



HAL
open science

MLVA-16 typing of 295 marine mammal *Brucella* isolates from different animal and geographic origins identifies 7 major groups within *Brucella ceti* and *Brucella pinnipedialis*.

Marianne Maquart, Philippe Le Flèche, Geoffrey Foster, Morten Tryland, Françoise Ramisse, Berit Djønne, Sascha Al Dahouk, Isabelle Jacques, Heinrich Neubauer, Karl Walravens, et al.

► **To cite this version:**

Marianne Maquart, Philippe Le Flèche, Geoffrey Foster, Morten Tryland, Françoise Ramisse, et al.. MLVA-16 typing of 295 marine mammal *Brucella* isolates from different animal and geographic origins identifies 7 major groups within *Brucella ceti* and *Brucella pinnipedialis*.. *BMC Microbiology*, 2009, 9, pp.145. 10.1186/1471-2180-9-145 . hal-00408931

HAL Id: hal-00408931

<https://hal.science/hal-00408931>

Submitted on 30 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Research article

Open Access

MLVA-16 typing of 295 marine mammal *Brucella* isolates from different animal and geographic origins identifies 7 major groups within *Brucella ceti* and *Brucella pinnipedialis*

Marianne Maquart¹, Philippe Le Flèche^{2,3}, Geoffrey Foster⁴, Morten Tryland⁵, Françoise Ramisse³, Berit Djønné⁶, Sascha Al Dahouk^{7,8}, Isabelle Jacques^{1,9}, Heinrich Neubauer^{7,10}, Karl Walravens¹¹, Jacques Godfroid⁵, Axel Cloeckart*¹ and Gilles Vergnaud^{2,12}

Address: ¹INRA, UR1282, Infectiologie Animale et Santé Publique, IASP, Nouzilly, F-37380, France, ²Université Paris-Sud 11, CNRS, UMR8621, Institut de Génétique et Microbiologie, Orsay 91405, France, ³Division of Analytical Microbiology, Centre d'Etudes du Bouchet, Vert le Petit, F-91710, France, ⁴SAC Veterinary Services, Inverness, IV2 4JZ, UK, ⁵Norwegian School of Veterinary Science, Department of Food Safety and Infection Biology, Section of Arctic Veterinary Medicine, Stakkevollveien 23, N-9010 Tromsø, Norway, ⁶Norwegian National Veterinary Institute, Ullevålsveien 68, N-0106 Oslo, Norway, ⁷Bundeswehr Institute of Microbiology, Department of Bacteriology, Neuherbergstr. 11, D-80937 Munich, Germany, ⁸RWTH Aachen University, Department of Internal Medicine III, Pauwelsstraße 30, D-52074 Aachen, Germany, ⁹Institut Universitaire de Technologie, Université François Rabelais, 29 rue du pont-volant, 37082 Tours cedex 2, France, ¹⁰Friedrich Loeffler Institute, Institute of Bacterial Infections and Zoonoses, Naumburger Str. 96a, D-07743 Jena, Germany, ¹¹Veterinary and Agrochemical Research Centre (VAR), Groeselenberg 99, B-1180 Brussels, Belgium and ¹²DGA/MRIS-Mission pour la Recherche et l'Innovation Scientifique, 7-9, rue des Mathurins, Bagneux F-92221, France

Email: Marianne Maquart - marianneaquart@yahoo.fr; Philippe Le Flèche - lefleche@igmors.u-psud.fr; Geoffrey Foster - geoffrey.foster@sac.co.uk; Morten Tryland - Morten.Tryland@veths.no; Françoise Ramisse - francoise.ramisse@dga.defense.gouv.fr; Berit Djønné - berit.djonne@vetinst.no; Sascha Al Dahouk - Sascha.Al-Dahouk@gmx.de; Isabelle Jacques - Isabelle.Jacques@tours.inra.fr; Heinrich Neubauer - heinrich.neubauer@fli.bund.de; Karl Walravens - Karl.Walravens@var.fgov.be; Jacques Godfroid - jacques.godfroid@veths.no; Axel Cloeckart* - Axel.Cloeckart@tours.inra.fr; Gilles Vergnaud - gilles.vergnaud@u-psud.fr

