



HAL
open science

Parasitism in Kansas in the 1800s - A glimpse to the past through the analysis of grave sediments from Meadowlark cemetery.

Matthieu Le Bailly, Marcelo L.C. Gonçalves, Christine Lefevre, Donna C. Roper, Jeremy W. Pye, Aduino Araujo, Françoise Bouchet

► **To cite this version:**

Matthieu Le Bailly, Marcelo L.C. Gonçalves, Christine Lefevre, Donna C. Roper, Jeremy W. Pye, et al.. Parasitism in Kansas in the 1800s - A glimpse to the past through the analysis of grave sediments from Meadowlark cemetery.. Memórias do Instituto Oswaldo Cruz, 2006, 101 (Suppl. II), pp.53-56. 10.1590/s0074-02762006001000009 . hal-00451144

HAL Id: hal-00451144

<https://hal.science/hal-00451144v1>

Submitted on 27 Sep 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Parasitism in Kansas in the 1800s – A glimpse to the past through the analysis of grave sediments from Meadowlark cemetery

Matthieu Le Bailly/⁺, Marcelo LC Gonçalves*, Christine Lefèvre**, Donna C Roper***, Jeremy W Pye****, Adauto Araujo*, Françoise Bouchet

Université de Reims, UFR de Pharmacie, EA 3798, Laboratoire de Paléoparasitologie, CNRS UMR 5197, 51, rue Cognacq-Jay, 51 096 Reims cedex, France *Escola Nacional de Saúde Pública-Fiocruz, Rio de Janeiro, RJ, Brazil **Muséum National d'Histoire Naturelle, Paris cedex 05, France ***Department of Sociology/Anthropology/Social Work, Kansas State University, Manhattan, KA, US ****Department of Anthropology, University of Arkansas, Fayetteville, AK, US

During the excavations of the XIX century Meadowlark cemetery (Manhattan, Kansas, US), samples of sediments were taken from around five skeletons, and analyzed to detect intestinal parasites. No helminth eggs were found, but immunological ELISA tests for Entamoeba histolytica were positive in three samples. The immunological techniques have been successfully used in paleoparasitology to detect protozoan infections. Amoebiasis could have been a severe disease in the past, especially where poor sanitary conditions prevailed, and there is evidence that this cemetery may have been used in a situation where poor sanitary conditions may have prevailed. The presence of this protozoan in US during the late XIX century gives information on the health of the population and provides additional data on the parasite's evolution since its appearance in the New World.

Key words: paleoparasitology - ancient diseases - paleoepidemiology - protozoal infection - amoebiasis

Scientists have been using different techniques to look for parasites in ancient material (Reinhard et al. 1987, Araujo et al. 1998, Bouchet et al. 2003a). Coprolites or desiccated feces, or fecal remains mixed with soil sediment are commonly found in archaeological excavations. The parasitological analysis of such material yields rich information about paleopathologies, human migrations, cultural changes, dietary habits, paleoclimates, parasite dispersion, and host migrations throughout space and time (Reinhard et al 2001, Bouchet et al. 2003b, Gonçalves et al. 2003).

MATERIALS AND METHODS

The analyzed sediments were collected during the excavation of the Meadowlark Cemetery, on the grounds of Meadowlark Hills Retirement Community in Manhattan, Kansas. This cemetery was used during the period ca. 1860 to 1900. It was excavated during the summer and early fall of 2004 to make way for construction of new residences on the retirement community property. A total of 17 graves were revealed. Thirteen of these graves contained skeletons. Sediment samples from the abdominal region of five skeletons were studied: MC F9, F10, F12, F13, and F17 (Table).

The material was submitted to the standard paleoparasitological examination using an optical microscope to

search for helminth eggs. A commercially available enzyme-linked immunosorbent assay (ELISA) with monoclonal antibodies was used to test for antigen from *Entamoeba histolytica* (Haque et al. 1995, 2000) and *Giardia duodenalis* (Aldeen et al. 1998, Boone et al. 1999), two common intestinal protozoan parasites.

