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Stable Thiol Groups

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The functionalization of superparamagnetic iron oxide nanoparticles (SPIOs) by meso-2,3-dimercaptosuccinic acid (DMSA) was investigated. Under ambient conditions, the thiol groups from DMSA are not stable and do not allow a direct functionalization without storage in stringent conditions or a chemical regeneration of free thiols. In this study, we have developed a protocol based on poly(ethylene glycol) (PEG) grafting of SPIO prior to DMSA anchoring. We have observed that PEGhelps to increase the stability of thiol groups under ambient conditions. The thiol functionalized SPIOs were stable under physiological pH and ionic strength as determined by Ellman's essay and allowed us to graft a thiol reactive fluorescent dye: tetramethylrhodamine-5maleimide (TMRM).

Superparamagnetic iron oxide (SPIO) nanoparticles have received great attention for a few decades in the fields of bioapplications such as early diagnostics¹ or treatment of diseases like cancer.² Most often, SPIOs are synthesized by aqueous coprecipitation method and the surface chemistry of the nanoparticles is governed by the FeOH surface groups. As a result, SPIOs are neutral in the physiological conditions and agglomerate very quickly.³ To prevent this process, various hydrophilic polymers have been attached to sterically stabilize the colloids.^{4,5} Polyethylene glycol (PEG) is one of the most common polymers used due to its high hydrophilicity and biocompatibility.^{4,6} Covalently attached PEG can stabilize the SPIO for long duration at rigorous physiological conditions.⁷ In the fields of biosensing or bioimaging, biomolecules can be covalently grafted onto iron oxide nanoparticles via disulfide bonds. As an example, meso-2,3-dimercaptosuccinic acid (DMSA) provides an easy way to functionalize SPIO with thiol groups^{8,9,10} but their oxidation to polysulfide is hardly prevented.¹⁰ To circumvent the formation of disulfide bonds between the DMSA molecules, coated SPIOs have to be stored in inert conditions and/or at low temperature. Otherwise dithiothreitol (DTT) or tris-(2-carboxyethyl)-phosphine (TCEP) are to be used prior to bio functionalization to regenerate the thiols.¹¹ Finally, a high concentration of DMSA is needed to have at least one free thiol on SPIO but excessive amount of DMSA should be avoided as it is used as a drug to prevent heavy metals toxicity.¹²

The present paper reports a convenient method to stabilize free thiols onto SPIOs by post functionalization using methoxyPEG₂₀₀₀ silane. The stability was proved by grafting a thiol reactive dye: tetramethylrhodamine-5-maleimide (TMRM).¹³

Iron oxide nanoparticles were synthesized by coprecipitation method as described in the literature.¹⁴ Briefly, a 1:2 molar ratio of ferrous and ferric chloride (respectively 1M and 2M) was added dropwise to a 0.75 M NaOH solution at 90 °C. After washing, the final product was kept at pH 3 with a SPIO concentration of 23 mg/mL. The crystallite size of the nanoparticles given by XRD (7.6 ± 0.1) nm is in close agreement with the mean TEM grain size (9 ± 1) nm (Figure 1). The specific surface area of SPIOs is $(110 \pm 1) \text{ m}^2/\text{g}.$

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To prepare DMSA coated SPIOs (DMSA-SPIO), 22 mg of DMSA dissolved in degassed water were mixed to 100 mg of SPIO dispersion at pH 3 under nitrogen flow to prevent the formation of disulfide bonds. The reaction mixture was stirred at room temperature for 2 hours.

After washing, DMSA-SPIO suspension was stored under nitrogen at 4 °C.

To functionalize iron oxide nanoparticles with mPEG₂₀₀₀ (mPEG-SPIO), a silanated methoxy polyethylene glycol (mPEG₂₀₀₀-Si) was synthesized as described in the literature.¹⁵ Briefly, 3-isocyanatopropyltriethoxysilane was attached to mPEG₂₀₀₀, using dibutyltindilaurate, at 60 °C in dried THF under nitrogen flow for 48 hours. mPEG₂₀₀₀-Si was precipitated in n-hexane. Then 400 mg of mPEG₂₀₀₀-Si (molecular weight ~2250 g/mol) was dissolved in 40 mL of ethanol/water mixture (50/50 v/v, pH 4.0) and 100 mg of SPIO dispersion was added to the polymer solution. After 48 hours the reaction mixture was dialyzed and freeze dried before use.