* Corresponding author

Published: 20 July 2009

Received: 9 March 2009

BMC Microbiology 2009, 9:145 doi:10.1186/1471-2180-9-145

Accepted: 20 July 2009

This article is available from: <http://www.biomedcentral.com/1471-2180/9/145>

© 2009 Maquart et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Since 1994, *Brucella* strains have been isolated from a wide range of marine mammals. They are currently recognized as two new *Brucella* species, *B. pinnipedialis* for the pinniped isolates and *B. ceti* for the cetacean isolates in agreement with host preference and specific phenotypic and molecular markers. In order to investigate the genetic relationships within the marine mammal *Brucella* isolates and with reference to terrestrial mammal *Brucella* isolates, we applied in this study the Multiple Loci VNTR (Variable Number of Tandem Repeats) Analysis (MLVA) approach. A previously published assay comprising 16 loci (MLVA-16) that has been shown to be highly relevant and efficient for typing and clustering *Brucella* strains from animal and human origin was used.

Results: 294 marine mammal *Brucella* strains collected in European waters from 173 animals and a human isolate from New Zealand presumably from marine origin were investigated by MLVA-16. Marine mammal *Brucella* isolates were shown to be different from the recognized terrestrial mammal *Brucella* species and biovars and corresponded to 3 major related groups, one specific of the *B. ceti* strains, one of the *B. pinnipedialis* strains and the last composed of the human isolate. In

the *B. ceti* group, 3 subclusters were identified, distinguishing a cluster of dolphin, minke whale and porpoise isolates and two clusters mostly composed of dolphin isolates. These results were in accordance with published analyses using other phenotypic or molecular approaches, or different panels of VNTR loci. The *B. pinnipedialis* group could be similarly subdivided in 3 subclusters, one composed exclusively of isolates from hooded seals (*Cystophora cristata*) and the two others comprising other seal species isolates.

Conclusion: The clustering analysis of a large collection of marine mammal *Brucella* isolates from European waters significantly strengthens the current view of the population structure of these two species, and their relative position with respect to the rest of the *Brucella* genus. MLVA-16 is confirmed as being a rapid, highly discriminatory and reproducible method to classify *Brucella* strains including the marine mammal isolates. The *Brucella2009* MLVA-16 genotyping database available at <http://mlva.u-psud.fr/> is providing a detailed coverage of all 9 currently recognized *Brucella* species.

Background

Brucellae are Gram-negative, facultative, intracellular bacteria that can infect many species of animals and man. Six species were classically recognized within the genus *Brucella*: *B. abortus*, *B. melitensis*, *B. suis*, *B. ovis*, *B. canis*, and *B. neotomae* [1,2]. This classification is mainly based on differences in pathogenicity, host preference, and phenotypic characteristics [1-3]. Three additional species have been recently included in the genus *Brucella*, i.e. *B. ceti* and *B. pinnipedialis* isolated from marine mammals, with cetaceans (dolphin, porpoise, and whale species) and pinnipeds (various seal species) as preferred hosts respectively [4], and *B. microti* isolated from the common vole [5]. From a phenotypic point of view, *B. ceti* and *B. pinnipedialis* can be distinguished by their growth requirement for CO₂ and their oxidative metabolism [6,7].

The phylogenetic significance of this separation is supported by molecular analyses. At the molecular level, evidence for two distinct marine mammal *Brucella* subpopulations subsequently given species rank and designated *B. ceti* and *B. pinnipedialis* has been initially provided by study of DNA polymorphism at the porin-encoding *omp2* locus [8]. This was further confirmed by an infrequent restriction site-PCR (IRS-PCR) method, reflecting the higher number of IS711 elements in the genome of marine mammal isolates compared to terrestrial mammal *Brucella* species [9-11]. IRS-PCR revealed six specific DNA fragments useful for the detection and identification of marine mammal *Brucella* isolates and the presence of a putative genomic island only in seal isolates except for hooded seal isolates [11,12]. Interestingly to date three human cases, one from New Zealand and two from Peru, with *Brucella* infections presumably of marine origin, have been described according to the specific molecular markers cited above, and may point towards a zoonotic potential of these marine mammal *Brucella* species [13,14]. One human case with laboratory acquired infection has also been reported [15].

