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Marie Jarry, Mickael Diallo, Céline Lecointre, Laurence Desrues, Tursonjan Tokay, et al.. The vasoactive peptides urotensin II and urotensin II-related peptide regulate astrocyte activity through common and distinct mechanisms. Involvement in cell proliferation. *Biochemical Journal*, 2010, 428 (1), pp.113-124. 10.1042/BJ20090867. hal-00479207

HAL Id: hal-00479207

<https://hal.science/hal-00479207v1>

Submitted on 30 Apr 2010

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The vasoactive peptides Urotensin II and Urotensin II-related peptide regulate astrocyte activity through common and distinct mechanisms. Involvement in cell proliferation

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Short title: Differential effects of URP and UII on astrocyte proliferation

Keywords: UT receptor, calcium, proliferation, G protein, BRET

Abbreviations used

2-APB, 2-aminoethoxydiphenylborane; BRET, Bioluminescence Resonance Energy Transfer; $[Ca^{2+}]_c$ cytosolic calcium concentration; ω -CgTx GVIA, ω -conotoxin GVIA; DBI, diazepam-binding inhibitor; DMEM, Dulbecco's modified Eagle's medium; EGTA, ethylene glycol-bis(2-aminoethylether)-*N,N,N',N'*-tetraacetic acid; ET-1, endothelin-1; FBS, fetal bovine serum; GPCR, G protein-coupled receptor; IL-1 β , interleukin-1 β ; ODN, octadecaneuropeptide; PAR, protease-activated receptor; PBS, phosphate buffered saline; PIP, polyphosphoinositides; PI3K, polyphosphoinositide kinase 3; PLC, phospholipase C; PKB, protein kinase B; PTX, pertussis toxin; Rluc, *Renilla reniformis luciferase*; Thr, Thrombin; UII, urotensin II; URP, urotensin II-related peptide; UT, UII receptor.

Synopsis

Urotensin II (UII) and its paralog urotensin II-related peptide (URP) are two vasoactive neuropeptides whose respective central actions are currently unknown. Here, we have compared the mechanism of action of URP and UII on cultured astrocytes. Competition experiments performed with [¹²⁵I]UII showed the presence of very high- and high-affinity binding sites for UII, and a single high-affinity site for URP. Both UII and URP provoked a membrane depolarization accompanied by a decrease of input resistance, stimulated the release of endozepines, neuropeptides specifically produced by astroglial cells, and generated an increase in cytosolic calcium concentration ($[Ca^{2+}]_c$). The UII/URP-induced $[Ca^{2+}]_c$ elevation was pertussis toxin (PTX)-insensitive, and was blocked by the PLC inhibitor U73122 or the IP₃ channel blocker 2-APB. The addition of Ca²⁺ chelator EGTA reduced the peak and abolished the plateau phase whereas the T-type calcium channel blocker mibefradil totally inhibited the calcium response evoked by both peptides. However, URP and UII induced a mono- and biphasic dose-dependent increase in $[Ca^{2+}]_c$ and provoked short- and long-lasting Ca²⁺ mobilization, respectively. Similar mono- and biphasic dose-dependent increase in [³H]inositol incorporation into polyphosphoinositides (PIP) in astrocytes was obtained but sole the effect of UII was significantly reduced by PTX, although BRET experiments revealed that both UII and URP recruited G_o protein. Finally, UII exerted a dose-dependent mitogenic activity on astrocytes, but not URP. Therefore, we described that URP and UII exert not only similar but also divergent actions on astrocyte activity, UII exhibiting a broader range of activities at physiological peptide concentrations.

Introduction

Urotensin II (UII) is a cyclic neuropeptide that has been initially isolated from the urophysis, a neurosecretory organ located in the caudal portion of the spinal cord of teleost fish [1]. Orthologs of UII have been subsequently characterized in the brain and spinal cord of various species of tetrapods including frog, mouse, rat, pig and human [2]. A paralog of UII called UII-related peptide (URP) has been recently identified in rat and

human [3]. UII and URP share a fully conserved C-terminal cyclic hexapeptide CFWKYC core that plays a major role for the biological activity of the peptides [4]. The cyclic region of UII and URP exhibits some structural similarities to that of somatostatin (Fig. 1) and it has recently been found that the genes encoding these three neuropeptides originate from a common ancestor [5].

The mRNAs encoding the UII and URP precursors have been localized both in the brain and in peripheral organs [3]. In the rat and mouse central nervous system (CNS), the UII and URP genes are primarily expressed in subpopulations of motoneurons located in discrete brainstem nuclei and in the ventral horn of the spinal cord [6, 7]. In the human brain, high levels URP mRNA are also present in the hippocampus and thalamus [3]. It has recently been shown that a mature form of UII in the rat brain could only imply atypical cleavage sites in prepro-UII, and that URP might be the sole UII-immunoreactive substance present in the rodent brain [8].

The biological action of UII and URP are mediated through activation of a common G protein-coupled receptor originally called GPR14 [3, 9] and now renamed UT [2]. In UT-transfected CHO and HEK293 cells, UII stimulates the phospholipase C pathway and causes an increase in cytosolic calcium concentration ($[Ca^{2+}]_c$) [10]. UII also exerts mitogenic activity in UT-transfected cells and in vascular smooth muscle cells expressing native UT [10, 11].

In the CNS, UT mRNA has been detected in motoneurons of the spinal cord in mouse [12] and rat [13] as well as in the thalamus, superior occipital gyrus and substantia nigra in human [9]. Previous studies have shown that central administration of UII modulates cardiovascular, locomotor, behavioral and endocrine functions [14, 15]. Although URP has been proposed as the sole endogenous ligand for urotensinergic receptors in the brain [3], little is known regarding the central activities of URP.

It is now clearly established that astroglial cells release various neuroactive compounds [16] including the gliopeptides endozepines. Endozepines were originally characterized as endogenous ligands for benzodiazepine receptors [17]. All endozepines characterized so far derive from an 86-amino-acid precursor called diazepam-binding inhibitor (DBI) which generates several biologically active fragments, notably the octadecaneuropeptide (ODN, DBI₃₃₋₅₀) [18]. The release of

endozepines from astrocytes is regulated by several neuropeptides such as pituitary adenylate cyclase-activating polypeptide (PACAP) and somatostatin [19, 20].

The expression of UII and its receptor has been previously reported in a human glioblastoma cell line [21] and the presence of functional UT has been demonstrated in astrocytes [22, 23]. In astroglial cells, UII specifically binds two affinity sites, stimulates polyphosphoinositides (PIP) metabolism and causes a robust increase in $[Ca^{2+}]$ [22, 24]. Up to now, nothing is known regarding the mechanism of action of URP on native cells, and the relevance for the existence of a redundant peptide for a unique UT receptor remains unexplored. In the present study, we have characterized functional binding sites for URP on rat cortical astrocytes and compared the effects of URP with those of UII on electrophysiological activity, endozepine release, Ca^{2+} , PIP turn-over and cell proliferation.

Experimental

Reagents

URP (ACFWKYCV), rat UII (UII, pQHGTAPECFWKYCI) and rat octadecaneuropeptide ODN (diazepam-binding inhibitor 33-50, DBI₃₃₋₅₀) were synthesized by the solid phase methodology on a Pioneer PerSeptive Biosystem peptide synthesizer (Applera France, Courtaboeuf, France) using the standard manufacturer's procedures as previously described [25]. All peptides were purified (> 98%) on a 2.2 X 25-cm Vydac C₁₈ column (Alltech, France) and characterized by MALDI-TOF MS on a Voyager DE-PRO mass spectrometer (Applera-France). Glutamine, N-2 hydroxyethylpiperazine-N'-2-ethane sulfonic acid (HEPES), the antibiotic-antimycotic solution and bovine γ -globulins were purchased from Invitrogen (Cergy-Pontoise, France). Dulbecco's modified Eagle's medium (DMEM), Ham-F12 culture medium, insulin, D(+)-glucose, mibefradil, nifedipine, ω -conotoxine GVIA, pertussis toxin (PTX), endothelin-1 (ET-1), bovine serum albumin (BSA) were obtained from Sigma-Aldrich (St-Quentin Fallavier, France). Fetal bovine serum (FBS) and interleukin-1 β (IL-1 β) were from Eurobio (Les Ulis, France). Fluo-4 AM and Fura-2 AM were from Molecular Probes (Leiden, Netherlands). Somatostatin-14

and [Tyr⁰,D-Trp⁸]somatostatin-14 were purchased from NeoMPs (Strasbourg, France). Pituitary adenylate cyclase-activating polypeptide (PACAP) was generously provided by Pr Alain Fournier (INRS, Pointe-Claire, Canada).

Astrocyte culture

Purified cultures of astrocytes were prepared as previously described [26]. Briefly, cerebral hemispheres from newborn Wistar rats were collected in DMEM/Ham-F12 (2:1, v/v) culture medium supplemented with 2 mM glutamine, 1% insulin, 5 mM HEPES, 0.4% glucose and 1% of the antibiotic-antimycotic solution. The tissues were disaggregated mechanically with a syringe equipped with a 1-mm gauge needle, and filtered through a 100- μ m pore size mesh filter (Falcon, Becton Dickinson, Grenoble, France). Dissociated cells were resuspended in culture medium supplemented with 10% heat-inactivated FBS and seeded in 150-cm² culture flasks (Falcon) at the density of 2×10^7 cells per flask. The cells were incubated at 37°C in a humidified atmosphere (5% CO₂) and the medium was changed twice a week. When cultures were confluent, the flasks were gently shaken on an orbital shaker at 250 rpm for 2 h. Dislodged cells were discarded and a second step of purification was performed at 250 rpm for 14-16 h. Remaining adhesive cells were collected by trypsination, centrifuged (100 g, 10 min) and plated in 150-cm² flasks. Suspended astrocytes were harvested and seeded in 96-well plates (for calcium studies), 24-well plates (for binding studies), 14-mm glass coverslips (for electrophysiological experiments and [Ca²⁺]_c measurements), 35-mm dishes (for measurement of PIP turnover) and in 60-mm dishes (for measurement of endozepine release). The purity of the cultures was assessed by GFAP immunostaining which showed that enriched cultures contained > 98% astrocytes [22].

