



HAL
open science

Parallel heterodyne detection of dynamic light scattering spectra from gold nanoparticles diffusing in viscous fluids.

Michael Atlan, Pierre Desbiolles, Michel Gross, Maite Coppey-Moisan

► **To cite this version:**

Michael Atlan, Pierre Desbiolles, Michel Gross, Maite Coppey-Moisan. Parallel heterodyne detection of dynamic light scattering spectra from gold nanoparticles diffusing in viscous fluids.. Optics Letters, 2010, 35 (5), pp.787-789. 10.1364/OL.35.000787 . hal-00463656

HAL Id: hal-00463656

<https://hal.science/hal-00463656>

Submitted on 14 Mar 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Parallel heterodyne detection of dynamic light scattering spectra from gold nanoparticles diffusing in viscous fluids.

Michael Atlan,¹ Pierre Desbiolles,² Michel Gross,² and Maïté Coppey-Moisan³

¹*Institut Langevin, CNRS UMR 7587, INSERM U 979, Fondation Pierre-Gilles de Gennes, Universités Paris 6 & 7, ESPCI ParisTech, 10 rue Vauquelin, 75005 Paris, France.*

²*Laboratoire Kastler-Brossel de l'École Normale Supérieure, CNRS UMR 8552, Université Pierre et Marie Curie - Paris 6, 24 rue Lhomond 75231 Paris cedex 05. France*

³*Département de Biologie Cellulaire, Institut Jacques Monod, CNRS UMR 7592, Université Paris 7, 2 Place Jussieu, Tour 43, 75251 Paris Cedex 05. France*

(Dated: March 26, 2010)

We developed a microscope intended to probe, using a parallel heterodyne receiver, the fluctuation spectrum of light quasi-elastically scattered by gold nanoparticles diffusing in viscous fluids. The cutoff frequencies of the recorded spectra scale up linearly with those expected from single scattering formalism in a wide range of dynamic viscosities (1 to 15 times water viscosity at room temperature). Our scheme enables ensemble-averaged optical fluctuations measurements over multispeckle recordings in low light, at temporal frequencies up to 10 kHz, with a 12 Hz framerate array detector. OCIS : 180.6900, 090.1995, 290.5820

Advances in dynamic light scattering microscopy [1–4] have recently raised the possibility of developing passive microrheology tools based on the measurement of local thermally-driven intensity fluctuations [5, 6]. The related homodyne measurement schemes involve usually single-mode detection [7–9]. Concurrently, heterodyne detection is a powerful method for investigating dynamic light scattering (DLS) in low-light conditions [10, 11]. In usual heterodyne detection schemes, the intensity fluctuations of the mix between the scattered field and a local oscillator (LO) field are measured on a single-mode detector. Since the probed scattered field matches the LO beam mode structure [12], such heterodyne schemes cannot perform multimode measurements. Multimode homodyne techniques based on intensity fluctuations measurements have been developed to take advantage of the spatial bandwidth of array detectors in order to perform speckle ensemble averaging and/or spatially-resolved measurements [13–17]. However, these parallel homodyne techniques suffer from a temporal frequency bandwidth imposed by the camera frame rate and from limited sensitivity in low light.

We present here a DLS microscope designed for probing the first-order Doppler spectrum of the light scattered by nanoparticles in Brownian motion. Our microscope performs heterodyne detection with a camera used as a multi pixel detector. The Doppler spectrum is explored by sweeping the LO beam optical frequency, so that the temporal frequency bandwidth is not limited by the camera frame rate.

