, using GraphPad Prism software.

Other analytical procedures

SDS/PAGE was performed using precast Bis-Tris 4–12% gradient polyacrylamide gels in the MOPS buffer system (Invitrogen). Proteins were transferred to nitrocellulose membranes (BioRad) using the Xcell II Blot Module (Invitrogen). Membranes were blocked for 1 hr in TBS (Tris-buffered saline) containing 5% (w/v) non-fat dried skimmed milk. The membranes were probed with appropriate antibody (0.1–1 µg/ml) in TBS-Tween and 2% (w/v) non-fat dried skimmed milk. Detection was performed using secondary antibody (1 µg/ml) coupled to IR 680 or IR 800 dye, and the membranes scanned using the Li-Cor Odyssey IR imager. Protein concentrations were determined by Coomassie Blue binding [32] with BSA as standard.

RESULTS

Alignment of sequences around activation loop phosphorylation sites

Fig. 1 shows an alignment of the activation loops (lying between the conserved DFG and APE motifs) of the AMPK-related kinases, as well as the four isoforms of CaMKI (CaMKI- α , - β , - δ and - γ) and the single form of CaMKIV. All contain a threonine residue in the position equivalent to Thr-172 in AMPK- α 2, and a conserved hydrophobic residue (usually leucine) two residues N-terminal to this. Many of the other residues conserved between these sequences are also found in other protein kinases not downstream of LKB1 and/or CaMKKs, such as the DFG motif prior to the threonine, and the CG and YAAPE motifs following it. Using a peptide library approach, it has been shown that LKB1 only phosphorylates threonine residues, and strongly prefers a leucine two residues N-terminal to the threonine (i.e. at P-2) [33]. Fig. 1 shows that leucine is present at P-2 in all AMPK-related kinases and in CaMK1- α and - β , although in CaMK1- δ , CaMKI- γ and CaMKIV, the residue at P-2 is methionine. The features of primary sequence recognized by CaMKK α and CaMKK β , if any, are not known.

CaMKKs activate AMPK but not AMPK-related kinases in HeLa cells

As an initial test of whether CaMKKs would activate any of the AMPK-related kinases we utilized HeLa cells, which express both CaMKK α and CaMKK β [13], but do not express LKB1 due to a biallelic deletion of the gene in the cervical cancer from which the cells were originally derived in 1951 [34]. We have shown previously that the Ca²⁺ ionophore A23187 caused activation of endogenous AMPK in HeLa cells, and that this effect was blocked by the CaMKK inhibitor, STO-609 [13]. Activation of AMPK (Fig. 2A), and phosphorylation of Thr-172 (not shown), occurred in T-RexTM HeLa cells not only in response to A23187, but also in response to stable expression of wild type, active LKB1. However, while stable expression of LKB1 in T-RexTM HeLa cells increased the basal activity of the endogenous AMPK-related kinases SIK1, SIK2, SIK3, NUA2, MARK1, MARK2/MARK3 and MARK4, unlike AMPK none were significantly activated by A23187 in the parental cells (Fig. 2B-H) (for these experiments, MARK2 and MARK3 were assayed together because we do not have an antibody that distinguishes these two closely related kinases [7]). These results show that the CaMKKs do not activate any of the endogenous AMPK-related kinases that are expressed in HeLa cells.

CaMKK β is activated by Ca²⁺ and calmodulin but also has a substantial basal activity [13]. We therefore examined the effect of over-expressing CaMKK β in HeLa cells in the absence of a Ca²⁺ ionophore. Fig. 3A shows that transient expression of CaMKK β in the parental T-Rex™ HeLa cells caused almost as much activation of AMPK as stable expression of LKB1 in the same cell line, even without addition of A23187. Based on the signal obtained in a Western blot using anti-FLAG antibody (both over-expressed proteins being FLAG-tagged), the expression of CaMKK β in these experiments was somewhat higher than that of LKB1 (Fig. 3B). Despite the large activation of AMPK by over-expression of CaMKK β , none of the AMPK-related kinases examined (SIK1, SIK2, SIK3 or NUA2) were activated compared with the untransfected cells, although they were activated by stable expression of LKB1 (Fig. 3C-F).

CaMKK β activates the AMPK kinase domain but not BRSK1/BRSK2 in cell-free assays

The brain-specific AMPK-related kinases, BRSK1 and BRSK2, are not expressed in HeLa cells. In order to address whether they could be phosphorylated and activated by CaMKK β , we expressed their combined kinase and UBA-like domains in bacteria as glutathione-S-transferase (GST) fusions, as well as the kinase domain of the α 1 isoform of the AMPK catalytic subunit (GST- α 1-KD). We also expressed GST-CaMKK β in bacteria and a GST-LKB1:FLAG-STRAD α :myc-MO25 α complex in HEK-293 cells. All fusion proteins/complexes were purified on glutathione-Sepharose and the upstream kinases were tested for their ability to phosphorylate and activate GST- α 1-KD, GST-BRSK1 and GST-BRSK2 in cell-free assays. Fig. 4A shows that while the LKB1 complex and CaMKK β were both able to activate GST- α 1-KD, only the LKB1 complex was able to activate GST-BRSK1 and GST-BRSK2. There was also no phosphorylation of BRSK1 or BRSK2 by CaMKK β under conditions where there was a robust phosphorylation of AMPK (not shown). These experiments were performed in the absence of Ca²⁺ and calmodulin, since in cell-free assays CaMKK β appears to be at least partially independent of Ca²⁺ and calmodulin [13]. However, Fig. 4B confirms that BRSK1 was not phosphorylated by CaMKK β even in the presence of Ca²⁺ and calmodulin.

Interestingly, although CaMKK β was ineffective, CaMKK α was able to activate (Fig. 4C) and phosphorylate (Fig. 4D) GST-BRSK1 in the presence of Ca²⁺ and calmodulin. However, the concentration of CaMKK α used in these experiments was about 20-fold higher than that required to obtain a comparable level of phosphorylation of AMPK- α 1 (data not shown).