For the standard paleoparasitological examination, the samples were rehydrated for a week in 0.5% trisodic phosphate and 5% glycerinated solution. The material was then crushed in a mortar. The suspension was submitted to an ultrasound device for 1 min at 50°C, on a frequency of 35 kHz, and the suspension strained through 315- μ m, 160- μ m, 50- μ m, and 25- μ m meshes. A portion of each sample was used for microscopic examination. The material was placed on a slide that was covered with a coverslip and examined for evidence of parasites and food remains. Twenty slides of each sample were examined at magnification of $\times 100$ and $\times 400$ (Bouchet et al. 2001).

For the ELISA tests, the *E. histolytica* II and the *Giardia* II assay (TechLab, Blackburg, VA, US) were used. Sample preparation was quite different to provide physicochemical parameter modifications, especially due to formalin (Hould 1984, Exbrayat 2000). The samples were rehydrated for one week in ultra-pure water obtained from a purification system manufactured by Millipore® (Direct-Q™ 5). Samples were preserved in the freezer (2-4°C) during the rehydration phase to avoid bacteria and fungi development. Manufacturer's directions were followed and results were interpreted by visual inspection and using a spectrophotometer. A test sample was considered positive if it had an obvious yellow color when compared to the negative control well. A test sample was considered negative if it was colorless or less yellow than the negative control well. The absorbance of each specimen was also measured at 450 nm wavelength (Haque et al. 2000).

Financial support: CNRS; Capes/Cofecub, CNPq, TechLab®, Fumouze, Millipore®

⁺Corresponding author: matthieu.lebailly@univ-reims.fr

Received 20 July 2006

Accepted 16 October 2006

RESULTS

Microscopic examination revealed no helminth eggs resulting from an infestation of any one of the various parasitic worm species (including roundworm and hookworm) in any of the five analyzed samples. Samples MC F10, F12, and F13 were positive for the presence of the *E. histolytica* antigen, the causal agent of amoebic dysentery and amoebic liver abscess (Table). All samples were negative for the *G. duodenalis* antigen.

Parasites are biological markers that can be used to track ancient host migrations (Araújo et al. 2003). The finding of parasite remains in archaeological material is a tool to trace the evolution of past infections worldwide. Well-preserved parasite eggs or protozoan antigen recovered by immunoassays techniques are now beginning to reveal the origin and evolution of infectious diseases in the past (Bouchet et al. 2002, Le Bailly et al. 2003a, Ortega & Bonavia 2003)

DISCUSSION

The former species *E. histolytica* was separated in 1993 into two species, *E. dispar*, a non-pathogenic intestinal parasite and *E. histolytica*, which can cause dysentery and extra-intestinal diseases in humans (Diamond & Clark 1993). Although microscopy cannot differentiate between them, isoenzyme analysis and specific molecular techniques are able to distinguish *E. histolytica* from *E. dispar* (Haque et al. 1998, Huston et al. 1999).

E. histolytica had previously been found in archaeological material using the immunoassay technique (Gonçalves et al. 2004) on samples from other sites. The older traces were found in samples from Switzerland dated from around 5300 years BP (Gonçalves et al. 2004). Although protozoan cysts are not as resistant to decay as are helminth eggs, cyst antigens can remain stable for several centuries. Immunoassays using monoclonal antibodies to detect such antigens are very specific and are a reliable tool in paleoparasitology (Gonçalves et al. 2002, 2004). Amoebiasis is a parasitic disease related to contamination of drinking water and food polluted with feces, reflecting poor sanitation conditions wherever it is found.