The optical configuration is sketched in Fig.1. The whole optical apparatus has been described elsewhere [18]. A laser beam ($\lambda_L = 532$ nm, field E_L , angular frequency ω_L , average power 50 mW, single axial mode, CW) is split into tunable-frequency local oscillator (LO) and illumination arms (fields E_{LO} and E_I) to

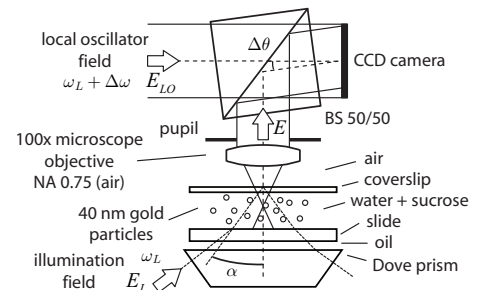


FIG. 1: Optical configuration. A colloidal gold suspension in a calibrated viscous fluid is illuminated by single mode laser light. The total internal reflection at the top interface ensures a dark-field illumination of the particles. The light field E scattered by the sample beats against a local oscillator field E_{LO} , detuned by $\Delta\omega$ with respect to the illumination field E_I . E_{LO} is tilted by $\Delta\theta$ with respect to E . The interference pattern $|E + E_{LO}|^2$ is recorded by a CCD camera.

form a Mach-Zehnder interferometer. The samples are thin chambers (thickness $h \sim 100 \mu\text{m}$, several millimeters wide) containing an aqueous solution of gold particles (diameter $d = 40$ nm : BBI international, EM.GC40) with added sucrose to increase the viscosity. The particle density is $\rho \sim 2.0 \times 10^{16} \text{ m}^{-3}$. At $\lambda_L = 535$ nm, the integral scattering cross-section σ_s of a 40 nm gold particle normalized by the particle volume v is $\sigma_s/v \sim 5.2 \mu\text{m}^{-1}$ [19]. Since the optical mean free path is $l = (\rho\sigma_s)^{-1}$, the ratio of the sample thickness to the mean free path is thus about $h/l \sim 3.5 \times 10^{-4}$, optical fluctuations are interpreted with single scattering formalism in dilute suspensions [10]. A Dove prism guides the illumination field E_I to the sample. Microscope immersion oil between the prism and the slide enables refractive index matching. To prevent the collection of the unscattered field, a total internal reflection is provoked at the glass-air in-

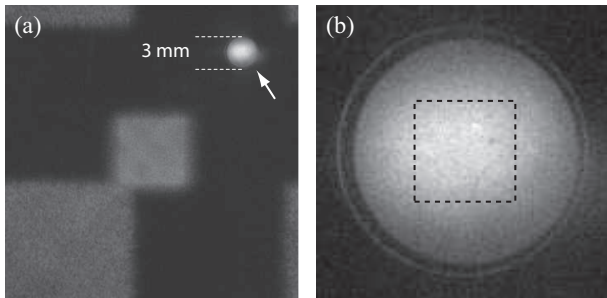


FIG. 2: (a) Hologram (1024×1024 pixels) calculated in the exit plane of the objective (arrow). (b) Zoom (141×141 pixels) of the objective pupil.

interface on top of the sample. The incident field makes an angle $\alpha \sim 50^\circ$ in water with respect to the optical axis. The field scattered by the gold beads $E \ll E_{\text{LO}}$ is collected by a microscope objective (Leica PL Fluotar: $100 \times$ magnification, $\text{NA} = 0.75$, air, exit pupil diameter $\simeq 3$ mm). The LO linear polarization is tuned to match the dominant polarization axis of the object field. Off-axis optical mixing ($\Delta\theta = 0.08$ rad tilt) of E and E_{LO} with a beam splitter (BS) results in a hologram $I = |E + E_{\text{LO}}|^2$ that is recorded with a CCD camera (PCO Pixelfly QE, 1392×1024 pixels of $d = 6.45 \mu\text{m}$, frame rate $\omega_{\text{S}}/(2\pi) = 12$ Hz, exposure time $\tau_{\text{E}} = 83$ ms) from which only the central $N_x \times N_y = 1024 \times 1024$ pixels region is used for the calculations.