AMPK and CaMKK β do not form a stable complex

Anderson *et al* [23] have recently reported evidence suggesting that CaMKK β can form a stable complex with the α and β subunits of AMPK in the absence of a γ subunit. To test this hypothesis, we initially studied whether over-expressed CaMKK β would co-precipitate with endogenous AMPK from extracts of HEK-293 cells. Fig. 5A shows that, although CaMKK β was readily detectable by Western blotting in the crude cell lysates, none was detected by Western blotting in immunoprecipitates made using anti-AMPK- α 1/ α 2 antibodies (although we could readily detect the α , β and γ subunits of AMPK). Anderson *et al* [23] claimed that the interaction between CaMKK β and AMPK was Ca²⁺-dependent, but the same results were obtained regardless of whether EDTA and EGTA were present in the lysis and immunoprecipitation buffers. When the converse experiment was performed, *i.e.* by immunoprecipitating using an anti-CaMKK β antibody, we could detect CaMKK β but not the α , β or γ subunits of AMPK in the precipitate, irrespective of the presence of EDTA and EGTA in the buffers (Fig. 5B).

Fig. 5C shows results of expressing GFP-tagged α 1 with or without various combinations of β 1, γ 1 and FLAG-tagged CaMKK β in HeLa cells. It has been previously shown that co-expression

with β and γ subunits greatly increases expression of GST-tagged $\alpha 1$ in COS7 cells, most likely because the heterotrimeric complex is more stable and is degraded less rapidly [35]. Consistent with this, co-expression of GFP- $\alpha 1$ with $\beta 1$ and $\gamma 1$, but not $\beta 1$ alone, led to a 3- to 4-fold increase in its expression; co-expression with $\gamma 1$ together with GFP- $\alpha 1$ also increased the expression of $\beta 1$. When CaMKK β was expressed, the protein was expressed at equal levels irrespective of whether or not particular AMPK subunits had been co-expressed. If, as suggested by Anderson *et al* [23], CaMKK β forms a complex with AMPK- α and - β subunits in the absence of γ , one might have expected that co-expression of CaMKK β would have stabilized and increased the expression of GFP- $\alpha 1$ and $\beta 1$ in the absence of $\gamma 1$. In fact, the opposite was observed: when GFP- $\alpha 1$ and $\beta 1$ were co-expressed with CaMKK β their expression level was reduced (compare lane 7 with lane 3 in Fig. 5C). Only when GFP- $\alpha 1$ and $\beta 1$ were co-expressed with $\gamma 1$ was there a marked increase in their expression.

Increasing Ca^{2+} and AMP causes a synergistic activation of AMPK in HeLa cells

Although it had been proposed that AMP promoted phosphorylation of AMPK by LKB1 [5], recent studies suggest that this may have been caused by trace contamination of the kinase preparations used with protein phosphatase, and that the effect of AMP on Thr-172 phosphorylation is entirely via inhibition of dephosphorylation [10]. This raised the question as to why drugs such as AICAR, which is converted to the AMP mimetic ZMP [36], or phenformin, which increases cellular AMP [5], do not activate AMPK in HeLa cells [5], even though CaMKK β is expressed and is capable of phosphorylating Thr-172 in those cells [13]. One explanation was that the activity of CaMKK β in cells with basal Ca^{2+} is so low that the endogenous protein phosphatase activity is sufficient to cause net dephosphorylation of Thr-172, even though dephosphorylation is being inhibited by AMP. To test this idea, we incubated HeLa cells with increasing concentrations of the Ca^{2+} ionophore A23187 in the presence and absence of phenformin. As found previously [5], in the absence of A23187 phenformin did not give a significant stimulation of AMPK activity, phosphorylation of Thr-172 on the AMPK α subunits, or phosphorylation of the AMPK site on acetyl-CoA carboxylase (ACC). As expected [13], in the absence of phenformin concentrations of A23187 up to 10 μM increasingly stimulated AMPK activity (Fig. 6A), as well as phosphorylation of AMPK and ACC (Fig. 6B). However, Fig. 6 reveals the new result that the presence of phenformin sensitized the cells to A23187. Thus, the presence of phenformin shifted the curve of AMPK phosphorylation and activation versus A23187 concentration to the left. A stimulation of the activation of AMPK, and the phosphorylation of AMPK and ACC, by phenformin was particularly evident at 3 μM A23187. These results show that, under appropriate conditions, phenformin can activate AMPK even in cells lacking LKB1.

We repeated the experiments using 3 μM A23187 in the presence and absence of phenformin or 2-deoxyglucose, another agent that can deplete cellular ATP, and measured AMPK activity, phosphorylation of ACC and AMPK, and levels of cellular nucleotides (Fig. 7). The level of AMP in HeLa cells under basal conditions was too low to measure accurately, although it could be detected in cells treated with phenformin and A23187 (not shown). We therefore used ADP:ATP as a surrogate for AMP:ATP, since the latter is expected to vary as the square of ADP:ATP [37]. Phenformin and 2-deoxyglucose did not cause phosphorylation of ACC or AMPK, or activation of AMPK, in HeLa cells that had not been treated with a Ca^{2+} ionophore, despite the fact that 2-deoxyglucose produced a large rise in ADP:ATP ratio (any small effect of phenformin on the ADP:ATP ratio was not significant). However, in cells treated with 3 μM A23187, both phenformin and 2-deoxyglucose significantly stimulated AMPK activity, and increased phosphorylation of ACC and AMPK, over and above the effect of A23187 alone, with the effect of phenformin being larger than that of 2-deoxyglucose (Fig. 7A, 7B). The increase in ADP:ATP induced by 2-deoxyglucose was not affected by A23187 but, surprisingly, a much larger increase in ADP:ATP occurred in the presence of A23187

in cells treated with phenformin (Fig. 7C). Despite the contrasting effects of phenformin and 2-deoxyglucose on cellular ADP:ATP ratios, the results in Fig. 7 confirm that, under appropriate conditions, both agents are capable of activating AMPK even in cells completely lacking LKB1.