It is possible to infer sanitation conditions and even lifestyle in ancient human settlements through the recovery and identification of parasites in archaeological material (Reinhard 1992, Araujo et al. 1998, Le Bailly et al. 2003a). Paleoparasitological studies in Neolithic settlements (Bouchet 1997, Bouchet et al. 1997, Le Bailly et al. 2003b, 2005), ancient Roman fortifications, and medieval villages

(Bouchet et al. 2000) have revealed a picture of the intensive relationship between parasites and humans in the Old World.

Although many intestinal parasites infected New World populations before contact, after colonization, these parasitoses seem to have become a major public health problem. As already noted, parasites are common where humans live in poor sanitary conditions. The parasitic agents most commonly are transmitted through contaminated food or water sources, as well as poor waste management and education about proper cleanliness. Due to the factors of transmission listed above, often individuals of low socio-economic status fall prey to such parasites. This was particularly the case in the XIX century.

Health and evidence of stress are largely a reflection of quality of life, and therefore are strongly related to socio-economic status. While there are degrees of environmental factors involved in the assessment of quality of life, such as pathological stressors, they are often tied into culturally created factors such as sanitation, access to resources, and education (Goodman et al. 1988). Thus, the identification of particular markers, which can elucidate the relationship between status and quality of life, such as the analysis of parasites, are important (Larsen 1997).

At the outset of the excavations of Meadowlark Cemetery, the researchers learned that William H Stillman had owned the land on which the cemetery was formed. A local informant, whose family lived on the property since 1915, claimed that Stillman was running the county poor farm and an orphanage in the late XIX century (Pye et al. 2004). A Manhattan brochure printed in 1888 AD states that an orphan home was arranging to relocate to Manhattan, however, no concrete evidence of this occurring has been found (Board of Trade – Manhattan, Kansas 1888). If an orphanage existed on this property during the use life of the cemetery, then one would expect to see a large proportion of subadults present, as well as a high incidence of infection due to low bacterial and parasitic thresholds. In fact, the largest number of burials was those of subadults.

Six burials were those of adults, four of them young adult males, one an elderly male, and one an adult female. This is consistent with a possibility put forth before the excavation began that this cemetery was used for burial of residents of a poor farm. Poor farms at this time in the US were established to provide care and work for the indigent and a means of subsistence for young laborers, often at the lowest possible cost to the federal or town gov-

TABLE

Characteristics of grave features and burials at the Meadowlark Cemetery, and positivity for *Entamoeba histolytica* ELISA test

Burials	Age	Sex	Coffin/Grave length	<i>E. histolytica</i> antigen
Feature 9	Subadult < 1yr	Undetermined	107.82 cm/ 139.12 cm	Negative
Feature 10	Subadult	Undetermined	154 cm/ 134.65 cm	Positive
Feature 12	Subadult	Undetermined	141.66 cm	Positive
Feature 13	Adult 18-25 yrs	Male	211.12 cm	Positive
Feature 17	Subadult	Undetermined	100.69 cm/ 122.58 cm	Negative

ernment (Bell 1990). There is some evidence that Stillman may have been receiving money from the county for care of the poor (*Manhattan Enterprise*, 8/13/1880), although there are no specific records that he ran a poor farm. By definition, orphans and poor migrant workers would not usually be able to attain high social status in the society, and therefore might also have been exposed to high levels of parasitic agents.

Conclusion

The finding of *E. histolytica* in organic sediment from the Meadowlark Cemetery reflects a crowded area and poor sanitation conditions in this particular Kansas community in the late XIX century. *G. duodenalis* was not found in the material. Since *G. duodenalis* is commonly found in ecosystems similar to those where *E. histolytica* is usually found, this could reflect a true absence of *G. duodenalis* or be a false negative result. However, caution should be exerted in the interpretation of negative results in ancient samples, as antigen structure may be compromised in the uncontrolled preservation conditions. In any event, the results obtained during the study of the Meadowlark Cemetery sediment samples should encourage others to use various immunological techniques, or immunological and molecular techniques together. Such techniques might include immunofluorescence or polymerase chain reaction-solution hybridization enzyme-linked immunoassay (Aguirre et al. 1995, 1997) that could be more sensitive for studying the evolution of the parasites genome.