The DLS spectra are acquired by sweeping the detuning frequency $\Delta\omega$ of the LO. At each frequency point $\Delta\omega$, 16 successive CCD frames $\{I_1, \dots, I_{16}\}$ are recorded. The field component E in the detector plane is calculated from the difference of consecutive images $I_{n+1} - I_n$. This quantity is back-propagated numerically up to the exit plane of the microscope objective by a standard convolution method [20] with a reconstruction distance $D = 34.9$ cm. A representation of the squared amplitude of the back-propagated hologram is shown in Fig.2 (a). Because of the angular tilt $\Delta\theta$ between the interfering beams, the image of the objective is shifted in the upper right corner of the hologram. Fig.2 (b) shows a zoom of the objective pupil image. The heterodyne signal $s_1 \propto |E|^2$ is averaged within the square region shown in Fig.2 (48×48 pixels with pixel size $\lambda D/(N_x d) = 28 \mu\text{m}$).

In the time domain, the normalized autocorrelation function g_{1d} of the fluctuating object field in the detector plane is [10] :

$$g_{1d}(q, \tau) = \exp(-Dq^2\tau) = \exp(-\tau/\tau_c), \quad (1)$$

where τ_c is the relaxation time, q is the scattering vector (momentum transfer), and D is the particle diffusivity. Since the beads undergo Brownian motion, the average mean-square displacement of each particle at time t scales as $\sim Dt$, where the diffusion constant D is linked to the thermal energy $k_B T$ and to the mobility μ of the parti-

cles: $D = \mu k_B T$. In the limit of low Reynolds number $\mu = 1/(6\pi\eta r)$, where η is the viscosity of the medium and r is the (hydrodynamic) radius of the spherical particles. The illumination and collection geometry introduce momentum transfer diversity, and because we seek to study dynamic fluctuations in non-varying optical scattering conditions, we retain

$$g_{1d}(\tau) \approx \exp(-Dak^2\tau) \quad (2)$$

where $k = 2\pi/\lambda_L$ is the angular wavenumber and a is an adjustment parameter used to describe the scattering structure factor of the optical configuration. Additionally, the electric field E that reaches the detector is a superposition of a field E_s statically scattered and a field E_d dynamically scattered by the sample [21, 22]. The normalized autocorrelation function g_1 of the total field $E = E_d + E_s$ is [21] :

$$g_1(\tau) = (1 - \rho) |g_{1d}(\tau)| + \rho, \quad (3)$$

where $\rho = |E_s|^2 / (|E_d|^2 + |E_s|^2)$. According to the Wiener-Khinchin theorem, the first-order power spectral density $s_1(\omega)$ of the field fluctuations is the Fourier transform of the field autocorrelation function $g_1(\tau)$. Let $s_{1d}(\omega)$ be the Fourier transform of $|g_{1d}(\tau)|$. Taking into account the instrumental response B in the frequency domain, we have :

$$s_1(\omega) = (1 - \rho) s_{1d}(\omega) * B(\omega) + \rho B(\omega), \quad (4)$$

where $*$ is the convolution product. In the absence of dynamic scattering, the off-axis component of the recorded image I_n at time $t_n = 2n\pi/\omega_{\text{S}}$ is the interference term $EE_{\text{LO}}^* \exp(i\Delta\omega t_n)$. The measured signal $I_n^{\text{off-axis}}$, integrated over an exposure time τ_{E} is :

$$I_n^{\text{off-axis}} = \frac{1}{\tau_{\text{E}}} \int_0^{\tau_{\text{E}}} EE_{\text{LO}}^* \exp[i\Delta\omega(t_n + \tau)] d\tau \quad (5)$$

where $i^2 = -1$. The normalized apparatus lineshape $B(\Delta\omega) = |I_{n+1}^{\text{off-axis}} - I_n^{\text{off-axis}}|^2 / (16|E|^2|E_{\text{LO}}|^2)$ is then :

$$B(\Delta\omega) = (\Delta\omega\tau_{\text{E}})^{-2} \sin^2(\Delta\omega\tau_{\text{E}}/2) \sin^2(\pi\Delta\omega/\omega_{\text{S}}). \quad (6)$$