Discussion

Despite the close similarity in the sequences surrounding Thr-172 on AMPK and the equivalent threonine residue on the AMPK-related kinases (Fig. 1), it is clear that the latter are not phosphorylated significantly by CaMKKs either in intact cells (Figs. 2-3) or in cell-free assays (Fig. 4). We have shown previously that HeLa cells lack LKB1 but express both CaMKK α and CaMKK β , although it was only when CaMKK β was knocked down using siRNA that the phosphorylation and activation of AMPK in response to Ca²⁺ ionophores was reduced [13]. Thus it seems that neither CaMKK α nor CaMKK β are capable of activating any of the AMPK-related kinases that are expressed in HeLa cells, i.e. SIK1 (SIK), SIK2 (QIK), SIK3 (QSK), NUAK2 (SNARK), or any of the MARKs. Similarly, over-expression of CaMKK β caused activation of AMPK in HeLa cells but failed to activate any of the AMPK-related kinases tested (Fig. 3).

BRSK1 (SAD-B) and BRSK2 (SAD-A) are brain-specific kinases that are not expressed in HeLa cells. Fujimoto et al [38] recently reported a search for proteins from rat brain extracts that interacted with the kinase domain of CaMKK α , and detected both the $\alpha 1$ subunit of AMPK and BRSK1 (SAD-B). They also reported that BRSK1 could be phosphorylated (at Thr-189, equivalent to Thr-172 on AMPK- $\alpha 1$) and activated by both CaMKK α and CaMKK β , although the rate of phosphorylation was much lower with CaMKK β than CaMKK α . The results in Fig. 4 confirm that CaMKK α is capable of phosphorylating and activating BRSK1 in cell-free assays. However, in our studies neither BRSK1 nor BRSK2 were significantly phosphorylated by CaMKK β , although the concentration used was sufficient to fully activate AMPK. It should also be pointed out that the concentration of CaMKK α used to phosphorylate BRSK1 in Fig. 4 was about 20-fold higher than that required to obtain a comparable level of phosphorylation of AMPK- $\alpha 1$ (data not shown). In addition, AMPK is itself a relatively poor substrate for CaMKK α , with CaMKK α being about 7-fold less active against AMPK than CaMKK β when the activities of the two CaMKKs were normalized using phosphorylation of calmodulin-dependent protein kinase I [13]. Thus, BRSK1 appears to be a poor substrate for CaMKK α and an even poorer substrate for CaMKK β , and we propose that LKB1 is the major upstream kinase for BRSK1 and BRSK2 *in vivo*. In support of this are results of Barnes et al [39], who knocked out LKB1 in pyramidal neurones of the cerebral cortex by crossing mice harbouring a conditional "floxed" allele with mice expressing Cre recombinase from the Emx1 promoter. This almost abolished phosphorylation of BRSK2 (SAD-A) and BRSK1 (SAD-B) at the threonine residues equivalent to Thr-172, while not affecting phosphorylation of AMPK at Thr-172. This finding can be explained if CaMKK β (or CaMKK α) provides an alternate upstream pathway for activation of AMPK, but not BRSK1 and BRSK2, in the cerebral cortex. Also in agreement with our results, Bright et al [40] reported that neither BRSK1 nor BRSK2 were activated in CCL13 cells over-expressing CaMKK β , while Uboha et al [41] reported that CaMKK partially purified from mouse cortex (isoform not specified) failed to phosphorylate MARK2 in cell-free assays.

Given that the sequences around the threonine in the activation loop are highly conserved between AMPK and the AMPK-related kinases, it remains unclear why the latter should be phosphorylated and activated by LKB1 complexes but not by CaMKK- α or - β . Our findings suggest that some feature of the structure of the AMPK-related kinases, other than the primary sequence of the activation loop, may be required to determine this specificity. This is supported by previous studies showing that a peptide derived from CaMKIV was a very poor substrate for CaMKK α compared with CaMKIV itself [42], and that thermal denaturation of CaMKI or CaMKIV abolished their

phosphorylation by CaMKK α [42, 43]. Thus, the CaMKKs appear to recognize determinants on CaMKI and CaMKIV other than just the primary sequence of the activation loop.

Our findings have important implications regarding the regulation of the AMPK-related kinases. None of them appear to interact with the regulatory β or γ subunits of the AMPK complex [44] and, consistent with this, those that are expressed in skeletal muscle (SIK2, SIK3, MARK2/3 and MARK4) are not phosphorylated or activated when AMPK is activated in muscle by contraction, AICAR or phenformin [8]. Our results now show that the AMPK-related kinases, unlike AMPK itself, are not regulated by increases in cytosolic Ca²⁺ via activation of CaMKK. That being the case, what are the upstream signals that regulate the AMPK-related kinases? In most cases this remains unclear, although there is some information available in the case of SIK1. Salt-inducible kinase (SIK1) was discovered as a protein kinase induced in the adrenal glands of rats fed a high-salt diet [45], and is also induced at the transcriptional level by glucagon and forskolin in primary rat hepatocytes [46]. In Y1 mouse adrenocortical tumour cells treated with adrenocorticotrophic hormone, SIK1 is phosphorylated at a C-terminal site by PKA, resulting in its translocation to the cytoplasm [47]. Thus, SIK1 may be regulated by altered transcription and/or by altered subcellular localization rather than by altered phosphorylation by LKB1.

Anderson et al [23] recently provided evidence suggesting that CaMKK β (also known as CaMKK2) can form a stable complex with the AMPK α and β subunits. They expressed FLAG-tagged CaMKK β in HEK-293 cells and, using anti-FLAG antibodies, showed co-precipitation not only of calmodulin but also of AMPK α subunits and acetyl-CoA carboxylase (ACC), the latter two detected using phosphospecific antibodies against Thr-172 (AMPK) and Ser-79 (ACC). These co-precipitation events appeared to be dependent on Ca²⁺ because they were not observed in the presence of EDTA and EGTA. Anderson et al also detected in the anti-FLAG immunoprecipitates the α 1, α 2, β 1 and β 2 subunits of AMPK, but not γ 1, although they could detect the latter in cell extracts. They therefore proposed that CaMKK β could replace the γ subunit within the AMPK complex. Our results do not support this concept. When CaMKK β was over-expressed in HEK-293 cells, we could not detect any AMPK- α , - β or - γ subunits in an immunoprecipitate made using anti-CaMKK β antibodies, either with or without EDTA and EGTA in the lysis and immunoprecipitation buffers (Fig. 5B). In the converse experiment (which was not reported by Anderson et al) we found that no CaMKK β was detected in an anti-AMPK- α 1/ α 2 immunoprecipitate, although we could readily detect the α , β and γ subunits of AMPK (Fig. 5A). This suggests to us that the AMPK subunits recovered by Anderson et al in their anti-FLAG immunoprecipitates may have been an artefact caused by the high level of FLAG-CaMKK β expression in the cells.