REFERENCES

- Aguirre A, Molina S, Blotkamp C, Verweij J, Vinuesa T, Valls ME, Guhl F, Polderman A, Jimenez de Anta MT, Corachan M, Gonzalez-Ruiz A, Frame IA, Warhurst D 1997. Diagnosis of *Entamoeba histolytica* and *Entamoeba dispar* in clinical specimens by PCR-SHELA. *Arch Med Res* 28: 282-284.
- Aguirre A, Warhurst DC, Guhl F, Frame IA 1995. Polymerase chain reaction-solution hybridization enzyme-linked immunoassay (PCR-SHELA) for the differential diagnosis of pathogenic and non-pathogenic *Entamoeba histolytica*. *Trans R Soc Trop Med Hyg* 89: 187-188.
- Aldeen WE, Carroll K, Robinson A, Morrison M, Hale D 1998. Comparison of nine commercially available enzyme-linked immunosorbent assays for detection of *Giardia lamblia* in fecal specimens. *J Clin Microbiol* 36: 1338-1340.
- Araújo A, Reinhard K, Bastos OM, Costa LC, Pirmez C, Iñiguez A, Vicente AC, Morel CM, Ferreira LF 1998. Paleoparasitology: perspectives with new techniques. *Rev Inst Med Trop São Paulo* 40: 371-376.
- Bell EL 1990. The historical archaeology of mortuary behavior: coffin hardware from Uxbridge, Massachusetts. *Hist Archaeol* 24: 54-78.
- Board of Trade, Manhattan, Kansas 1888. Manhattan, Riley County, Kansas – Kansas (Manhattan Brochure/Map), Kansas State University Libraries, Special Collections, Manhattan, Kansas.
- Boone JH, Wilkins TD, Nash TE, Brandon JE, Macias EA, Jerriss RC, Lyerly DM 1999. TechLab and alexon *Giardia* enzyme-linked immunosorbent assay kits detect cyst wall protein 1. *J Clin Microbiol* 37: 611-614.
- Bouchet F 1997. Intestinal capillariasis in neolithic inhabitants of Chalais (Jura, France). *Lancet* 349 (9047): 256.
- Bouchet F, Bentrard S, Dommelier S, Paicheler JC, Pétrequin P 1997. Capillarioses intestinales: nématodoses du Néolithique. *Bull Soc Fr Parasitol* 15: 49-54.
- Bouchet F, Harter S, Paicheler JC, Araujo A, Ferreira LF 2002. First recovery of *Schistosoma mansoni* eggs from a latrine in Europe (15-16th centuries). *J Parasitol* 88: 404-405.
- Bouchet F, Harter S, Le Bailly M 2003a. The state of the art of paleoparasitological research in the Old World. *Mem Inst Oswaldo Cruz* 98 (Suppl. 1): 95-101.
- Bouchet F, Guidon N, Dittmar K, Harter S, Ferreira LF, Chaves SM, Reinhard K, Araújo A 2003b. Parasites remains in archaeological sites. *Mem Inst Oswaldo Cruz* 98 (Suppl. 1): 47-52.
- Bouchet F, West D, Lefèvre C, Corbett D 2001. Identification of parasitoses in child burial from Adak Island (Central Aleutian Islands, Alaska). *CR Acad Sc Paris (série III)* 324: 123-127.
- Diamond LS, Clark CG 1993. A redescription of *Entamoeba histolytica* Shaudinn, 1903 (amended Walker, 1911) separating it from *Entamoeba dispar* Brumpt, 1925. *J Eukariot Microbiol* 40: 340-344.
- Exbrayat JM 2000. *Méthodes Classiques de Visualisation du Génome en Microscopie Photonique*, Tec et Doc, Paris.
- Gonçalves MLC, Araujo A, Duarte R, Pereira da Silva J, Reinhard K, Bouchet F, Ferreira LF 2002. Detection of *Giardia duodenalis* antigen in coprolites using a commercially available enzyme-linked immunosorbent assay. *Trans R Soc Trop Med Hyg* 96: 640-643.
- Gonçalves MLC, Araujo A., Ferreira LF 2003. Human intestinal parasites in the past: new findings and a review. *Mem Inst Oswaldo Cruz* 98 (Suppl. 1): 103-118.
- Gonçalves M, Silva V, Andrade C, Rocha G, Le Bailly M, Bouchet F, Ferreira LF, Araujo A 2004. Amebiasis distribution in the past: first steps in using an immunoassay technique. *Trans R Soc Trop Med Hyg* 98: 88-91.
- Goodman AH, Thomas RB, Swedlund AC, Armelagos GJ 1988. Biocultural Perspectives on Stress in Prehistoric, Historical, and Contemporary Population Research. *Yearb Phys Anthropol* 21: 169-202.
- Haque R, Ali IKM, Akther S, Petri Jr WA 1998. Comparison of PCR, isoenzyme analysis, and antigen detection for diagnosis of *Entamoeba histolytica* infection. *J Clin Microbiol* 36: 449-452.
- Haque R, Mollah NU, Ali KM, Alam K, Eubanks A, Lyerly D, Petri Jr WA 2000. Diagnosis of amebic liver abscess and intestinal infection with the TechLab *Entamoeba histolytica*. II antigen detection and antibody tests. *J Clin Microbiol* 38: 3235-3239.
- Haque R, Neville LM, Hahn P, Petri WA 1995. Rapid diagnosis of *Entamoeba* infection by using *Entamoeba* and *Entamoeba histolytica* stool antigen detection kits. *J Clin Microbiol* 33: 2558-2561.
- Hould R 1984. *Techniques d'Histopathologie et de Cytopathologie*, Maloine, Paris.