Binding studies

Three μ g of UII or [Tyr⁰,D-Trp⁸]somatostatin-14, in phosphate buffer (0.375 mM, pH 7.4), were labeled with 0.5 mCi Na¹²⁵I (Amersham Biosciences) by the lactoperoxidase method as previously described [25]. Mono-iodinated [¹²⁵I]UII or [¹²⁵I-Tyr⁰,D-Trp⁸]somatostatin-14 used for the radioligand binding assays were

purified by reversed-phase HPLC on an Adsorbosphere C₁₈ column (0.46 X 25-cm, Alltech) using a linear gradient (25-65% over 40 min) of acetonitrile/trifluoro acetic acid (99.9 : 0.1, v/v) at a flow rate of 1 mL/min, and stored at 4°C. The specific radioactivity of each radioiodinated peptide was approximately 2000 Ci/mmol.

Cultured astrocytes were rinsed three times with phosphate buffered saline (PBS), dried under a cold air stream and stored at -80°C until binding experiments. Frozen cells were washed twice with assay buffer (50 mM Tris buffer, 1 mM MnCl₂ and 0.5% BSA) and incubated at 22°C in the same buffer in the presence of radiolabeled peptide. At the end of the incubation, cells were washed three times with assay buffer, solubilized with 1% SDS and the radioactivity was counted in a gamma counter (LKB Wallac, Evry, France). For competition experiments, cells were incubated for 3 h with [¹²⁵I]UII (0.2 nM) or [¹²⁵I-Tyr⁰,D-Trp⁸]somatostatin-14 (0.4 nM) in the presence of graded concentrations of unlabeled UII, URP or somatostatin-14. Non-specific binding was determined by addition of 1 μM unlabeled UII or [Tyr⁰,D-Trp⁸]somatostatin-14.

Measurement of cytosolic Ca²⁺ concentration

Purified astrocytes were plated into 96-well plates at 30,000 cells/well or in 14-mm glass coverslips in complete medium added with FBS (10%). After 2 days in culture, astrocytes were rinsed twice with a modified Hank's balanced salt solution (HBSS) containing (in mM): 135 NaCl; 5 KCl; 1 MgCl₂; 1 CaCl₂; 10 HEPES, 3 glucose; 2.5 probenidic (pH 7.4). Cells were incubated at 37 °C with 40 μL of 2 μM fluo-4 AM or Fura-2 AM dye containing 20% pluronic acid for 40 min in a 5% CO₂ atmosphere. Cells were washed twice with modified HBSS, and the effects of graded concentrations of URP and UII on [Ca²⁺]_c were measured with a fluorometric imaging plate reader FlexStation II (Molecular Devices, Sunnyvale, CA). Agonist-induced changes in [Ca²⁺]_c were also measured (fluorescence at 510 nm following excitation at 340/380 nm from a 300 W xenon lamp) in individual cells from a single visual field using an integrating CCD camera coupled to a Leica DMLFSA microscope. Cell calcium imaging and fluorescence quantification were carried-out using the Metafluor Imaging System (Molecular Devices, Sunnyvale, CA).

Measurement of polyphosphoinositide metabolism

Two to five-day-old (2-5 DIV) cultured astrocytes were incubated at 37°C with 10 $\mu\text{Ci/mL}$ myo-[^3H]inositol in glucose-, insulin- and serum-free medium, for 80 to 120 minutes. The incubation was stopped by removing the medium and adding ice-cold 10% trichloroacetic acid. The cells were homogenized and centrifuged (15,000 rpm, 4°C, 10 min). The pellet was washed with 500 μL chilled water and [^3H]PIP were extracted with 500 μL of chloroform/ethanol (2:1, v/v). The radioactivity was counted in a beta counter (LKB 1217 Rack Beta, EG and G Wallac, Evry, France).

Electrophysiological experiments

Electrophysiological recordings were performed at room temperature (22°C) on 2-4 DIV cultured astrocytes seeded on glass coverslips using the standard amphotericin B-perforated configuration of the patch-clamp technique. The patch pipettes were fabricated from 1.5- μm (outer diameter) soft glass tubes on a two-step vertical pipette puller (List-Medical, L/M-3P-A, Darmstadt, Germany). Patch electrodes had a final resistance of 5-7 M Ω . The internal pipette solution contained 130 mM KMeSO₄; 20 mM KCl; 5 mM MgCl₂; 1 mM EGTA; 10 mM HEPES (adjusted at pH 7.4 with KOH). Before each experiment, the medium was replaced by a bath solution: 145 mM NaCl; 5.4 mM KCl; 1.8 mM CaCl₂; 0.8 mM MgCl₂; 10 mM HEPES; 10 mM glucose (pH 7.4 adjusted with NaOH). Cells were transferred to the stage of an upright microscope (DMLFSA, Leica, Heidelberg, Germany). Voltage signals were recorded from an Axopatch 200A amplifier (Axon Instruments, Foster City, CA, USA) and filtered at 2 kHz (3 db, four-pole, Low-pass Bessel filter). Cell input resistance was monitored by applying hyperpolarizing pulses (2 pA, 10 ms, 0.125 Hz) to the cell under study. Data acquisition and analysis were performed through a digidata 1200 interface using the pClamp 8 suite programs (Axon Instruments) and the Origin 4.1 analysis software (Microcal Software, Northampton, MA, USA).

Measurement of endozepine release

5-DIV astrocytes were incubated for 60 min at 37°C with fresh serum-free medium in the absence or presence of test substances. Peptides contained in culture media were concentrated on Sep-Pak C₁₈ cartridges (Waters, Saint-Quentin en Yvelines, France). Bound material was eluted with 50% (v/v) acetonitrile/water containing 0.1% TFA (v/v) and dried by vacuum centrifugation (Speed Vac concentrator, Savant, Hicksville, NY, USA) until radioimmunoassay (RIA).

The concentrations of ODN-like immunoreactivity (ODN-LI) in the Sep-Pak-purified samples were quantified by RIA using an antiserum raised against synthetic rat ODN [20]. The analog [Tyr⁰]-ODN was iodinated using the chloramine-T procedure and purified on a Sep-Pak C₁₈ cartridge. The final dilution of the ODN antiserum was 1:30,000 and the total amount of tracer was 6,000 cpm/tube. The antibody-bound ODN fraction was precipitated by addition of 100 µL bovine γ -globulins (1%, w/w) and 2 mL polyethylene glycol 8000 (20%, w/v). After centrifugation, the pellet containing the bound fraction was counted in a gamma counter (LKB Wallac, Rockville, MI, USA).

Plasmid construction, HEK transfection and BRET assay

Human UT receptor (hUT) and protease-activated receptor 1 (PAR-1) in pCDNA3.1 were purchased from cDNA Ressources Center (Rolla, MO, USA). The ability of receptors to activate G_{i/o} proteins was controlled by measuring BRET between G protein subunits. Thus, we used G_α-Rluc fused proteins, the *Renilla reniformis luciferase* (Rluc) fused to the G protein α subunit G_{α0} in the loop between helices α A and α B in the h- α domain [28], and the Venus tagged G_{γ2} subunit generously provided by Dr C. Galès (Inserm U858, Toulouse, France).

HEK-cells (HEK293) were grown in complete medium (DMEM supplemented with 10% (v/v) FBS, 4.5 g/L glucose, 100 U/mL penicillin, 0.1 mg/mL streptomycin, and 1 mM glutamine (Invitrogen)). Transient transfections were performed with the indicated plasmids using lipofectamine 2000 following the manufacturer's protocol (Invitrogen). All constructs were verified by sequencing, and their expression in HEK293 cells was confirmed by BRET signals.

For BRET experiments, 48 h after transfection, HEK293 cells were washed with PBS, and coelenterazine *h* substrate (Promega, Charbonnières, France) was added at a final concentration of 5 μM in the total volume of 30 μL /well at 37 °C. For kinetic analysis, BRET readings were then performed immediately after coelenterazine addition. After BRET signal stabilization (≈ 100 sec), the different ligands were injected by the Mithras LB 940 plate reader (Berthold Biotechnologies, Bad Wildbad, Germany). BRET signals were expressed in milliBRET units of BRET ratio as previously described [29].

Cell proliferation

To determine cell survival and proliferation, 2-DIV cultured astrocytes were grown in the complete culture medium supplemented with 10% FBS. Cells were either incubated in the absence or presence of URP, UII, endothelin-1, PACAP or interleukin-1 β (IL-1 β) for 24 h in the same complete medium (WST-1 assay, 2 h) or rinsed in FBS-free medium and incubated after 24 h in the absence or presence of URP or UII for 48 h (cell counting). Astrocytes were rinsed three times in PBS (37°C) and cell proliferation was quantified by fluorescence measurement with a FL600 microplate reader (450/750 nm) or by counting the cell number with an electronic cell counter (Z2, Beckman Coulter, Roissy, France).

Data analysis

To calculate the Hill coefficient (n_H), competition curves were first fitted to the Hill equation *via* a nonlinear regression using the computerized curve-fitting package GraphPad Prism version 4 for Windows (GraphPad Software, San Diego, CA, USA). The data were then re-fitted using equation for displacement of radioligand by competitors to one or two binding sites. A two-site curve-fitting model was retained when this model fitted the data significantly better than a one-site model, as determined by a *F*-test at a significance level of $p < 0.05$. IC_{50} values derived from these latter fits were converted to apparent competition-receptor dissociation equilibrium constants K_1 for the very high affinity and K_2 for the high-affinity binding sites. For biological activities, values presented in the figures are means \pm S.E.M.

Statistical significance of differences was determined by using the one-way ANOVA followed by a Dunnett's post hoc test or the non-parametric Friedman's test.

Results

Binding characteristics of URP on cultured rat astrocytes

Competition experiments performed with [125 I]UUII (0.2 nM) as a tracer and unlabelled UUII (1 pM-1 μ M) generated a biphasic curve with a Hill coefficient of 0.81 ± 0.06 (Fig. 2A). In contrast, graded concentrations of unlabeled URP (1 pM-1 μ M) generated a monophasic displacement curve with a Hill coefficient close to 1. Statistical analysis of the competition experiments using the *F*-test showed that data best fitted with a two-site model ($p < 0.05$) and the calculated dissociation constants for the high- (K_1) and low-affinity (K_2) sites were 0.31 ± 0.12 and 32.9 ± 18.7 nM for UUII and with a one-site model ($p < 0.05$) yielding a dissociation constant for a high-affinity (K) site of 1.62 ± 0.74 nM, for URP (Fig. 2A, inset). We thus tested the ability of URP and UUII, which exhibit sequence similarities with somatostatin in their cyclic portion, to displace the binding of the somatostatin analog [125 I-Tyr 0 ,D-Trp 8]somatostatin-14 (0.4 nM). Graded concentrations of unlabeled URP (10 pM-10 μ M) showed a biphasic competition curve with a Hill coefficient of 0.32 ± 0.09 (inset) and dissociation constants for high (K_1)- and low (K_2)-affinity sites of 9.16 ± 5.32 nM and 4.42 ± 1.72 μ M, respectively. In contrast, UUII (3 nM-30 μ M) failed to displace [125 I-Tyr 0 ,D-Trp 8]somatostatin-14 below micromolar concentrations (Fig. 2B).