We also addressed the same question in a different way, making use of previous findings that the α subunits of AMPK are unstable unless co-expressed with β and γ subunits [35]. This phenomenon can be explained by recent structural studies showing that the C-terminal region of the β subunit forms extensive interactions with the C-terminal domain of α and with the N-terminal region of γ [4]. If these subunits do not find their interaction partners, they may be recognized as misfolded proteins and become degraded. We argued that, if the AMPK- α and - β subunits can form a complex with CaMKK β in the absence of the γ subunit, then co-expression with CaMKK β should stabilize the α subunit in the presence of a β subunit. However, Fig. 5C shows that this is not the case: in the presence of β 1 the γ 1 subunit stabilizes the α 1 subunit, but CaMKK β does not substitute for γ 1 in this respect. Rather than stabilizing the AMPK α and β subunits, over-expression of CaMKK β decreased the level of expression of all three AMPK subunits (Fig. 5C), perhaps because the transfected DNA was competing for the available biosynthetic machinery.

Finally, the results shown in Figs. 6 and 7 show that the two known mechanisms by which phosphorylation of AMPK is activated, i.e. inhibition of dephosphorylation by AMP and increased

phosphorylation via activation of CaMKK β by Ca²⁺, can under the right circumstances be synergistic. They also provide an explanation for our previous findings [5] that agents that cause increases in AMP (e.g. phenformin) or that are converted to an AMP mimetic (AICAR) do not increase phosphorylation and activation of AMPK in HeLa cells. The basal rate of phosphorylation of Thr-172 in these cells, which lack LKB1 and would only have the Ca²⁺-independent activity of CaMKK β , appears to be too low for any effect of increased AMP on dephosphorylation to become evident. However, if CaMKK β is activated using the Ca²⁺ ionophore A23187 at sub-optimal concentration (3 μ M), the effect of a rise in AMP to inhibit dephosphorylation now becomes apparent (Figs. 6 and 7). In agreement with our findings, Yu *et al* [48] have recently reported that A23187 and AICAR act in an additive manner to inhibit cell proliferation and/or survival in HeLa cells.

A somewhat unexpected finding shown in Fig. 7 was that while treatment with 3 μ M A23187 alone did not significantly increase the ADP:ATP ratio, when combined with 10 mM phenformin it caused a much larger increase in this ratio than that caused by phenformin alone. By contrast, 2-deoxyglucose treatment increased ADP:ATP but there was no additional effect when it was added together with A23187. Phenformin is an inhibitor of complex I of the respiratory chain [49, 50], thus potentially causing inhibition of mitochondrial ATP synthesis, whereas A23187 is a Ca²⁺ ionophore that causes release of cytosolic Ca²⁺, potentially increasing ATP turnover due to activation of Ca²⁺ pumps. Although 2-deoxyglucose is often described as an inhibitor of glycolysis, its major effect may be to accelerate ATP consumption, because it is rapidly phosphorylated within the cell to 2-deoxyglucose-6-phosphate. We propose that a combination of treatments that both inhibit ATP synthesis and accelerate ATP turnover (phenformin and A23187), as opposed to two different treatments that accelerate ATP turnover (2-deoxyglucose and A23187) may make it more difficult for the cells to maintain their energy status. The very large increase in AMPK activity in response to addition of both A23187 and phenformin (Fig. 7A) could be explained by an increase in cytosolic Ca²⁺ coupled with a larger increase in AMP than that caused by 2-deoxyglucose. Whatever the explanation, the results in Fig. 7 show that treatments that elevate cellular AMP (phenformin and 2-deoxyglucose) can under the right circumstances (elevated cytosolic Ca²⁺) cause increased phosphorylation and activation of AMPK even in cells lacking LKB1. Thus, the effect of AMP on phosphorylation of AMPK is independent of the upstream kinase utilized.

Increased cytosolic Ca²⁺ tends to switch on many processes that consume ATP, such as operation of ion pumps or channels, interaction of motor proteins, and membrane trafficking and secretion. The ability of Ca²⁺ and AMP to additively activate AMPK to restore cellular energy balance may therefore have physiological relevance.

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FIGURE LEGENDS

- Figure 1:** Alignment of activation loops of the AMPK-related kinases and calmodulin-dependent protein kinases. Sequences between the conserved DFG and YAAPE motifs were manually aligned, with gaps indicated by dots. The arrow shows the threonine residue (P) whose phosphorylation is required for kinase activity. Residues identical between all sequences are boxed, while the conserved hydrophobic residue at P-2 is in bold type.
- Figure 2:** Effect of treatment of T-Rex™ HeLa cells with A23187, or stable expression of LKB1 in the same cells, on the activity of endogenous AMPK and AMPK-related kinases. The indicated kinase was immunoprecipitated from cell lysates and assayed using the AMARA peptide as substrate. Note that we do not have an antibody that distinguishes MARK2 and MARK3 so the activity measured with this antibody could represent either MARK2 or MARK3, or a combination of the two. Data are means ± SD for duplicate cell incubations; mean significantly different from control without A23187 or LKB1, $p < 0.05^*$; $p < 0.01^{**}$; $p < 0.001^{***}$.
- Figure 3:** Effect of transient expression in T-Rex™ HeLa cells of CaMKKβ, or stable expression of LKB1, on the phosphorylation and activity of AMPK and AMPK-related kinases. (A) endogenous AMPK activity; (B) Western blots of duplicate cell lysates shown in (A); (C) – (F) activities of endogenous SIK1, SIK2, SIK3 and NUA2. Data are means ± SD for duplicate cell incubations; mean significantly different from control without CaMKKβ or LKB1, $p < 0.05^*$; $p < 0.01^{**}$; $p < 0.001^{***}$.
- Figure 4:** Phosphorylation and activation of GST-AMPK-α1 (kinase domain only), GST-BRSK1 or GST-BRSK2 (kinase domains plus UBA-like domains) by CaMKKs. (A) Activity of GST-AMPK-α1, -BRSK1 or -BRSK2 after incubation in the presence of MgATP for 1 hr with or without CaMKKβ (minus Ca²⁺/calmodulin), or LKB1:STRADα:MO25α. (B) Time course of phosphorylation of GST-BRSK1 by CaMKKβ plus Ca²⁺/calmodulin, or LKB1:STRADα:MO25α. (C) Activity of AMPK-α1 and BRSK1 after incubation in the presence of MgATP for 1 hr with or without CaMKKα plus Ca²⁺/calmodulin, or LKB1:STRADα:MO25α. (D) Time course of phosphorylation of GST-BRSK1 in the experiment shown in (C). Data are means ± SD for duplicate cell incubations; means significantly different from control without CaMKKβ or LKB1, $p < 0.001^{***}$.