In the past few years, polymorphic tandem repeat loci have been identified by analysing published genome sequences of *B. melitensis* 16 M, *B. suis* 1330, and *B. abortus* 9-941 [16-18]. Hundreds of *Brucella* strains have been typed to allow the development of an assay, called MLVA-16 assay (Multiple Locus VNTR Analysis) [5,17-23]. The sixteen loci have been grouped in 3 panels, called panel 1 (8 minisatellite loci), panel 2A (3 microsatellite loci) and panel 2B (5 microsatellite loci) [17,20]. Panel 1 has shown to be useful for species identification. Panel 2A and panel 2B increased the discriminatory power. Panel 2B was selected to contain the more highly variable markers, which is why this panel is often given a lower weight in clustering analysis [20,21]. Three of the five octamers in panel 2B have been initially evaluated by Bricker *et al.* [16]. The MLVA-16 assay provides a clustering of strains that is in accordance with the currently recognized *Brucella* species and biovars isolated from terrestrial mammals.

The aim of this study was to evaluate the MLVA-16 assay for the classification of marine mammal *Brucella* isolates, using 294 marine mammal *Brucella* strains obtained from 173 animals representing a wide range of marine mammal species from different European geographic origins (excluding the Mediterranean sea). This study also included the strain, presumably of marine origin, isolated from a patient in New Zealand [14]. A tentative overview of the global *Brucella* population structure was produced by comparison with published typing data.

Results

All strains could be typed at all loci, with few exceptions for panel 2B loci. At the loci *bruce04*, *bruce09* and *bruce16*, multiple bands were observed in the PCR products of 12, 9 and 6 strains, respectively. This may suggest that in some occasions multiple alleles are present in the DNA preparation. Besides, two strains were negative in PCR either for *bruce07* or *bruce30*. In 69 animals, strains were initially isolated from different organs, contributing

121 extra strains. In sixteen among these animals, more than one genotype was observed (in one animal 5 different genotypes were found). In most cases, these genotypes were also observed in at least one other animal. In five cases, at least one of the genotypes was unique in the present collection, suggesting that the presence of multiple genotypes could be the result of a mutation event that occurred in the course of infection. Three of these new genotypes were the result of one repeat unit changes at a single locus. The other two were a 2 repeat units change in bruce04 and a four repeat units change in bruce09. These observations suggest that occasionally the most highly mutable loci may vary in the course of infection. They also do not exclude the possibility that animals carrying multiple variants may have been infected by multiple strains present within the community.

The 294 investigated marine mammal *Brucella* isolates which originated from 173 animals and one patient clustered in 117 different genotypes using the complete MLVA-16 assay. One representative for each genotype and animal was used for analysis, totalling 196 strains (Figures 1, 2, 3). Three main groups were identified, the *B. ceti* group, the *B. pinnipedialis* group and a third group comprising the human isolate from New Zealand. The 117 representative genotypes were compared with the 18 terrestrial mammal *Brucella* reference strains and published data (Figure 4). The 3 clusters were clearly separated from all the terrestrial mammal isolates.

The cetacean group composed of 102 strains presenting 74 genotypes (1–74) (Figure 1) could be separated into three major subclusters. The first two subclusters A1 and A2, from genotype 1 to 18 were only composed of dolphin isolates and one minke whale isolate (9 isolates from striped dolphins (*Stenella coeruleoalba*), 11 from Atlantic white sided dolphins (*Lagenorhynchus acutus*), 2 from bottlenose dolphins (*Tursiops truncatus*) and 1 from a common dolphin (*Delphinus delphis*)). These two subclusters correspond to sequence type ST26 [24], MLVA panel 1 genotype 24 (subcluster A1) and 77 (subcluster A2, Figure 1 and Figure 3), and together correspond to cluster A in [25] (Figure 3). The third subcluster, from genotype 19 to 74 corresponds to MLST sequence type 23, MLVA-16 panel 1 genotypes 23, 69 and 70, and is cluster B in [25] (Figure 1 and Figure 3). This subcluster was composed of 78 strains. Sixty-four were obtained from porpoises, 12 from 4 species of dolphins (9 from Atlantic white sided dolphin (*Lagenorhynchus acutus*), one from a white-beaked dolphin (*Lagenorhynchus albirostris*), one from a bottlenose dolphin (*Tursiops truncatus*), one from a common dolphin (*Delphinus delphis*), and one from a minke whale (*Balaenoptera acutorostrata*) isolated in Norway in 1995 [10] (Figure 1). An exception was the bmar111 (strain number M490/95/1), with the genotype 20, isolated in

Scotland from a harbour (or common) seal (*Phoca vitulina*) and which belongs to the *B. ceti* group (Figure 1). This is, however, in agreement with previous observations, either phenotypic [26] or molecular, including MLVA typing [25]. This particular strain carries the two specific IRS-PCR fragments (II and III) of the *B. ceti* strains [11], and the PCR-RFLP pattern of the *omp2* genes is similar to that of *Brucella* strains isolated from porpoises [8].