Similar effects of URP and UUII on resting membrane potential and endozepine release from cultured rat astrocytes

To investigate the action of URP and UUII on the electrical activity of cultured astrocytes, amphotericin-perforated patch-clamp recordings were performed. Under current-clamp conditions (holding potential at -52.12 ± 3.08 mV; $n = 32$), focal application of URP (Fig. 3A, *upper*; 100 nM; 2 min) or UUII (Fig. 3A, *lower*; 100 nM; 2 min) elicited a membrane depolarization in 69% of astrocytes (20/29) accompanied by

a decrease of the input resistance. The majority of responding astrocytes remained in a depolarized state during the washout period (Fig. 3A). The amplitudes of membrane depolarization induced by URP at concentrations of 10 nM (9.6 ± 2.6 mV, $n = 5$) and 100 nM (16.6 ± 4.3 mV, $n = 8$) were very similar to those evoked by UII (10 nM, 9 ± 4 mV, $n = 3$; 100 nM, 15.5 ± 2.8 mV, $n = 12$, Fig. 3B).

Graded concentrations of URP (1 pM-10 μ M, 1 h) induced a dose-related increase in the release of ODN-LI with an EC_{50} value of 3.95 ± 1.52 nM and a maximum effect at a concentration of 1 μ M (*data not shown*). Incubation of cells with 0.1 nM, 10 nM and 1 μ M URP or UII provoked a concentration-dependent increase in ODN-LI release, non significantly different between URP and UII (Fig. 3C).

Similar mechanism of URP and UII on $[Ca^{2+}]_c$ mobilization in cultured rat astrocytes

URP and UII (0.1 μ M, each) induced a rapid $[Ca^{2+}]_c$ increase (peak) followed by a sustained response (plateau) [22]. Preincubation of astrocytes with the $G_{i/o}$ protein inhibitor pertussis toxin (PTX, 0.2 μ g/mL, 18 h) did not affect the Ca^{2+} responses induced by both peptides (Fig. 4A). In contrast, the URP- and UII-induced Ca^{2+} mobilizations were completely blocked by the PLC inhibitor U73122 (10 μ M, 20 min) and were strongly reduced by the IP_3 -induced Ca^{2+} release and store-operated Ca^{2+} channels blocker 2-APB (50 μ M, 20 min), by 83 and 80%, respectively (Fig. 4B, C). In order to gain insight into the Ca^{2+} pools recruited by UII and URP, $[Ca^{2+}]_c$ were measured in the absence or presence of the extracellular Ca^{2+} chelator EGTA (1 mM, 20 min). Peak responses were partially attenuated by 53% (URP) and 58% (UII) and plateau phases were totally blocked in the presence of both peptides (Fig. 4D). The $[Ca^{2+}]_c$ increase evoked by UII and URP was also suppressed in the presence of the voltage-dependent T-type Ca^{2+} channel blocker mibefradil (1 μ M, Fig. 4E, 20 min), whereas the specific L- and N-type Ca^{2+} channel blockers, nifedipine and ω -conotoxin GVIA (1 μ M, each, 20 min), respectively, did not significantly affect the peptide-evoked elevation of $[Ca^{2+}]_c$ in astrocytes (Fig. 4F, G).

Divergent actions of URP and UII on $[Ca^{2+}]_c$ mobilization and PIP turnover in cultured rat astrocytes

As shown in Fig. 5A, exposure of astrocytes to increasing concentrations of URP and UII (1 pM-10 μ M, each) provoked a robust and sustained increase in $[Ca^{2+}]_c$, followed by a plateau phase. The dose-response study of the effects of URP and UII on $[Ca^{2+}]_c$ mobilization revealed that URP and UII induced a mono- and biphasic dose-dependent $[Ca^{2+}]_c$ increase (Fig. 5A). The maximum stimulations (URP, 157%; UII, 151%) were obtained at 1 μ M for both peptides. The peptide-induced Ca^{2+} response were also investigated at a single astrocyte level using a digital imaging system. Application of 0.1 μ M URP resulted in a rapid and transient increase in $[Ca^{2+}]_c$ levels ($136.9 \pm 3.5\%$), in a subset of 59% astrocytes (16/24, Fig. 5B). Surprisingly, the UII-evoked $[Ca^{2+}]_c$ response was characterized by a rapid increase followed by a sustained or developing increase of the $[Ca^{2+}]_c$ level, reaching $137.1 \pm 5.2\%$ stimulation after 400 sec of recording, in 70% of cell (21/26, Fig. 5C).

As observed on Ca^{2+} levels, exposure of astrocytes to graded concentrations of URP (1 pM-1 μ M) induced a dose-dependent increase in $[^3H]$ inositol incorporation into PIP with an EC_{50} value of 2.19 ± 1.42 nM (Fig. 6A). In contrast, UII (1 pM-1 μ M) evoked a biphasic dose-dependent stimulation of PIP turnover yielding an EC_{50}^1 of 9.1 ± 4.6 pM and an EC_{50}^2 of 11.0 ± 4.6 nM (Fig. 6A). The amplitude of the stimulatory effect of URP (10 nM; $141.5 \pm 8.6\%$ of control) on PIP metabolism was not significantly different from that obtained with UII (10 nM; $154.4 \pm 12.8\%$ of control). Co-application of URP (10 nM) with a saturating concentration of UII (10 nM) induced an increase in $[^3H]$ inositol incorporation into PIP ($160 \pm 12.8\%$) that did not significantly exceed the effect of each peptide administered alone (Fig. 6B). In same sets of experiment, PTX (0.2 μ g/mL; 18 h) significantly decreased the incorporation of $[^3H]$ inositol into PIP induced by UII (100 nM, -45%) but failed to significantly affect the stimulatory effect of URP (100 nM, -26%) (Fig. 6C).

Specific activation of G_o protein by UII and URP in cells expressing recombinant hUT

To control whether UT might be coupled to $G_{i/o}$ proteins in the presence of UII, but also URP, we used the BRET approach to directly monitor $G_{\alpha o}$ protein activation in living cells [30, 31]. This assay is based on the G protein complex conformational change and the dissociation of the $G_{\alpha o}$ fused with the energy donor Rluc, from the $G_{\gamma 2}$ fused with the energy acceptor YFP upon receptor activation. These fused G protein subunits were shown to be correctly expressed and to allow an efficient coupling of receptors to their effectors (*data not shown*). In HEK293 cells coexpressing $G_{\alpha o}$ -Rluc, $G_{\gamma 2}$ -YFP and either recombinant hUT (HEK293-hUT) or PAR-1 (HEK293-PAR-1), a significant BRET signal was detected compared to cells expressing only $G_{\alpha o}$. When HEK293-hUT cells were treated with PBS or Thrombin (Thr, 50 U/mL), the BRET signal is not modified (Fig. 7A). In contrast, kinetic analysis showed that UII and URP (10 μ M, each) injection provoked BRET signal decrease between $G_{\alpha o}$ and $G_{\gamma 2}$ subunits, which occurred rapidly and persisted for more than one minute (Fig. 7B). In order to control the specificity of the UT- G_o coupling, BRET signal was measured on HEK293-PAR-1 in the presence of Thr, UII and URP. In these cells, Thr induced BRET decrease whereas URP and UII failed to activate G protein dissociation (Fig. 7C).

Effect of URP and UII on cell viability/proliferation in cultured rat astrocytes

2-DIV cells (non synchronized) were cultured in complete medium additionned with FBS and then incubated in a FBS-free medium containing URP or UII (10 nM, each). Cell viability was measured 24 h later by using WST-1 assay, allowing detection of mitochondrial activity related to the number of viable cells. Fig. 8A showed that URP failed to affect the cell viability whereas UII stimulated by 132.7% the astrocyte growth. PACAP (10 nM), interleukin-1 ($IL-1\beta$, 10 ng/mL) and endothelin-1 (ET-1, 10 nM) were also tested as positive controls and gave rise respectively to 126.3, 169.3 and 171.1% increase (Fig. 8A). The dose-response effects of URP and UII on cell growth were also investigated in non-synchronized (WST-1, Fig. 8B) and synchronized (18 h of harvesting, counting, Fig. 8C) astrocytes. A 24 h

(Fig. 8B) or 48 h-treatment (Fig. 8C) with graded concentrations of URP failed to significantly modify the rate of cell growth. In contrast, UII induced a dose-dependent cell growth, yielding an EC_{50} value of 2.37 ± 0.02 nM, and a maximum stimulatory effect of $132.9 \pm 7.0\%$ at 1 μ M (Fig. 8B) in non-synchronized cultures, and evoked a significant cell number increase ($120.1 \pm 2.9\%$) at 0.1 μ M in synchronized conditions (Fig. 8C).

Discussion

Although the cardiovascular effects of the urotensinergic system are now well characterized, little is known concerning the roles of UII and URP in the CNS. Immunohistochemical studies have shown the presence, in the rat brain, of UT-like immunoreactivity in GFAP-positive cells within the brainstem, hypothalamus and thalamus [23]. In agreement with these observations, we have recently demonstrated that rat cortical astrocytes express UT mRNA, contain the UT protein, and exhibit very high- and high-affinity UII binding sites [22]. In these cells, UII specifically activates a PLC/PIP/ Ca^{2+} transduction pathway, *via* both PTX-sensitive and -insensitive G proteins [22]. The identification of a UII paralog, URP, which apparently exhibits the same pharmacological characteristics as UII [3], raises the question as to whether URP functions as a UII backup in the brain. In the present study, we have looked for the existence of URP binding sites in cultured rat astrocytes and we have compared the mechanisms of action of URP and UII on astroglial cell activity.