Figure 5: AMPK and CaMKK β do not form a stable complex. (A) CaMKK β was over-expressed in HEK-293 cells; the top panel show Western blots of CaMKK β and actin in the cell lysates, while the bottom panel shows Western blots performed on immunoprecipitates made with control immunoglobulin or anti-AMPK- α 1/ α 2 antibodies in the presence and absence of EDTA plus EGTA. (B) As (A), except that Western blots were performed on immunoprecipitates made with control immunoglobulin or anti-CaMKK β antibodies in the presence and absence of EDTA plus EGTA. (C) HeLa cells were transfected with various combinations of plasmids encoding GFP-tagged AMPK- α 1, AMPK- β 1, AMPK- γ 1 or CaMKK β . Lysates were analyzed by Western blotting using antibodies against GFP, pan- β , γ 1, CaMKK β or actin.

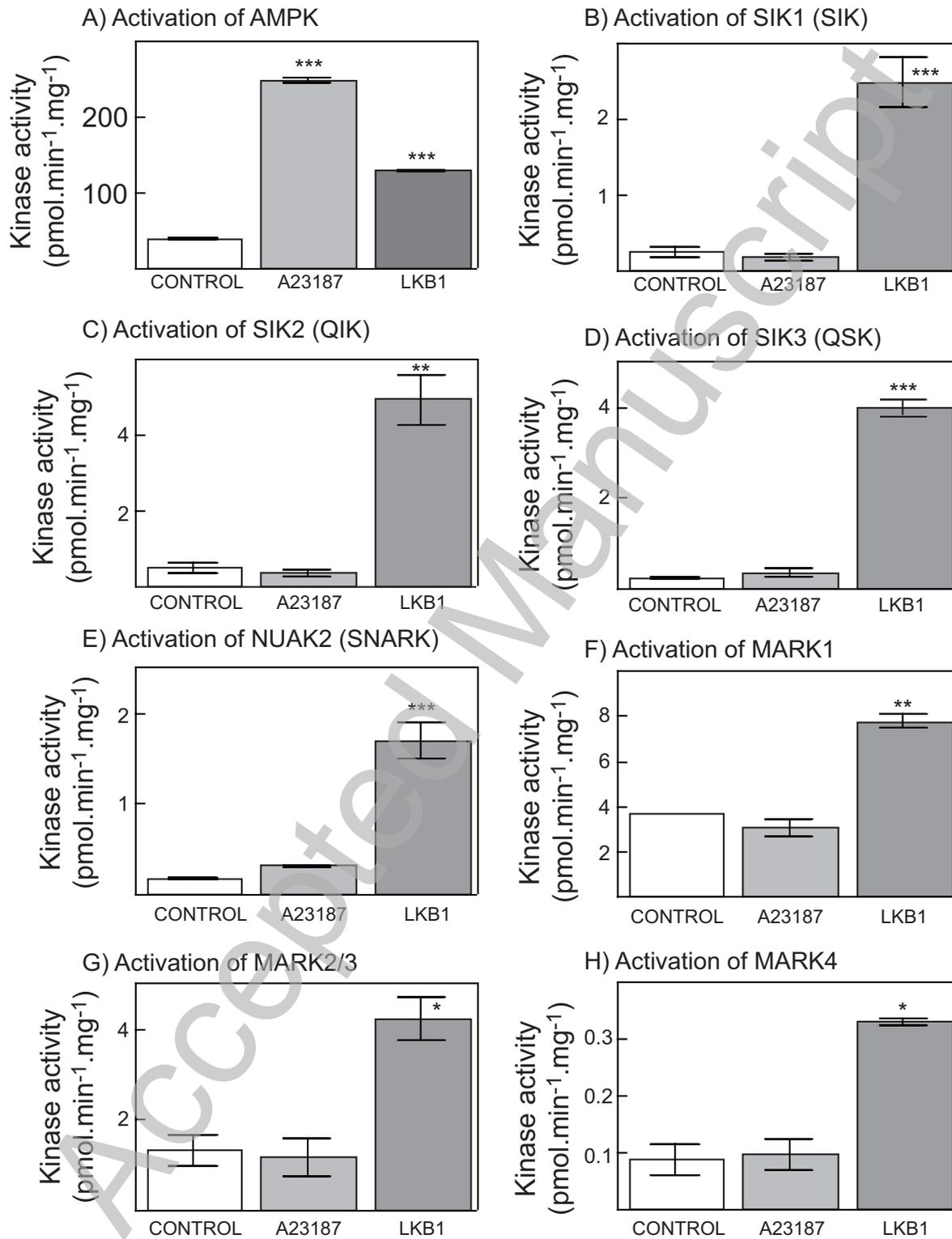
Figure 6: In cells lacking LKB1, phenformin increases activation and phosphorylation by the Ca²⁺ ionophore A23187. (A) HeLa cells were treated with increasing concentrations of A23187 in the presence and absence of 10 mM phenformin. AMPK was immunoprecipitated and assayed at 200 μ M AMP. Results are mean \pm range of duplicate cell incubations. (B) Cell lysates treated as in (A) were analyzed by Western blotting using antibodies against the phosphorylated form of ACC (pACC), streptavidin to detect total ACC, phosphospecific antibodies against the Thr-172 site on AMPK- α (pT172) and antibodies against total AMPK- α 1/ α 2 (AMPK- α).