The 93 representative *B. pinnipedialis* strains presented 42 different genotypes (75–116) (Figure 2) corresponding to cluster C in [25]. This group of isolates could similarly be further divided in three major subclusters. The first subcluster (genotype 75 to 101) was composed of several seal isolates (harbour seal and grey seal (*Halichoerus grypus*)) and the isolate from a European sea otter (*Lutra lutra*). It corresponds to MLST sequence type 25, MLVA panel 1 genotypes 25, 72, 73, and cluster C2 in [25]. The second subcluster (MLVA genotypes 102 to 107) corresponds to MLST sequence type 24, MLVA panel 1 genotypes 71 and 79 and is cluster C1 in [25]. Interestingly, the hooded seal isolates (15 strains) were exclusively clustered in 9 closely related genotypes, forming the third subcluster of the pinniped isolates (genotype 108 to 116) called C3 in [25]. Most of the hooded seal isolates analysed in this study were isolated in Norway in 2002 [27] and there were also 4 hooded seal isolates from Scotland that clustered with the Norwegian isolates. One of the 93 strains of the *B. pinnipedialis* group was obtained from a cetacean. This strain (M192/00/1), identified as bmar160 with the genotype 107 in Figure 2, was isolated from a minke whale in Scotland in 2000. This strain was also demonstrated as a *B. pinnipedialis* strain by other molecular markers, as described by Maquart *et al.* [12] and Groussaud *et al.* [25].

The human isolate 02/611 (bmar265) from New Zealand (genotype 117 (Figure 2)), clearly separates from the clusters described above but fits in an intermediate position within the marine mammal groups of isolates in agreement with previous MLST analyses [28]. Figure 3 suggests a position closer to the *B. ceti* group in agreement with the phenotypic behaviour [14], but the typing of more strains from Pacific waters [29–31] will be needed in order to achieve a more conclusive cluster analysis.

Owing to the inclusion of 40 representative strains in duplicate, the results described above could be compared to those recently described by Groussaud *et al.* who studied 74 marine mammal isolates by multilocus sequence typing, multilocus sequence analysis (MLST, MLSA) and MLVA. Duplicate typing was useful since Groussaud *et al.* used a partially different set of 21 VNTRs [25] (9 loci are common, including three loci from panel 1 (Bruce 08, 45, 55), one from panel 2A (Bruce18) and the whole panel 2B).