Competition experiments revealed that URP could displace [125 I]UII at nanomolar concentrations ($K = 1.62$ nM) with a Hill coefficient close to 1, suggesting the existence of an homogeneous population of URP binding sites in astrocytes. However, the occurrence of two affinity binding sites for UII has been previously described in CHO cells expressing recombinant human UT [27] and in rat cortical astrocytes [22]. In our model, URP is five fold less potent in competing for the very high affinity UII binding site and twenty fold more potent in displacing UII from the high-affinity site, indicating that UII and URP exhibit different binding characteristics in astrocytes. Interestingly, URP and the C-terminal octapeptide of human UII, UII₄₋₁₁,

were 10 times more potent than human UII in competing with [125 I]UII at recombinant UT [3, 32], suggesting that the N-terminal segment of UII decreases the affinity of the peptide for its receptor. It has previously been shown that cultured rat astrocytes express UT mRNA and that UT-like immunoreactivity is located at the plasma membrane of astroglial cells [22, 23]. Thus, the present study suggests that the high affinity site bound by URP actually correspond to UT. The cyclic hexapeptide core of UII and URP exhibits substantial similarities with the biologically active region of somatostatin-14 ([33, 34]; Fig. 1) and UT shares sequence identity with somatostatin receptors (sst) [35]. In spite of these relatively high degree of sequence homologies, UII exhibits very low affinity for somatostatin receptors (sst1, sst2 and sst4) either transfected in cell lines [36] or naturally expressed in astrocytes [20, 22, 37]. The present data show that URP could displace [125 I-Tyr⁰,D-Trp⁸]somatostatin-14 binding to both high- and low-affinity sites. This observation indicates that, unlike rat UII that selectively binds to UT, URP can interact with both UT and somatostatin receptors expressed in rat cortical astrocytes. In agreement with these data, a recent comparative genomic study has established the existence of a close evolutionary relationship between urotensinergic and somatostatinergic gene families, particularly between URP and somatostatin-14 peptides [5]. Moreover, it has recently been demonstrated that high concentration of URP stimulates [Ca^{2+}]_c levels and proliferation of recombinant sst-2 expressing CHO cells [38]. Together, it is suggested that shorter urotensinergic peptides in their N-terminal sequence (Fig. 1; human UII and URP) were more potent than longer UII (rat UII) in competing for the high affinity UII binding site UT, and also acquired the ability to bind sst.

Astrocytes have long been considered as non-excitabile support of neuronal cells in the brain. It is now well established that astroglial cells express functional channels and receptors enable to sense synaptic transmission [39]. In addition, astrocytes participate in neuronal functioning not only *via* feeding and structural support, but also through release of gliotransmitters such as glutamate, D-serine, ATP, cytokines or endozepines [16]. Application of URP or UII in the vicinity of astrocytes in culture induced a slow in onset membrane depolarization accompanied by a decrease of the input resistance. It has also been demonstrated that UII depolarizes nervous cells on

brainstem cholinergic neurons and evokes a membrane conductance decrease [40, 41]. Astrocytes in culture or acutely dissociated from different brain regions, may express various ion channels including both T- and L-type Ca^{2+} channels, different K^{+} channels and also store-operated Ca^{2+} channels [42]. Thus, URP as well as UII may inhibit K^{+} channel activity and/or stimulate voltage-activated Ca^{2+} channel opening either directly or upon depolarizing stimulus. Astrocytes are able to synthesize, store and release endozepines *via* Ca^{2+} -dependent and/or cAMP/PKA-dependent ways [19]. It is likely that depolarization of glial cells evoked by both peptides subserves neurotransmitter secretion, transcription or proliferation.

It is now clearly established that activation of UII receptor expressed in native cells or in transfected-cell lines, is associated with an increase in PIP turnover causing mobilization of Ca^{2+} from intracellular stores [43, 44]. Exposure of astrocytes to URP and UII was associated with $[\text{Ca}^{2+}]_c$ mobilization, *i.e.* a rapid peak response followed by a plateau phase. Peptide-induced $[\text{Ca}^{2+}]_c$ increase was PLC-dependent (U73122 sensitive) and resulted from activation of IP_3 -receptor pools (2-APB sensitive). These data are in a good agreement with previous observations made in sartorius muscle, vascular tissues, or in rhabdomyosarcoma cell lines [44-46]. Since PLC-inducing Ca^{2+} mobilization is typically mediated *via* G_q and/or $\text{G}_{i/o}$ protein, and UII receptor sites might be linked to $\text{G}_{i/o}$ protein types [3, 22, 46], the ability of URP *versus* UII was tested in PTX-pretreated astrocytes. PTX, which catalyses ADP-ribosylation of $\text{G}_{i/o}$ -type G proteins and consequently inactivates their transduction mechanisms [47], did not affect the Ca^{2+} response evoked by both peptides, indicating that URP and UII tested at micromolar range activate a receptor coupled to a PLC/ IP_3 / Ca^{2+} pathway in a $\text{G}_{i/o}$ independent manner. The involvement of the extracellular Ca^{2+} pool was also tested by means of experiments in the presence of EGTA, and voltage-gated Ca^{2+} channel blockers. It is thus demonstrated that extracellular chelation of Ca^{2+} attenuated the peak, and totally blocked the plateau responses, indicating that the transient Ca^{2+} peak is partially attributable to Ca^{2+} release from the sarcoplasmic reticulum, whereas the sustained plateau phase is due to Ca^{2+} influx. Surprisingly, $[\text{Ca}^{2+}]_c$ increase induced by both peptides was completely abolished by the specific inhibition of the T-type calcium channel (mibefradil) and appeared insensitive to both blockage of L- and N-

type channels by nifedipine and ω -conotoxine GVIA, respectively. These observations suggest that high level of UT activation leads to the recruitment of both extra- and intracellular Ca^{2+} pools and to a cross-talk between IP_3 receptors and at least voltage-dependent T-type channels. T-type channels are represented by three different Ca^{2+} channels, *i.e.* $\alpha 1\text{G}$ or Cav3.1, $\alpha 1\text{H}$ or Cav3.2 and $\alpha 1\text{I}$ or Cav3.3 [48] and mRNA transcripts encoding the functional $\alpha 1\text{G}$ T-type subunit channels have been found in cultured astrocytes [49]. An interesting property of T-type Ca^{2+} channels is their ability to sustain a continuous Ca^{2+} influx in glia at rest by a window current mechanism [50]. Interestingly, changes in membrane potential and intracellular Ca^{2+} may affect Gq-protein coupled IP_3 production itself [51, 52]. The components of the basic $\text{G}_q\text{PCR}/\text{IP}_3$ production pathway would be located in or associated with the plasma membrane, making them potentially susceptible to changes in membrane potential lipid in the inner leaflet of the plasma membrane [53]. These observations may unveil a similar UT- G_q associated mechanism in astrocytes, where blockage of T-type Ca^{2+} channels may hyperpolarize astrocytes and inhibit IP_3 -induced Ca^{2+} release.

The dose-response curves of URP and UII on $[\text{Ca}^{2+}]_c$ show mono- and bi-phasic profiles, respectively. The agonist potency of URP ($\text{EC}_{50} \sim 0.7 \text{ nM}$) is similar to that one of the two sites of UII ($\text{EC}_{50} \sim 0.7 \text{ nM}$), and in close agreement with the binding characteristic at the common high affinity sites of URP and UII, positively coupled to a PLC- G_q protein pathway. It thus appears that the very high affinity binding site of UII, is not activated by URP and is functionally coupled to $[\text{Ca}^{2+}]_c$. Single-cell digital imaging techniques demonstrated that a majority of cells responded to URP and UII, even if the cell-cell variations in shape and amplitude can be observed. More surprisingly, URP ($0.1 \mu\text{M}$) evoked a transient and reversible increase in $[\text{Ca}^{2+}]_c$ whereas a single UII ($0.1 \mu\text{M}$) infusion induced a Ca^{2+} response that remained stable during more than 400 sec of recording. This last effect may be attributable to the slow dissociation rate of UII, as already described for rat and human UII on UT-transfected cells, skeletal muscle myoblasts and astrocytes [22, 27, 46, 54]. Thus, the slow dissociation might be specific of UII and would likely accounts for the sustained and washout-resistant contractile responses induced by UII on primate arteries and $[\text{Ca}^{2+}]_c$ increase in rat cortical astrocytes.

As observed for Ca^{2+} responses, [^3H]inositol incorporation in PIP shows mono- and biphasic profiles for URP and UII, respectively. Thus, URP and UII produced a concentration-dependent increase of $[\text{Ca}^{2+}]_c$ consistent with a UT- G_q coupling to PLC and IP_3 formation-inducing Ca^{2+} release from intracellular stores. The percentage increase of PIP metabolism produced by a concomitant administration of URP and UII at their maximum effective concentrations was not significantly different from the effects of both peptides individually tested, indicating that this pathway might be common to the both peptides.

Here, we observed that the binding and the stimulatory effect of URP on the PIP metabolism were not significantly affected by a pre-treatment with PTX whereas half of the UII-induced PIP formation was blocked by PTX. This observation indicates that URP would not be promiscuously linked to the $\text{G}_{i/o}$ PTX-sensitive binding site in astrocytes. In order to clarify a possible URP and UII-induced UT coupling to $\text{G}_{i/o}$ proteins, kinetic BRET signal changes were measured in HEK293 cells transiently cotransfected with $\text{G}_{\alpha o}$ -Rluc and $\text{G}_{\gamma 2}$ -YFP, and with either hUT or the control receptor PAR-1. In these conditions, UII but also URP provoked a fast decrease of BRET exclusively in cells expressing hUT, resulting from UT- G_o activation and $\text{G}_{\alpha o}$ - $\text{G}_{\gamma 2}$ dissociation. These data indicate that in a recombinant system, both UII and URP are able to activate a UT- $\text{G}_{i/o}$ PTX sensitive coupling system.

In astrocytes, PTX partially inhibited the effect of UII, and in a very less extent of URP, on PIP whereas it failed to block the UII/URP-evoked Ca^{2+} response, suggesting the existence of a specific PTX-insensitive UT- G_q -PLC- IP_3 - Ca^{2+} preferential coupling, and a PTX-sensitive UT- $\text{G}_{i/o}$ -PIP pathway more specific to UII. These two coincident cascades likely converge to an increase of the [^3H]inositol incorporation into the PIP. Indeed, the stimulatory effect of UII and URP on the PIP turnover can be ascribed to *i*) the activation of the PIP_2 hydrolysis into diacylglycerol and IP_3 by PLC, culminating in the release of calcium from internal stores, and *ii*) the production of PIP, PIP_2 , and PIP_3 upon the activation of PI-kinases in a Ca^{2+} independent manner. In particular, PIP synthesized by phosphatidylinositol-3 kinase (PI3K), are known to play a critical role in cell survival by membrane recruitment and activation of Akt/PKB kinase [55]. Since the PI3K has recently been shown to be

involved in the UII-induced natriuretic peptide release from rat atria [56] and the MAPK pathway appeared clearly implicated in several UII processes including vascular and airway smooth muscle cell proliferation and migration, fibroblast proliferation or cardiomyocyte hypertrophy [57], we hypothesize that a UT-G_{i/o}-PI3K-PTX sensitive pathway mainly activated by UII, and in a less extent by URP, might thus be involved in Ca²⁺-independent astrocyte activities.