- Huston CD, Haque R, Petri Jr WA 1999. Molecular-based diagnosis of *Entamoeba histolytica* infection. *Expert Rev Mol Med* 22: 1-11.
- Larsen CS 1997. *Bioarchaeology: Interpreting Behavior from the Human Skeleton*, Cambridge Studies in Biological Anthropology 21, Cambridge University Press, Cambridge.
- Le Bailly M, Harter S, Bouchet F 2003a. A l'interface de la biologie et de l'archéologie: la paléoparasitologie. *Archéopages* 11: 12-17.
- Le Bailly M, Leuzinger U, Bouchet F 2003b. *Dioctophymidae* eggs from coprolites from neolithic site of Arbon-Bleiche 3 (Switzerland). *J Parasitol* 89: 1073-1076.
- Le Bailly M, Leuzinger U, Schlichtherle H, Bouchet F 2005. *Diphyllobothrium*: neolithic parasite? *J Parasitol* 91: 957-959.
- Manhattan Enterprise 1880. *Issue of Manhattan Enterprise, August 13, 1880*, Kansas State Historical Society, Manhattan.
- Ortega YR, Bonavia D 2003. *Cryptosporidium*, *Giardia*, and *Cyclospora* in ancient Peruvians. *J Parasitol* 89: 635-636.
- Pye JW, Smith HC, Roper DC 2004. Excavations at Meadowlark Cemetery, Manhattan. *Curr Archaeol Kansas* 5: 77-92.
- Reinhard KJ 1992. Parasitology as an interpretive tool in archaeology. *Am Antiquity* 57: 231-245.
- Reinhard KJ, Araújo A, Ferreira LF, Coimbra JR CE 2001. American hookworm antiquity. *Med Anthropol* 20: 97-101.
- Reinhard KJ, Hevly RH, Anderson GA 1987. Helminth remains from prehistoric Indian Coprolites from the Colorado Plateau. *J Parasitol* 70: 630-639.