For the bifunctionalization of SPIOs (mPEG-SPIO-DMSA) 7.7 mg of DMSA was added to a suspension containing 500 mg of mPEG-SPIO and was stirred for 2 days. The purification of mPEG-SPIO-DMSA was done by washing with dilute HCl solution (pH 3). The samples were stored under ambient conditions.



Figure 1. TEM images of bare SPIO nanoparticles

Six weeks after the synthesis of bifunctionalized SPIOs tetramethylrhodamine-5-maleimide (TMRM) was added to mPEG-SPIO-DMSA in water at pH 7.5 with five times molar excess of TMRM - previously dissolved in DMSO - and stirred at room temperature for 24 hours. Excess of dye by removed by dilute HCl washing (pH 3) and the suspension was analyzed by UV-visible spectrometry, then kept at -18 °C in the dark.

As a matter of comparison, TMRM was added 24 hours after synthesis of DMSA-SPIO in water at pH 7.5 and stirred at room temperature for 24 hours. After removal of excess TMRM the suspension was analyzed by UV-visible spectrometry.

DMSA-SPIOs formed stable dispersion in 0.15 M NaCl solution from pH 4 to pH 11. Dynamic light scattering experiments showed a mean hydrodynamic diameter of 50 nm in this range of pH. Furthemore the nanoparticles are negatively charged due to the carboxylate groups of DMSA. However, the thiol groups could not be detected by Ellman's reagent (in PBS 20 mM solution) even just after synthesis (Table 1). It has been reported in the literature that DMSA forms tetramers on the surface of nanoparticles.⁹ Indeed these tetramers once adsorbed on the surface can form polysulfides very quickly due to close proximity of intermolecular thiol groups as observed in the template polymerization of sulfides.¹⁰

When DMSA was added to mPEG-SPIO nanoparticles, a stable suspension was observed in 0.15 M of NaCl whatever the pH and the mean hydrodynamic diameter was approximately the same than previously observed. The zeta potential of the nanoparticles was negative for all the pH, for the same reasons than previously explained. But free thiols were sterically protected by the polymer chains and hence Ellman's reagent was able to detect thiol groups even 10 weeks after storage under ambient conditions (see Table 1).

		Molecules/nm ² after grafting after			
Molecule	Nanoparticles	1 day	1 week	6 weeks	10 weeks
DMSA	Bare SPIO	0	0	0	0
DMSA	mPEG-SPIO	nd	0.19	0.14	0.14
TMRM	DMSA-SPIO	0	nd	nd	nd
TMRM	mPEG-SPIO- DMSA	nd	nd	0.28	0.28

Table 1. UV-visible titration of DMSA (using Ellman's reagent) or TMRM on bare and functionalized SPIO

The detection of thiols was then correlated with the estimation of grafted TMRM both by UV visible absorbance at respectively 412 nm and 555 nm (Table 1). After storing mPEG-SPIO-DMSA suspension for 6 weeks under ambient conditions, the average number of thiol groups per nm² was about **0.28**, which means **0.14** DMSA molecules per nm² (see Table 1). **0.28** TMRM molecules were grafted per nm² of SPIO which proved the accessibility of all free thiols and the ability to graft active molecules on there. Scheme 1 shows what could happen when mPEG₂₀₀₀-Si protects thiols of DMSA. These results show a good correlation between still active thiols and grafted TMRM.

In the case of DMSA-SPIO, no grafted TMRM was detected by UV-visible spectrometry confirming the absence of free thiols on the SPIO surface.

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In this study, we have shown that the stability of thiols can be increased significantly when DMSA is protected by PEG chains on the surface of SPIOs. The thiol groups of DMSA were stable even after 10 weeks of storage under ambient conditions. These thiols can be used to attach proteins or thiol reactive dyes without using DTT or stringent storage conditions. Furthermore mPEG-SPIOs-DMSA are stable at physiological conditions. Then this synthesis allows to manipulate well dispersed nanoobjects with functionalizable sites for a long period of time compared to the classical strategies.

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