Pathological processes in the brain are usually accompanied by a significant astrocyte proliferation referred as to astrogliosis. Here, we demonstrated that UII, but not URP, significantly stimulated rat cortical astrocyte proliferation. Numerous studies have demonstrated that vasoactive peptides or interleukins play important roles in astrogliosis [58]. Thus, we demonstrated that in the same set of experiments, PACAP and ET-1 tested at the same concentration than URP and UII, as well as IL1- β , also markedly increase cell growth. Moreover, graded concentrations of URP and UII were tested on either non synchronized or synchronized cells, and the data demonstrate the existence of a UII-specific dose-dependent increase of cell viability. A similar activity for UII has already been observed on rat vascular smooth muscle cells [11], on human endothelial cells [59], and on UT transfected CHO cells [10]. Considering that URP has been shown to activate porcine sst2 [36], we proposed that the absence of mitogenic role of URP would be linked to its ability to bind sst expressed by astrocytes, as sst2 [37, 20]. It is conceivable that concomitant UT-G_q and sst2-G_i recruitments by URP may *i)* also lead to [Ca²⁺]_c increase and/or *ii)* antagonize some specific very high-affinity UII/UT-mediating transduction pathways involved in cell proliferation.

In conclusion, the present study has demonstrated for the first time that URP and UII activate functional receptors in native rat cortical astrocytes, exhibiting similar and distinct ways of action. URP and UII bind a high affinity site that exhibits the pharmacological characteristics of UT, mediating cell depolarization, endozepine release and PLC-dependent Ca²⁺ mobilization through intra-(IP₃ receptor) and extracellular (T-type Ca²⁺ channels) pools (Fig. 9). Ca²⁺ entry through T-type channels appears essential for IP₃-induced Ca²⁺ increase whereas Ca²⁺ release from intracellular stores induces Ca²⁺ release. In addition, UII, and in a less extent URP, are able

to activate a higher affinity site coupled to a PTX-sensitive G protein, that would mediate an hypothetical PI3K activation, relaying an important physiological astrocyte behavior, *i.e.* cell proliferation (Fig. 9). Finally, URP, but not UII, may bind sst expressed in rat astrocytes whose activation would mimick the UT-Ca²⁺ coupling and/or counteract the mitogenic role of UT (Fig. 9). These data provide the first understanding of the involvement of the two vasoactive peptides UII and URP in nerve cell activity, stressing the specific role of UII in the (patho)physiology of astrocytes.

Acknowledgements

We gratefully acknowledge Mr Sébastien Arthaud, Mrs Huguette Lemonnier and Mr Gérard Cauchois for skillful technical assistance. We thank Laetitia Comps-Agrar, Claire Vol, and Mohammed Akli Ayoub for their technical and scientific assistance, and the Pharmacology Screening-Interactome Platform facilities of the Institut Fédératif de Recherche 3 of Montpellier (France). This work was supported by Inserm (U982), the European Institute for Peptide Research (IFRMP 23), the Lille-Amiens-Rouen-Caen Neuroscience network and the Conseil Régional de Haute-Normandie. M.D. was recipient of a fellowship from the Lille-Amiens-Rouen-Caen Neuroscience network and the Conseil Régional de Haute-Normandie. H.V. was recipient of the De Bétancourt-Perronet award for scientific cooperation between France and Spain. We thank Pr Alain Fournier (INRS, Pointe-Claire, Canada), who has generously provided PACAP.

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Figure legends

Figure 1 Comparison of the primary structures of rat and human urotensin II (UII), urotensin II-related peptide (URP) and somatostatin-14.

Common residues are indicated in bold characters.

Figure 2 Displacement curves and binding parameter tables comparing competition of [¹²⁵I]UII and [¹²⁵I-Tyr⁰,D-Trp⁸]somatostatin-14 binding by UII, URP and somatostatin-14 on cultured rat astrocytes.

(a) Cells were incubated for 3 h at 22°C with 0.2 nM of [¹²⁵I]UII in the absence or presence of graded concentrations (1 pM to 1 μM) of URP (■) or UII (▣). (b) Cells were incubated for 3 h at 22°C with 0.4 nM of [¹²⁵I-Tyr⁰,D-Trp⁸]somatostatin-14 in the absence or presence of graded concentrations of URP (■, 10 pM to 10 μM), UII (●, 3nM to 30 μM) or somatostatin-14 (□, 10 pM to 0.3 μM). Data are mean of three to five independent experiments performed in triplicate. *K*, dissociation constant; *n_H*, Hill coefficient.

Figure 3 Effects of URP and UII on the resting membrane potential and on the release of octadecaneuropeptide immunoreactivity (ODN-LI) from cultured rat astrocytes.

(a) Typical voltage responses to URP and UII (100 nM, 2 min, each) obtained in the amphotericin B-perforated configuration with the current-clamp mode. Hyperpolarizing pulses (2 pA, 10 ms, 0.125 Hz) were applied to monitor the cell input resistance. (b) Quantitative analysis of URP- and UII-induced membrane depolarizations. (c) Cells were incubated for 1 h in the absence or presence of graded concentrations (0.1 nM, 10 nM, 1 μM) of URP or UII. Data are mean ± S.E.M. of three to twelve independent experiments. *, *p* < 0.05; **, *p* < 0.01; one way ANOVA followed by a Dunnett's *post-hoc* test.

Figure 4 Mechanisms of URP- or UII-induced increase of $[Ca^{2+}]_c$ in cultured rat astrocytes.

Cells were loaded with fluo-4 AM (40 min) and exposed to PTX (0.2 μ g/mL, **a**), U73122 (10 μ M, **b**), 2-APB (50 μ M, **c**), EGTA (3 mM, **d**), mibefradil (1 μ M, **e**), nifedipine (1 μ M, **f**) or ω -conotoxin GVIA (1 μ M, **g**) (white histograms) and/or to URP or UII (0.3 μ M, each, dashed/colored histograms). Data are expressed as percentages of the corresponding control values in the absence of blockers and peptides. Data are mean \pm S.E.M. of at least 7 independent experiments. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; NS, non-significant; one way ANOVA followed by a Dunnett's *post-hoc* test.

Figure 5 Divergent effects of URP and UII on $[Ca^{2+}]_c$ in cultured rat astrocytes.

(**a**) Cells were loaded with fluo-4 AM (40 min) and exposed to graded concentrations of UII or URP (1 pM, 1 nM, 10 nM, 0.1 μ M and 1 μ M, each). Representative recordings of cells exposed to URP (left) and UII (middle). Dose response curve of the mean of maximum amplitude of $[Ca^{2+}]_c$ transient induced by URP or UII of at least 10 independent experiments (right). The results are expressed as percentages of the corresponding control values in the absence of peptides. (**b** and **c**) Cells were loaded with fura-2 AM (30 min) and perfused (\sim 100 sec) with URP (0.1 μ M, **b**) or UII (0.1 μ M, **c**). Galleries of images illustrate the time course change of fluorescence before perfusion (control), at the maximum amplitude (peak), at 300 sec and at 400 sec of recording. Bar graphs represent means of response at peak, at 300 and at 400 sec of recording. Curves show typical recordings of single astrocytes responding to URP (**b**) and UII (**c**). The results are expressed as percentages of the corresponding control values in the absence of peptide (control). UII or URP vs control, * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; one way ANOVA followed by a Dunnett's *post-hoc* test. Mean values at 300 or 400 sec vs peak, $^{\#\#}$, $p < 0.01$; NS, non-significant; non-parametric Friedman's test.

Figure 6 Distinct activities of URP and UII on PIP metabolism in cultured rat astrocytes.

Cells were incubated with [³H]inositol in the absence or presence of test substances. (a) Effects of graded concentrations of URP and UII (1 pM to 1 μM, each). (b) Comparison of the effect of URP (10 nM), UII (10 nM) or URP+UII (10 nM, each). (c) Effect of PTX (0.2 μg/mL, 18 h) on URP- and UII- (100 nM, each) induced [³H]inositol incorporation. Data are mean ± S.E.M. of four independent experiments performed in triplicate. PTX, URP or UII vs control, *p < 0.05; **p < 0.01; ***p < 0.001; NS, not statistically different from the control; UII vs URP, #p < 0.05; ###p < 0.001; UII or URP vs UII+PTX or URP+PTX, †p < 0.05; one way ANOVA followed by a Dunnett's *post-hoc* test.

Figure 7 BRET kinetic analysis of the effect of UII and URP on G_o protein activation.

Cells transiently coexpressing G_{oo}-Rluc, γ₂-YFP with hUT (a and b) or PAR-1 (c) were used for BRET experiment. Repetitive signals were recorded during 200 sec immediately before and after injection (arrow) of PBS or Thrombin (Thr, 50 U/mL) on hUT expressing HEK293 (HEK293-hUT, a), UII (0.1 μM) or URP (0.1 μM) on HEK293-hUT (b) and Thr, UII or URP on HEK293 expressing PAR-1(HEK293-PAR-1, c).

Figure 8 Effects of UII and URP on cultured rat astrocyte viability.