Figure 7: Phenformin and 2-deoxyglucose cause increased phosphorylation and activation of AMPK in HeLa cells treated with low concentrations of Ca²⁺ ionophore. (A) HeLa cells were treated with or without 3 μ M A23187 in the presence and absence of 10 mM phenformin or 100 μ M 2-deoxyglucose (2DG) for 1 hr. AMPK activity was determined in immunoprecipitates, and results are mean \pm SD (n = 3). (B) Samples from the experiment in (A) were analyzed in duplicate by Western blotting using antibodies as in Fig. 6C. (C) ADP:ATP ratios in an experiment identical to that shown in (A). Cells were extracted with perchloric acid and acid-free extracts analyzed by capillary electrophoresis. ADP:ATP ratios were determined and results are expressed as mean \pm SD (n = 3). KEY: ***significantly different by 1-way ANOVA (p<0.001) from relevant control without phenformin or 2DG; †††significantly different by 1-way ANOVA (p<0.001) from control without A23187.

Fogarty et al Figure 1

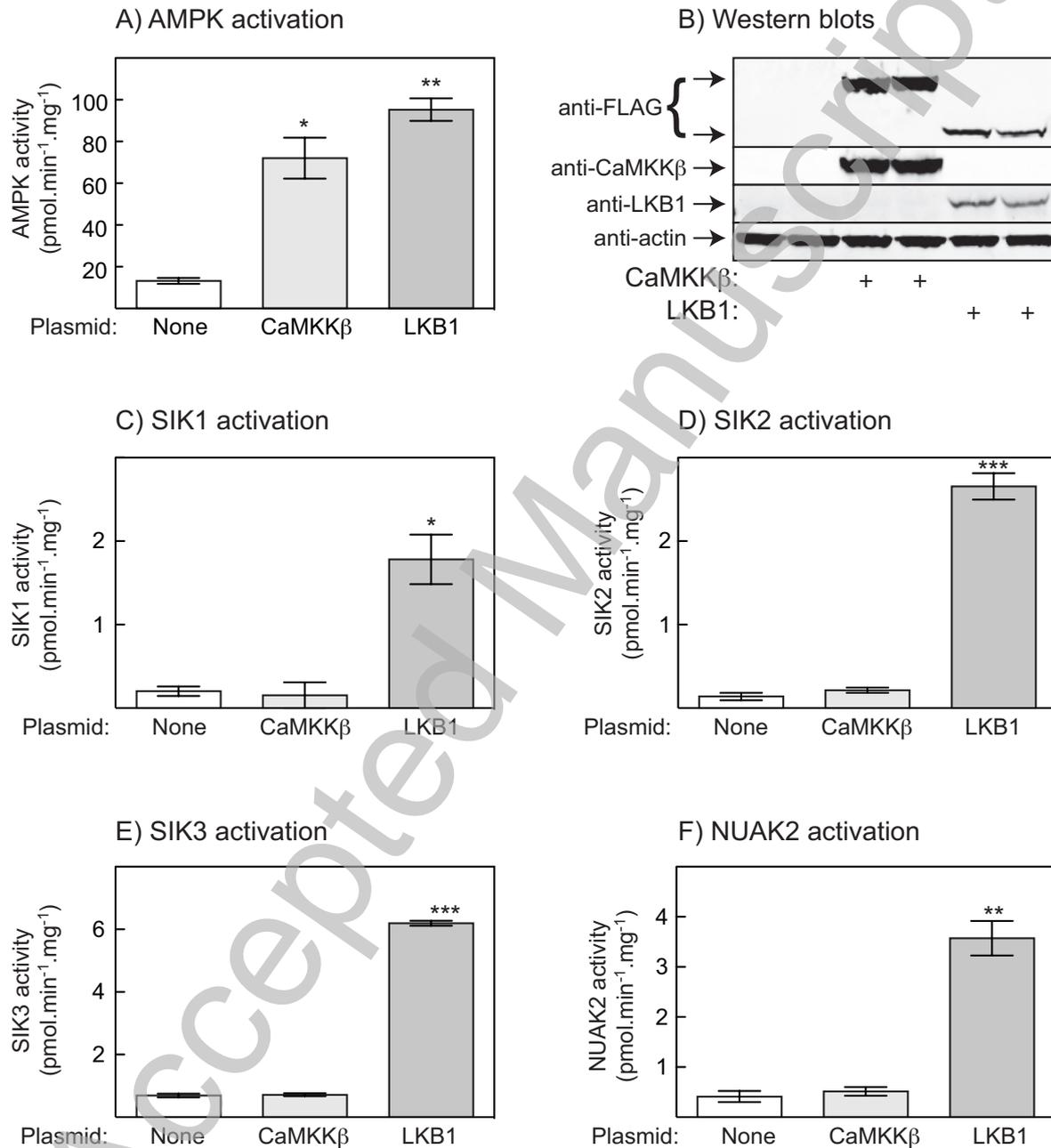
AMPK- α 1 (159-185):	DFGLSNMMSDGEFL LR TS	SCGSPNYAAPE
AMPK- α 2 (157-183):	DFGLSNMMSDGEFL LR TS	SCGSPNYAAPE
BRSK1 (190-216):	DFGMASLQVGDSL LE TS	SCGSPHYACPE
BRSK2 (159-185):	DFGMASLQVGDSL LE TS	SCGSPHYACPE
MARK1 (200-226):	DFGFSNEFTVGNK LD TF	CGSPPYAAPE
MARK2 (193-219):	DFGFSNEFTVGNK LD TF	CGSPPYAAPE
MARK3 (219-245):	DFGFSNEFTVGGK LD TF	CGSPPYAAPE
MARK4 (199-225):	DFGFSNEFTLGSK LD TF	CGSPPYAAPE
NUAK1 (196-222):	DFGLSNLYQKDK FL QT	FCGSPLYASPE
NUAK2 (1913-219):	DFGLSNLYHQGK FL QT	FCGSPLYASPE
SIK1 (167-193):	DFGFGNFYKSGEP LS TW	CGSPPYAAPE
SIK2 (160-186):	DFGFGNFFKSGEL LA TW	CGSPPYAAPE
SIK3 (206-232):	DFGFSNLFTPGQL LK TW	CGSPPYAAPE
SNRK (158-184):	DFGFSNKFQPGK LT TS	CGSLAYSAPPE
CaMKI α (162-188):	DFGLSKMEDPGSV LS TAC	GTPGYVAPE
CaMKI β (157-182):	DFGLSKIQ.AGN ML GTAC	GTPGYVAPE
CaMKI δ (165-191):	DFGLSKMEGKGDV MS TAC	GTPGYVAPE
CaMKI γ (164-189):	DFGLSKME.QNG IM STAC	GTPGYVAPE
CaMKIV (185-211):	DFGLSKIVEHQV LM KTV	CGTPGYCAPE