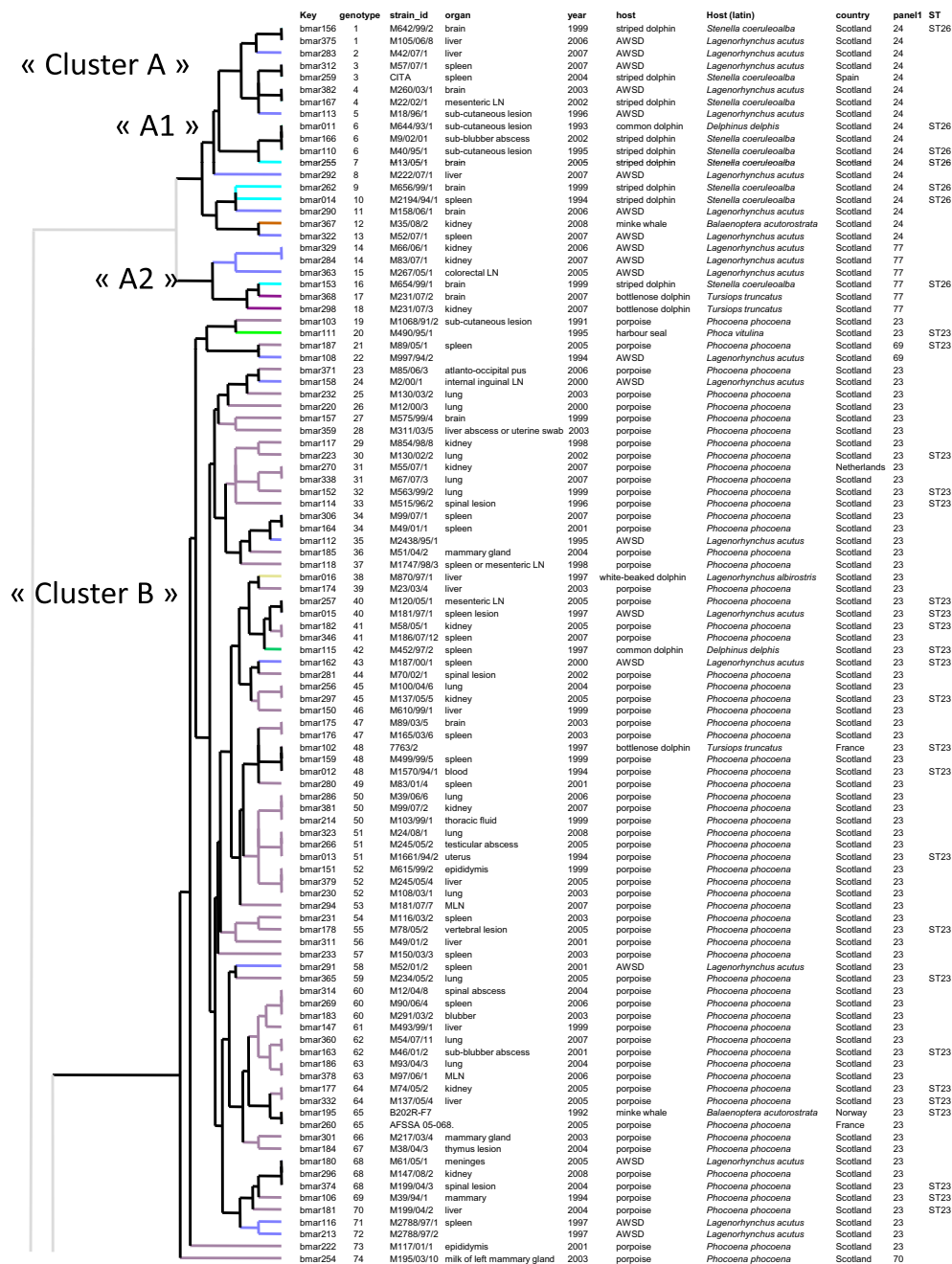


Figure 1
MLVA-16 clustering analysis of 102 *B. ceti* strains defines three groups of strains. All *B. ceti* isolates cluster into a first part (genotypes 1 to 74) of the dendrogram constructed from MLVA-16 testing of 294 *Brucella* strains obtained from 173 marine mammals (pinnipeds, otter and cetaceans) and one human patient from New Zealand. One strain per genotype and per animal is included (consequently some animals are represented by more than one strain), 196 entries are listed corresponding to 117 genotypes. In the columns, the following data are presented: DNA batch (key), genotype, strain identification, organ, year of isolation, host (AWSD: Atlantic White Sided Dolphin), host (Latin name), geographic origin, MLVA panel I genotype, sequence type when described by Groussaud *et al.* [25]. The colour code reflects the host species (see Figure 3 for detailed correspondence). No colour was used when identical genotypes were observed in different host species. The letter nomenclature proposed by Groussaud *et al.* is used (*B. ceti*, cluster A (ST26) further subdivided into A1 and A2 and cluster B (ST23)).