(a) Non-synchronized cells were incubated for 24 h in the absence or presence of URP (10 nM), UII (10 nM), PACAP (10 nM), IL-1β (10 ng/mL) or ET-1 (10 nM). (b) Non-synchronized cells were incubated for 24 h in the absence or presence of graded concentrations of URP or UII (10 pM to 1 μM). (c) Synchronized cells were incubated for 24 h in the absence or presence of graded concentrations of URP or UII (0.1 pM to 0.1 μM). For a and b, viability was determined with WST-1; for c, the mitogenic activity was determined by counting cell number. The results are expressed as percentages of control values. Data are mean ± S.E.M of three independent

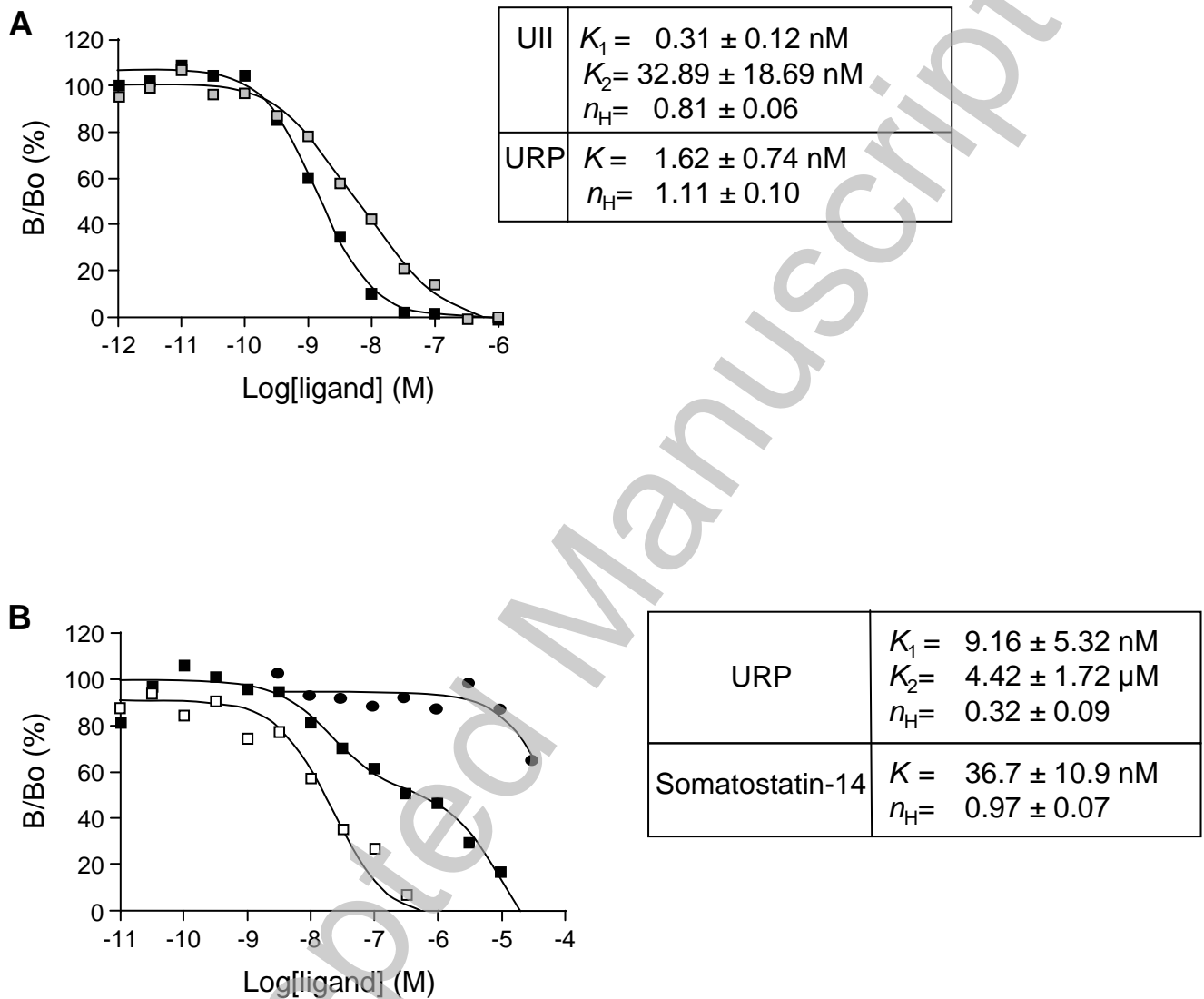
experiments performed in quintuplicate. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; NS, not statistically different from the control; one way ANOVA followed by a Dunnett's *post-hoc* test.

Figure 9 Proposed model depicting distinct and common transduction pathways activated by URP and UII on cultured rat astrocytes.

At low concentrations, UII binds a very high affinity site and activates a PTX-sensitive $G_{i/o}$ protein-coupled UT signaling pathway involved in PIP turnover, likely through IP kinases as PI3K, which would be involved in astrocyte proliferation. This specific UT coupling may be also, but in a very less extent, activated by URP. At higher concentrations, URP and UII recruit a high affinity site also attributable to UT that can be both associated to a $G_q/PLC/IP_3/Ca^{2+}$ pathway and T-type voltage-dependent Ca^{2+} channels. These extra- and intracellular Ca^{2+} pools appear cross-dependent, and would be involved in astrocyte depolarization and endozepine release. At high concentrations, URP may in addition bind sst expressed by astrocytes, such as sst2. This URP specific activation of sst could synergize the $[Ca^{2+}]_c$ increase evoked by UT, and/or counteract the UT-associated proliferation.

Peptides	Sequences		
rat Ull	QHGTAPEC	FWKY	CI
human Ull	ETPDC	FWKY	CV
human Ull ₄₋₁₁	DC	FWKY	CV
URP	AC	FWKY	CV
Somatostatin-14	AGCKNFFWKTFTSC		

Figure 1
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Figure 2
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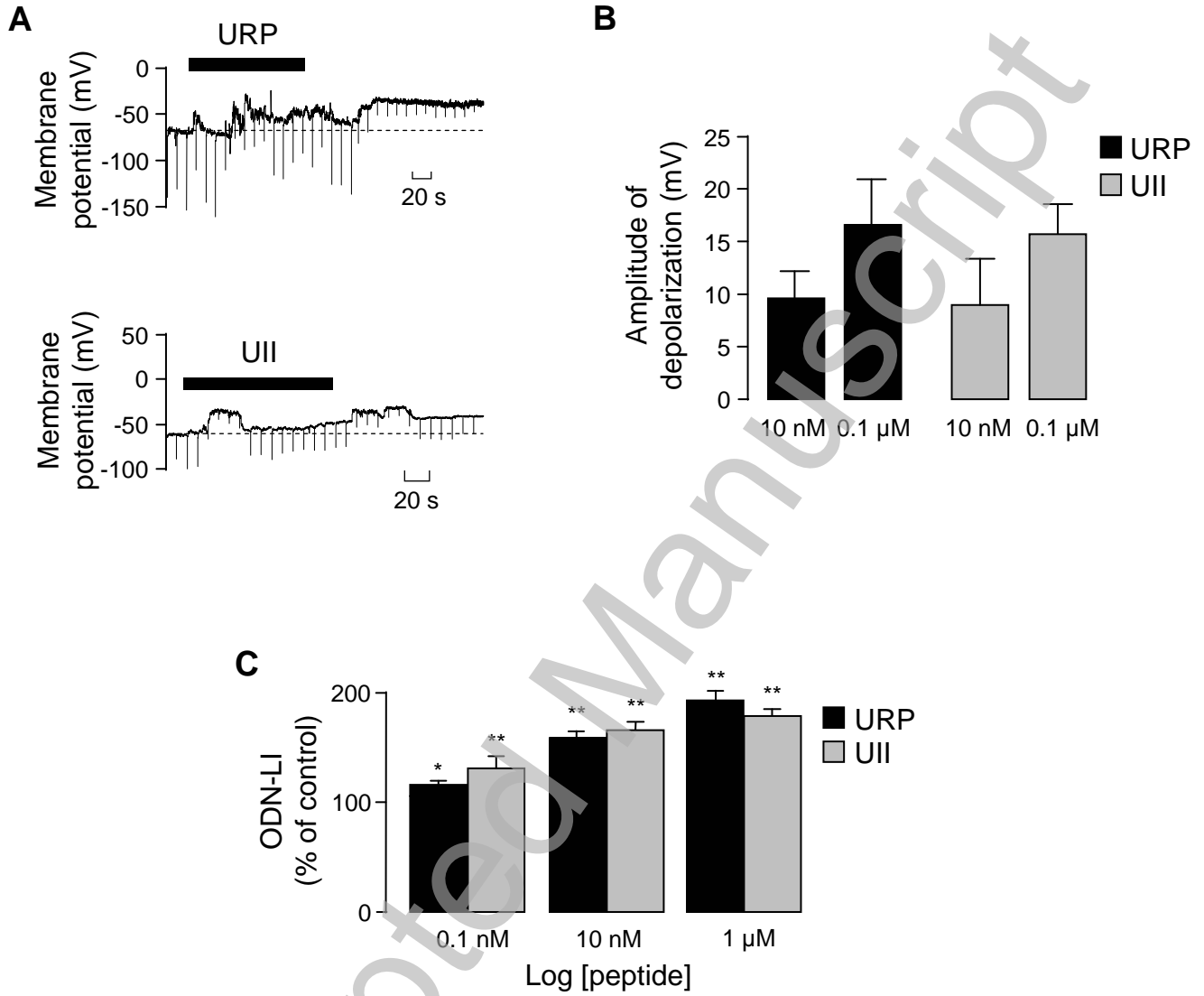


Figure 3
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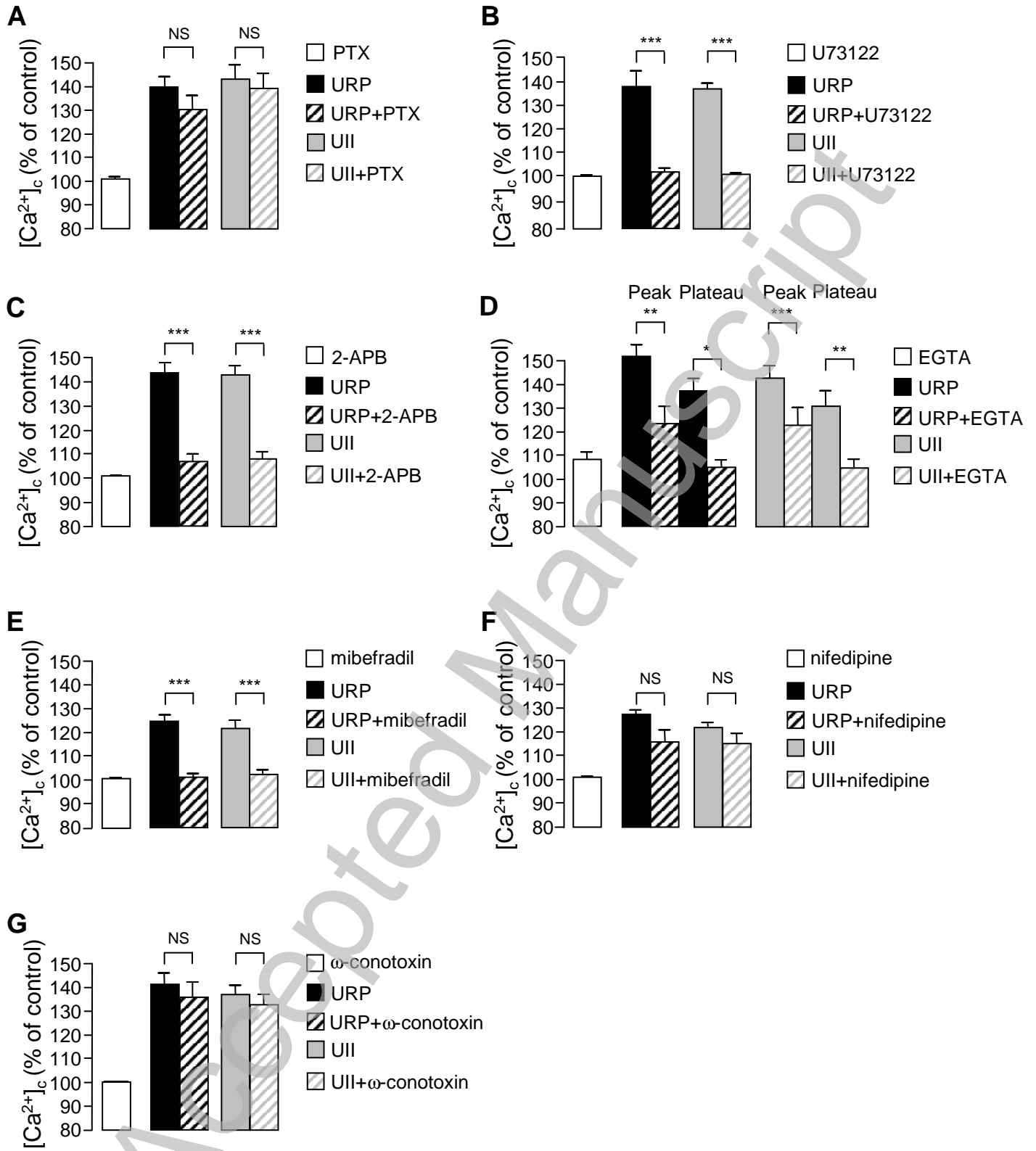