Fogarty et al Figure 2



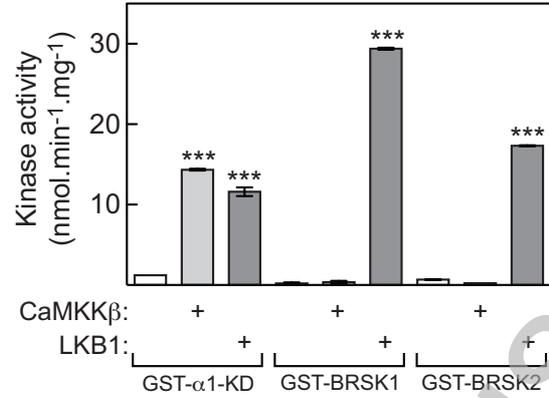
THIS IS NOT THE VERSION OF RECORD - see doi:10.1042/BJ20091372

Fogarty et al Figure 3

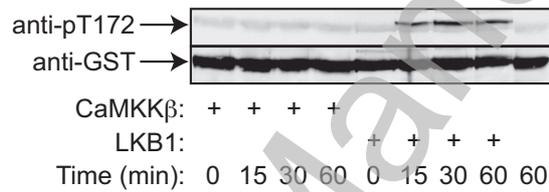


Fogarty et al Figure 4

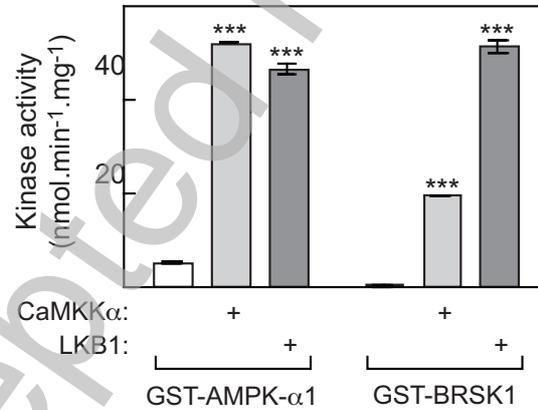
A) Activation of AMPK, BRSKs by CaMKK β /LKB1



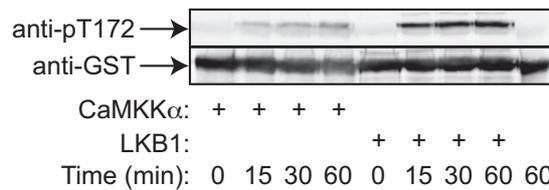
B) Phosphorylation of BRSK1 by CaMKK β /LKB1



C) Activation of BRSK1 by CaMKK α /LKB1



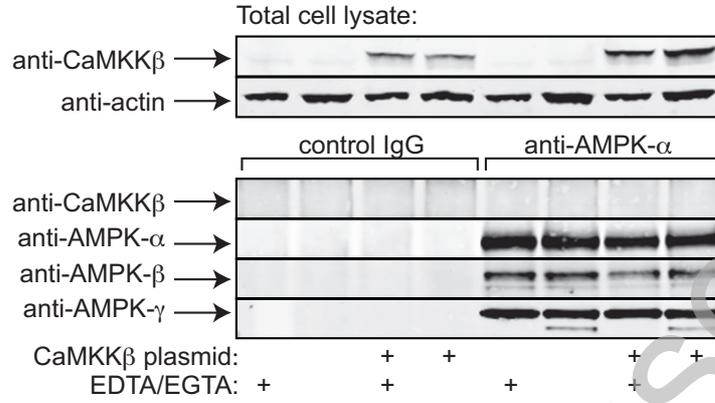
D) Phosphorylation of BRSK1 by CaMKK α /LKB1



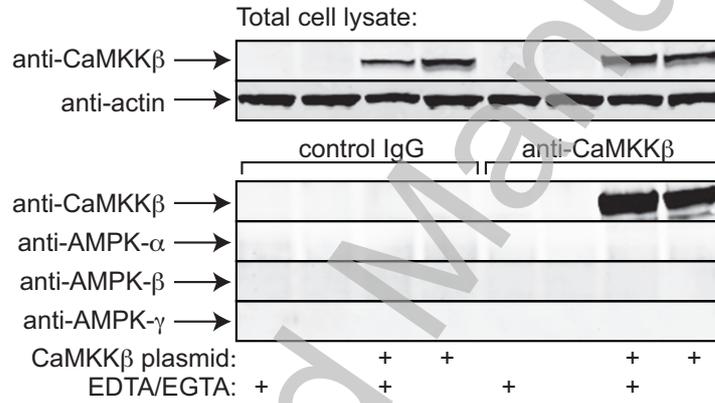
THIS IS NOT THE VERSION OF RECORD - see doi:10.1042/BJ20091372

Fogarty et al Figure 5

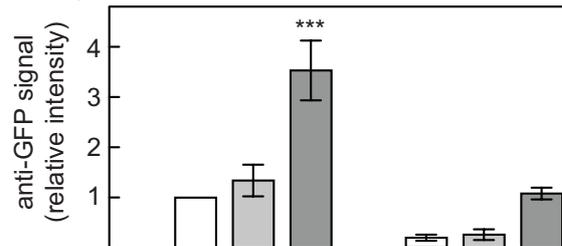
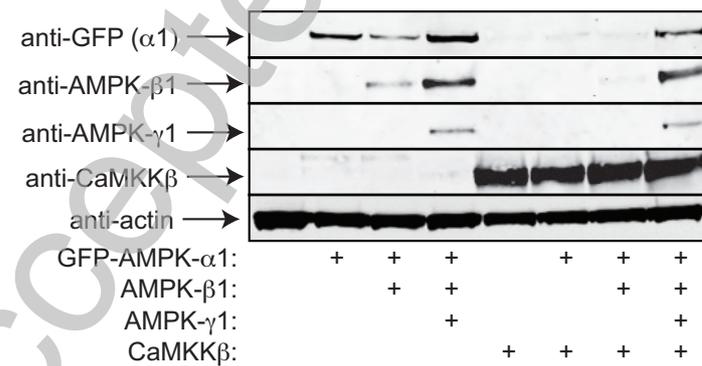
A) CaMKK β does not co-precipitate with AMPK antibodies