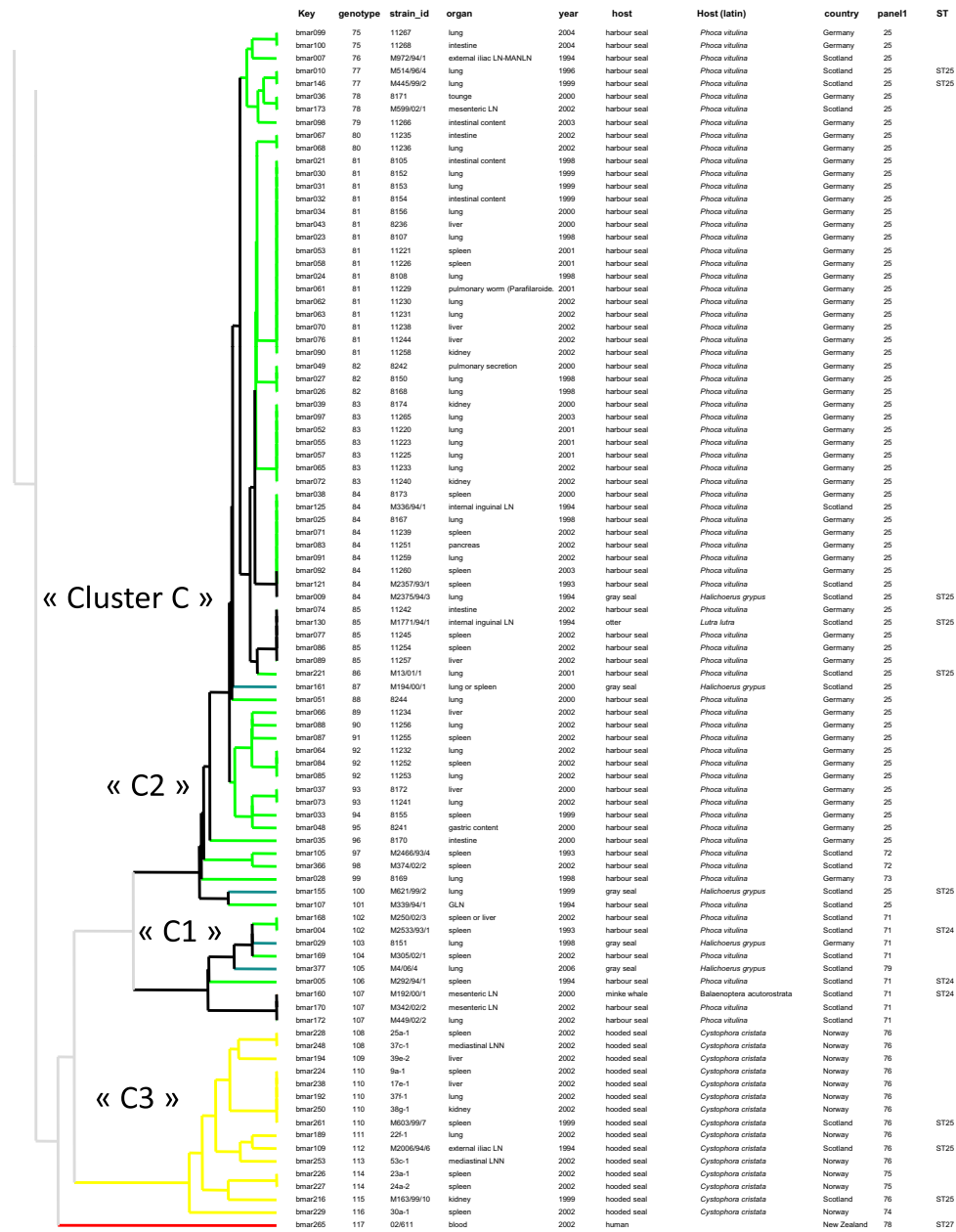


Figure 2
MLVA-16 clustering analysis of 93 *B. pinnipedialis* strains defines 3 groups of strains. All *B. pinnipedialis* isolates cluster together in the second part (genotypes 75 to 117) of the dendrogram constructed from MLVA-16 testing of 294 *Brucella* isolates obtained from 173 marine mammals (pinnipeds, otter and cetaceans) and one human patient from New Zealand. In the columns, the following data are presented: DNA batch (key), genotype, strain identification, organ, year of isolation, host (AWSD: Atlantic White Sided Dolphin), host (Latin name), geographic origin, MLVA panel I genotype, sequence type when described by Groussaud *et al.* [25]. The colour code reflects the host species (see Figure 3 for detailed correspondence). No colour was used when identical genotypes were observed in different host species. The red branch (genotype 117) corresponds to the human isolate (ST27). The letter nomenclature proposed by Groussaud *et al.* is used (*B. pinnipedialis*, cluster C, including C1 (ST24), C2 (ST25) and C3 (ST25)).