Figure 4
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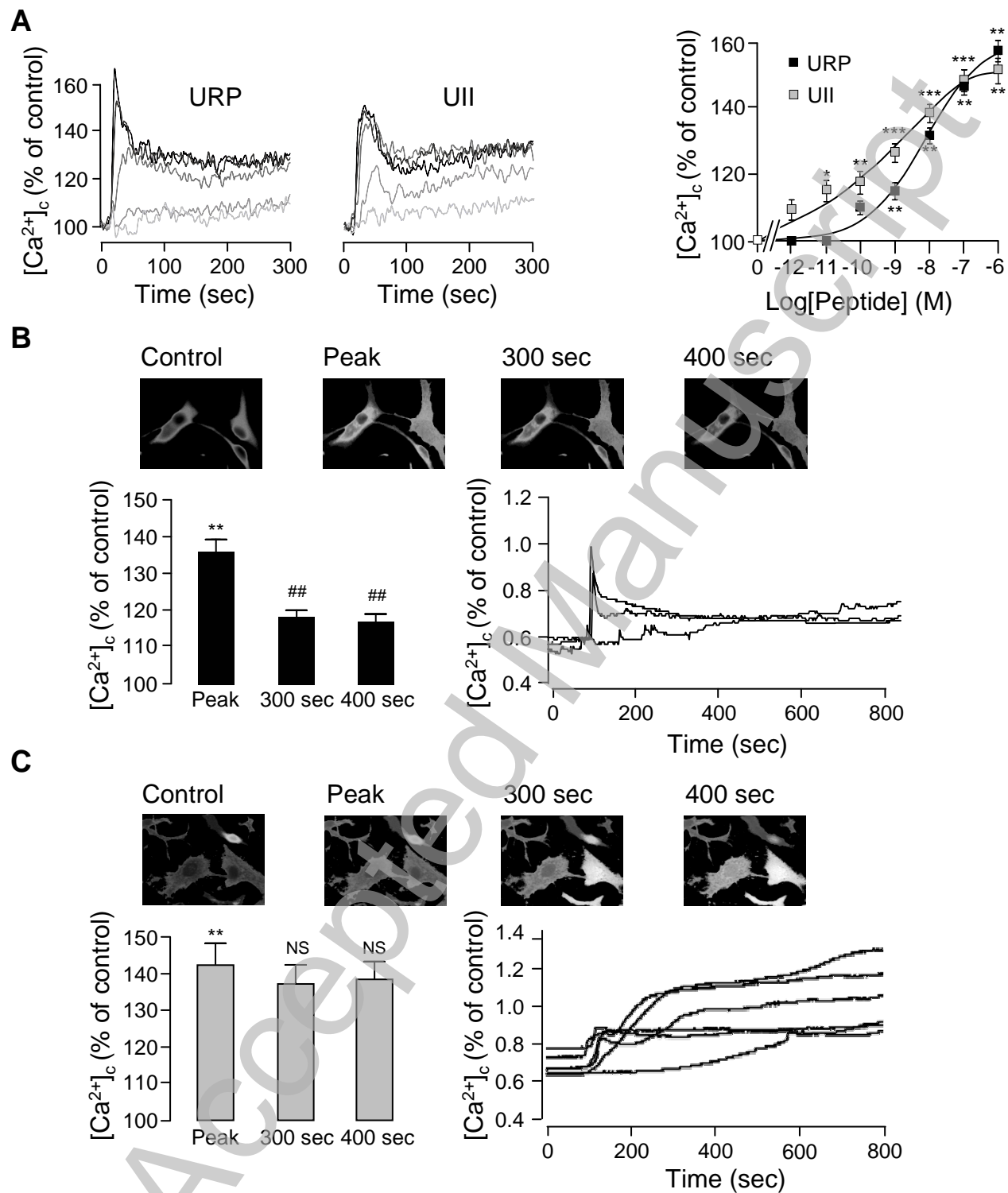


Figure 5
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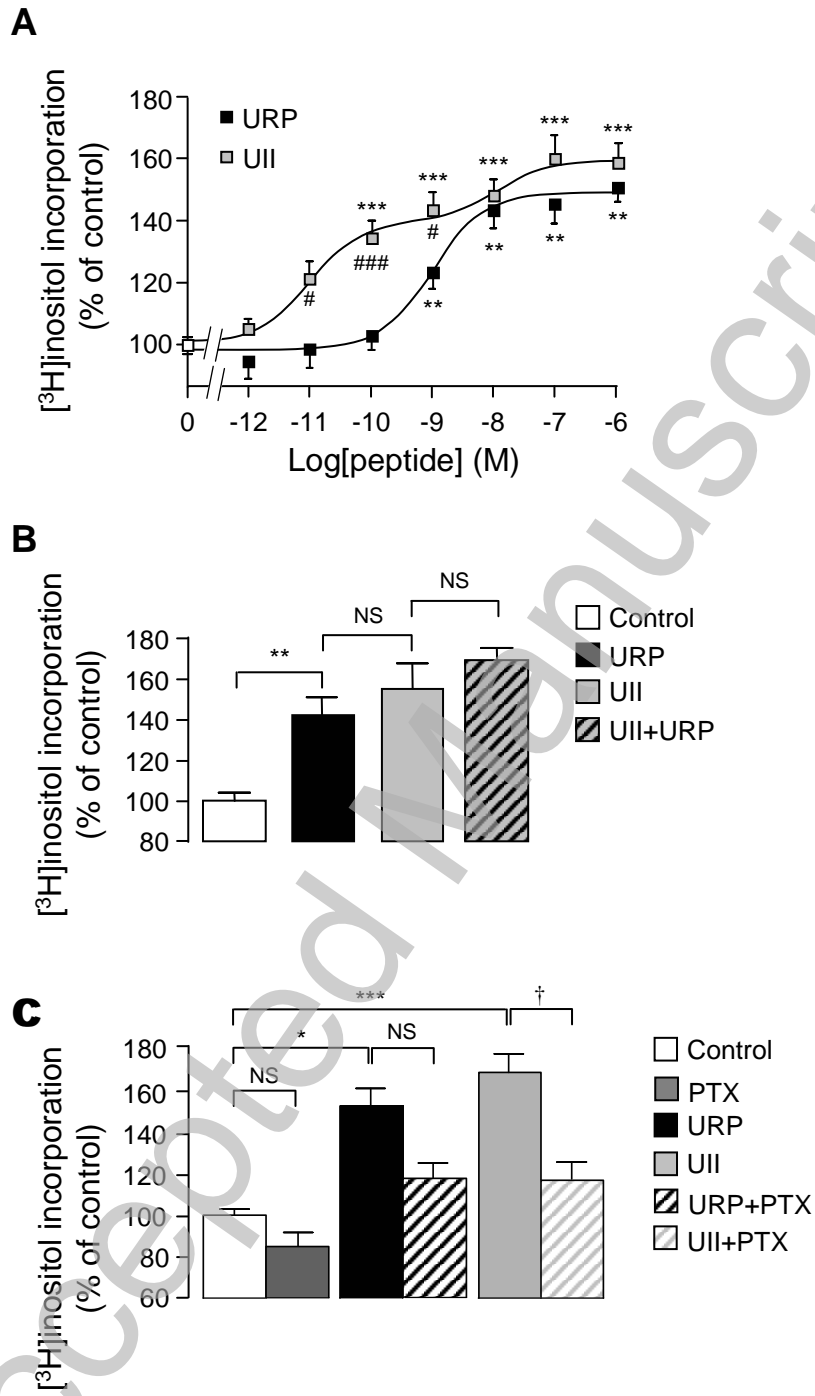
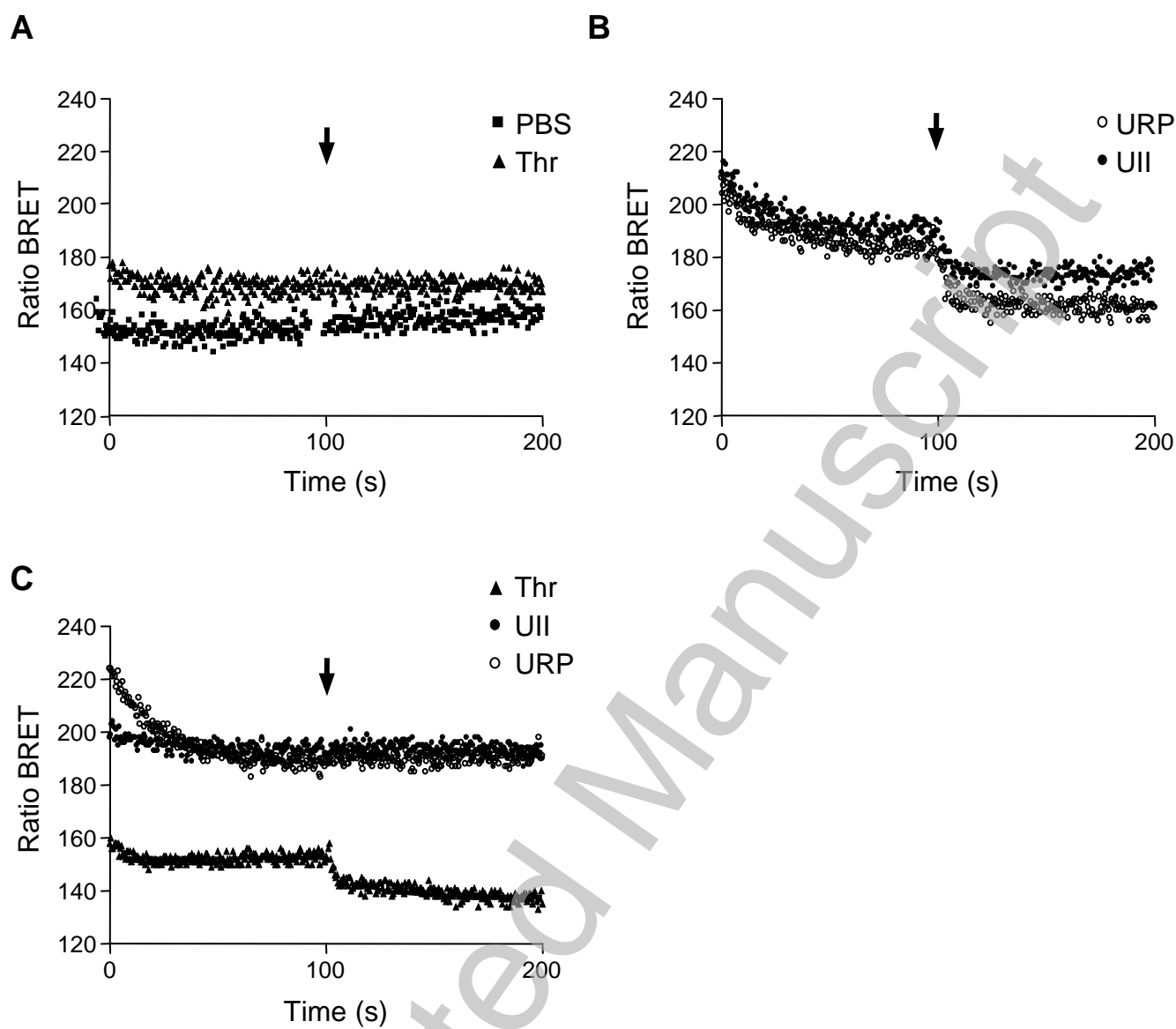