This function is plotted on fig. 3 (line 5). Hence, in good approximation, frequency shifts $\Delta\omega$ probe actual fluctuations frequencies ω . The normalized first-order power spectrum of the fluctuating field is the Fourier transform of equation 1 :

$$s_{1d}(\omega) = 1/(1 + \omega^2\tau_c^2), \quad (7)$$

where $\tau_c = 6\pi\eta r/(q^2 k_B T)$. Experimentally, the samples must be prepared with great care to avoid stray light. Otherwise, the estimation of the cutoff frequency τ_c^{-1} is made difficult because both fluctuating and non fluctuating light are recorded within the available CCD dynamic

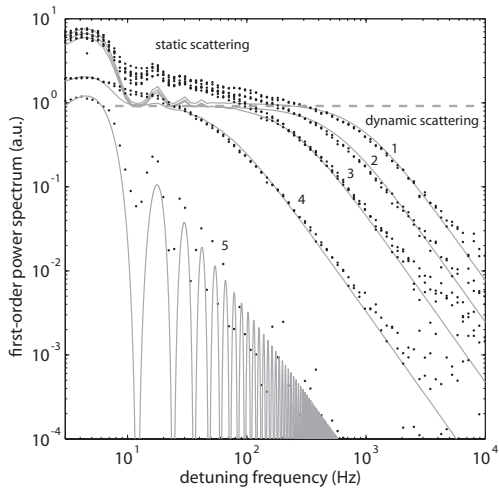


FIG. 3: First-order light fluctuations spectra normalized by the relative weight of dynamically scattered light. The relative viscosities of the samples [23] are $\eta/\eta_0 = 1.00$ (1), 1.69 (2), 3.94 (3), and 15.40 (4). Continuous lines are plotted from Eq.4. Data points (5) represent the apparatus lineshape in the frequency domain, and the continuous line is plotted from Eq.6. The horizontal axis is the detuning frequency $\Delta\omega/(2\pi)$.

range. A high relative weight of static light prevents an accurate detection at high frequencies because it reduces the effective dynamic range.

The measured spectra from samples of relative viscosity $\eta/\eta_0 = 1.00$ (1), 1.69 (2), 3.94 (3), and 15.40 (4) are represented in Fig.3, where $\eta_0 = 10^{-3}$ Pa.s is pure water viscosity. For each measurement, the frequency is detuned from 3 Hz to 10^4 Hz over 80 points with logarithmic spacing. Each scan lasts ~ 106 s and 5 minutes are needed to save the data. The frequency scans are performed twice with 15 minutes intervals and signal variability from one measurement to another is visible at 10^{-3} from the maximum typically (points clouds). The s_1 lines plotted in Fig.3 are calculated from Eq.4, with $k_B T = 4.11 \times 10^{-21}$ J at room temperature (298 K), $r = 40/2 = 20$ nm, and $q^2 = 3.7k^2$. The adjustment parameter $a = 3.7$ has been chosen to get a good agreement with the experimental data points. The expected momentum transfer from particle scattering of the incident and reflected beams, in DLS conditions [10] is $q^2 = 4n_w^2 \sin^2(\beta/2) k^2$. $n_w = 1.33$ is the refractive index of water. For the incident beam, we have $q^2 = 1.3k^2$ ($\beta = \alpha = 50^\circ$). For the reflected beam, we have $q^2 = 5.8k^2$ ($\beta = \pi - \alpha = 180 - 50^\circ$). The derived cutoff frequencies are $\tau_c^{-1} = 888$ Hz (1), 524 Hz (2), 225 Hz (3), 58 Hz (4); these cutoffs are inversely proportional to the viscosity and spread over a wide frequency range. The values of the relative weight of the dynamic light component for the different preparations used to fit $s_1(\omega)$ to the experimental data are $1 - \rho = 0.48$ (1), 0.15 (2), 0.14 (3), 0.13 (4), 0.0 (5). Data points (5) of Fig.3

are obtained from a preparation without Brownian particles; they are in agreement with the apparatus lineshape derived from Eq.6.