B) AMPK does not co-precipitate with CaMKK β antibodies



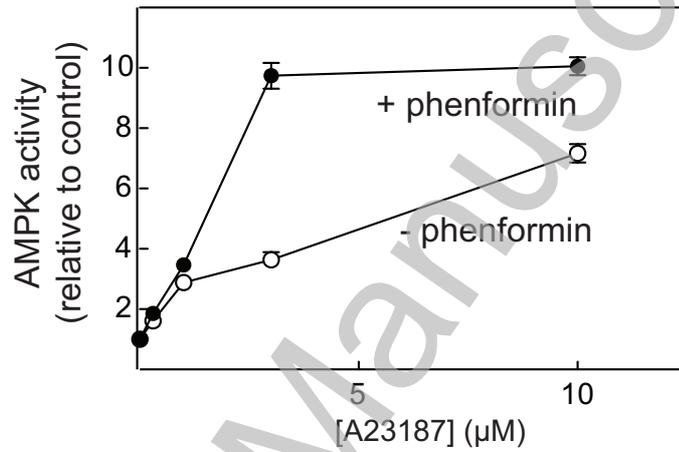
C) Stabilization of GFP- α 1 by β 1/ γ 1, but not by CaMKK β



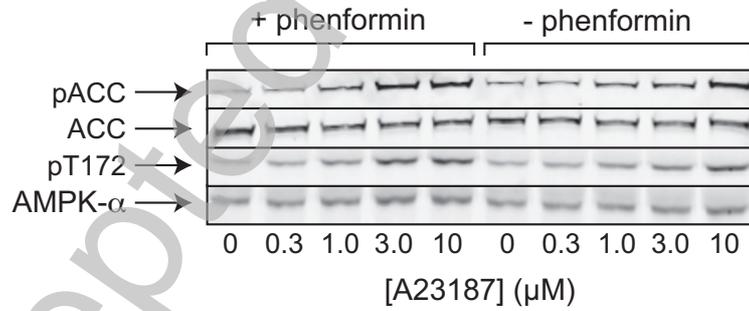
THIS IS NOT THE VERSION OF RECORD - see doi:10.1042/BJ20091372

Fogarty et al Figure 6

A) AMPK activation by A23187 ± phenformin (HeLa cells)



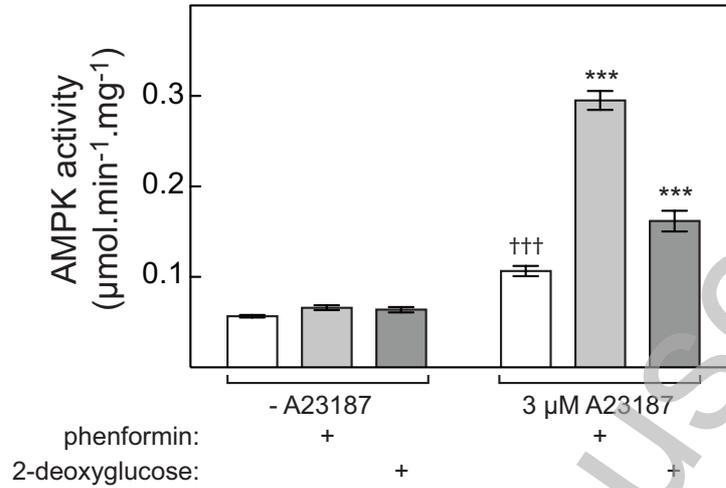
B) Phosphorylation by A23187 ± phenformin



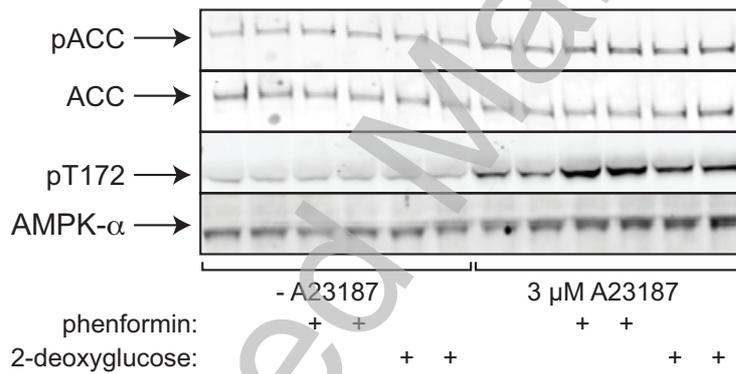
THIS IS NOT THE VERSION OF RECORD - see doi:10.1042/BJ20091372

Fogarty et al Figure 7

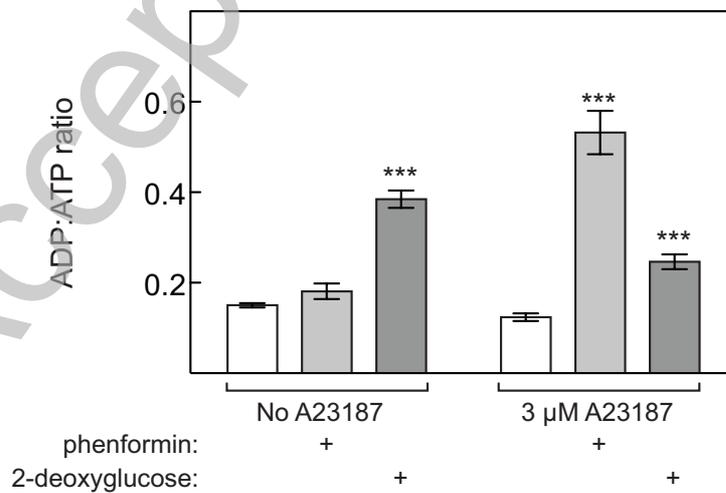
A) AMPK activation by phenformin or 2-DG ± A23187



B) Effects on phosphorylation of ACC and AMPK



C) Effects on cellular ADP:ATP ratio



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