Figure 6
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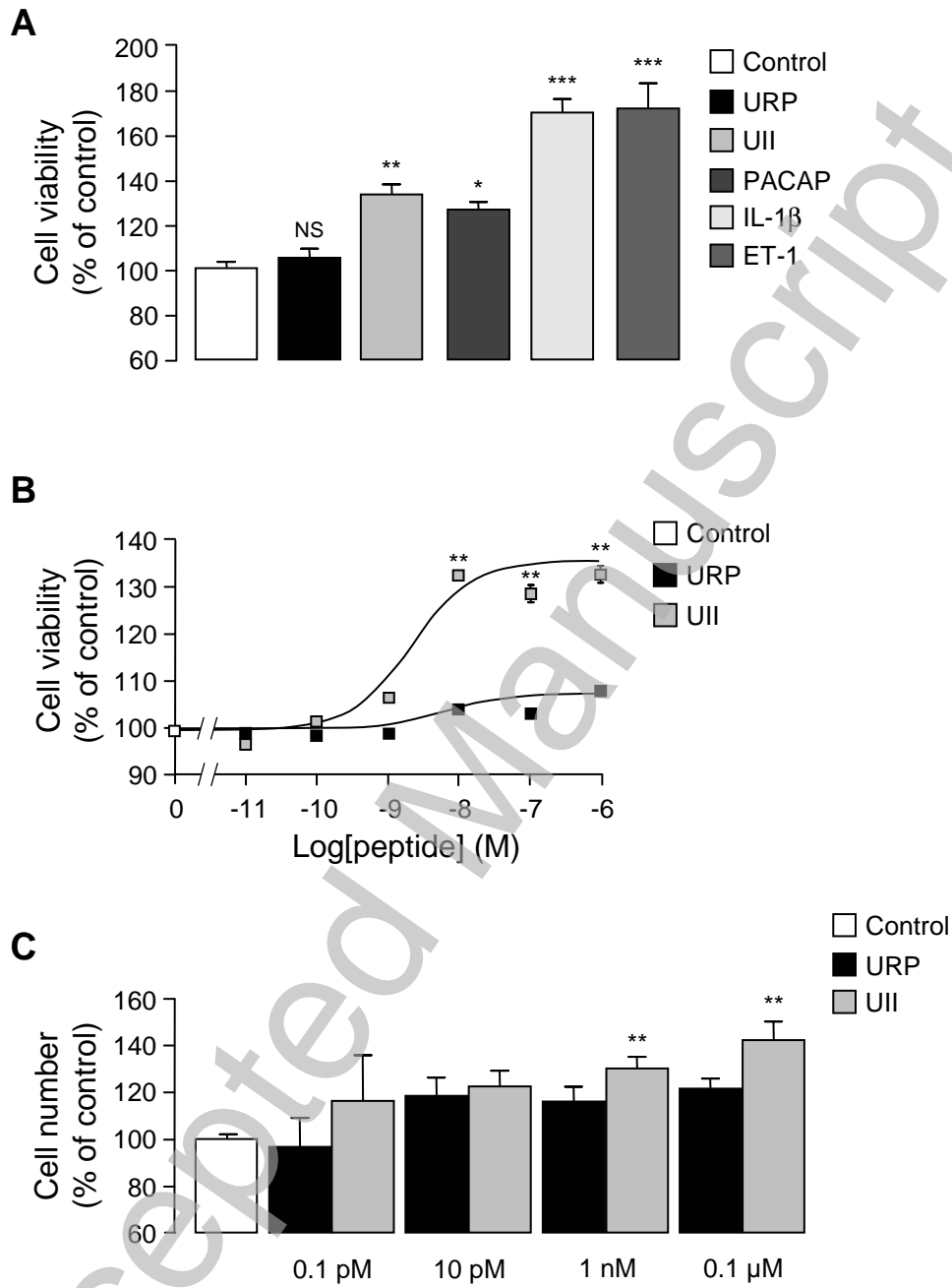


Figure 8
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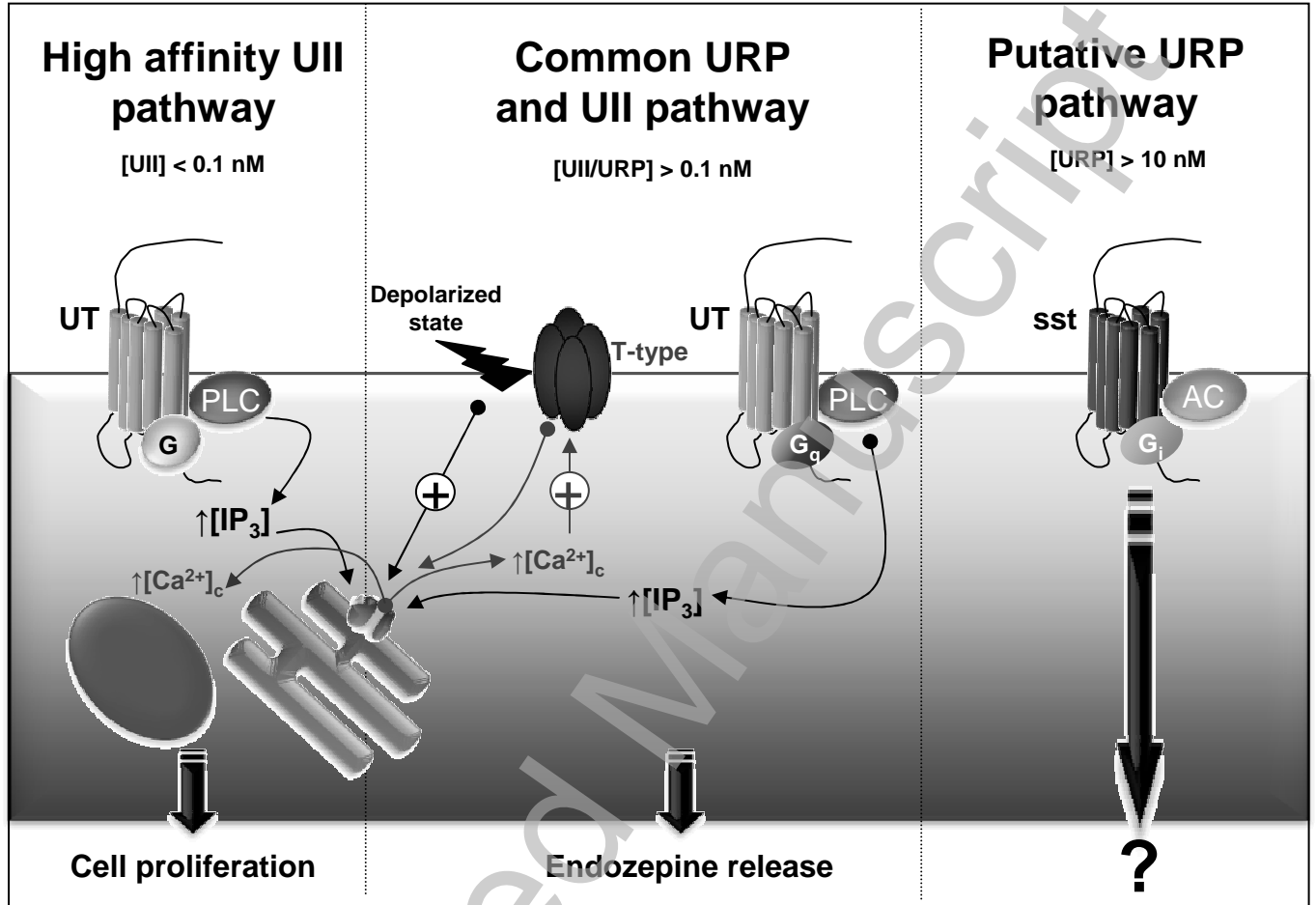


Figure 9
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