These results demonstrate the suitability of the proposed method for parallel heterodyne dynamic light scattering measurements of nanoparticles dynamics in microscopic volumes. The resolution, range and sensitivity of first-order spectrum measurements potentially enable a robust exploration of a wide range of viscosity-dependent fluctuations at microscopic scales.

The authors acknowledge support from Agence Nationale de la Recherche (ANR-09-JCJC-0113 grant), Fondation Pierre-Gilles de Gennes (FPGG014 grant), Région Ile-de-France (C’Nano grant) and CNRS.

-
- [1] T. Maeda and S. Fujime, *Review of Scientific Instruments* **43**, 566 (2003).
 - [2] I. Nishio, J. Peetermans, and T. Tanaka, *Cell Biochemistry and Biophysics* **7**, 91 (1985).
 - [3] P. Blank, R. Tishler, and F. Carlson, *Applied Optics* **26**, 351 (1987).
 - [4] R. Bar-Ziv, A. Meller, T. Tlusty, E. Moses, J. Stavans, and S. Safran, *Physical Review Letters* **78**, 154 (1997).
 - [5] P. Kaplan, V. Trappe, and D. Weitz, *Applied Optics* **38** (1999).
 - [6] T. Mason, H. Gang, and D. Weitz, *Journal of Molecular Structure* **383**, 81 (1996).
 - [7] R. Brown, *Applied Optics* **26**, 4846 (1987).
 - [8] J. Ricka, *Appl. Opt.* **32**, 2860 (1993).
 - [9] T. Gisler, H. Ruger, P. Schurtenberger, S. U. Egelhaaf, J. Tschumi, and J. Ricka, *Appl. Opt.* **34**, 3546 (1995).
 - [10] B. J. Berne and R. Pecora, *Dynamic Light Scattering* (Dover, 2000).
 - [11] R. Hohler, V. Labiausse, and S. Cohen-Addad, *JOSA A* **20**, 2179 (2003).
 - [12] S. Cohen, *Applied Optics* **14**, 9153 (1975).
 - [13] A. P. Y. Wong and P. Wiltzius, *Review of Scientific Instruments* **64**, 2547 (1993).
 - [14] S. Kirsch, V. Frenz, W. Schartl, E. Bartsch, and H. Sillescu, *The Journal of Chemical Physics* **104**, 1758 (1996).
 - [15] L. Cipelletti and D. A. Weitz, *Review of Scientific Instruments* **70**, 3214 (1999).
 - [16] V. Viasnoff, F. Lequeux, and D. J. Pine, *Review of Scientific Instruments* **73**, 2336 (2002).
 - [17] F. Scheffold and R. Cerbino, *Current Opinion in Colloid and Interface Science* (2007).
 - [18] M. Atlan, M. Gross, P. Desbiolles, Emilie Absil, G. Tessier, and M. Coppey-Moisand, *Optics Letters*. **33**, 500 (2008).
 - [19] P. Jain, K. Lee, I. El-Sayed, and M. El-Sayed, *J. Phys. Chem. B* **110**, 7238 (2006).
 - [20] U. Schnars and W. Juptner, *Appl. Opt.* **33**, 179 (1994).
 - [21] D. Boas and A. Yodh, *J. Opt. Soc. Am. A* **14**, 192 (1997).
 - [22] P. Zakharov, A. Volker, A. Buck, B. Weber, and F. Scheffold, *Opt. Lett.* **31**, 3465 (2006).
 - [23] D. Lide, *CRC handbook of chemistry and physics* (CRC Press, 2004).