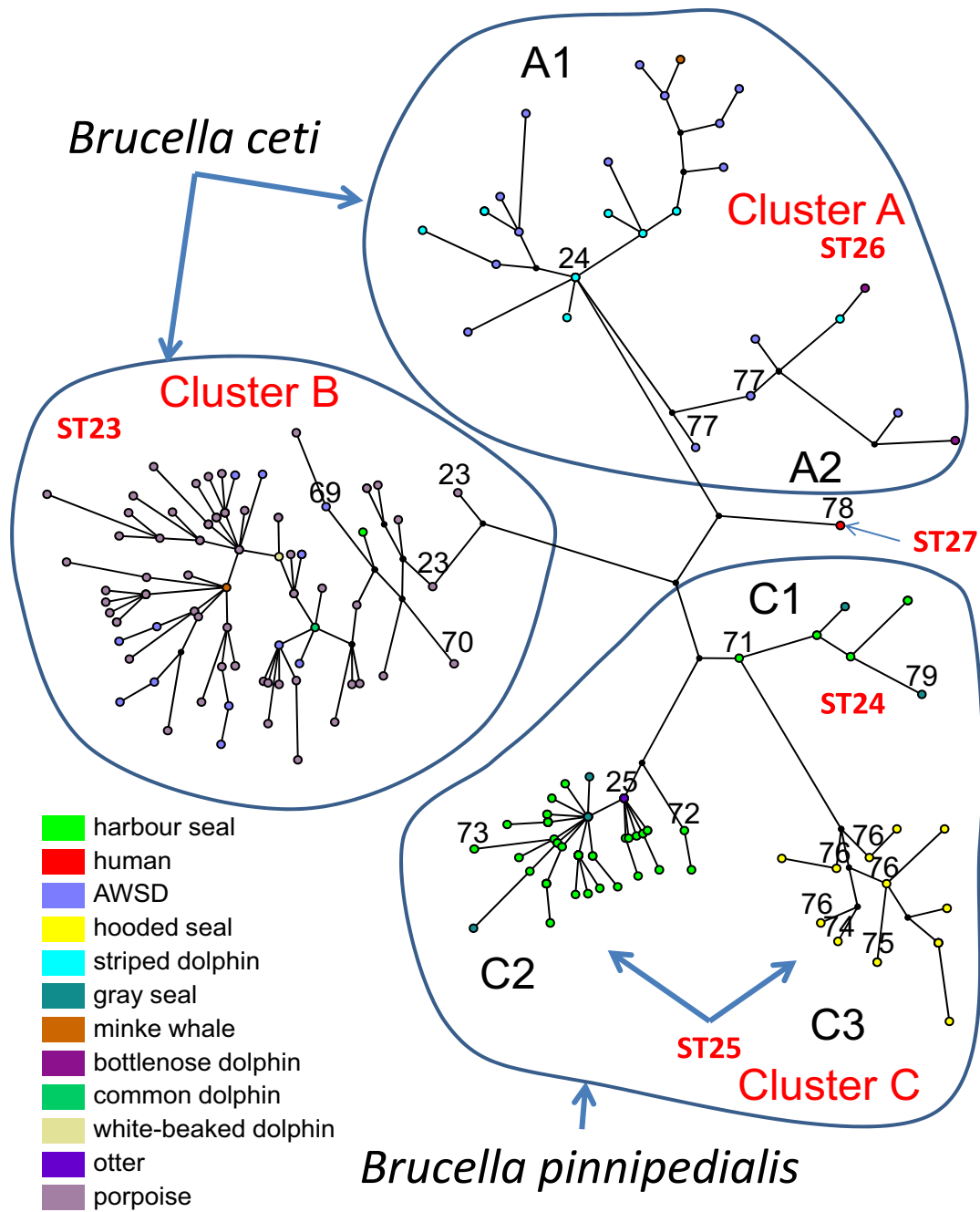


Figure 3

Maximum parsimony analysis on 117 marine mammal *Brucella* genotypes. Each coloured circle corresponds to one MLVA-16 genotype from a marine mammal species. Numbers in black (23, 24, 25, 69 to 79) indicate the MLVA the panel I genotype for the colour circle below. The panel I genotype along daughter branches is indicated only when it is different from the proposed parent node (i.e. in cluster A, all strains are panel I genotype 24 in subcluster A1 or 77 in subcluster A2). The tentative MLST sequence type (ST23 to ST27) as predicted from strains shared between this study and [25] is indicated, together with species assignment. The host species colour code indicated is the same as in Figures 1 and 2 (AWSD: Atlantic White Sided Dolphin).

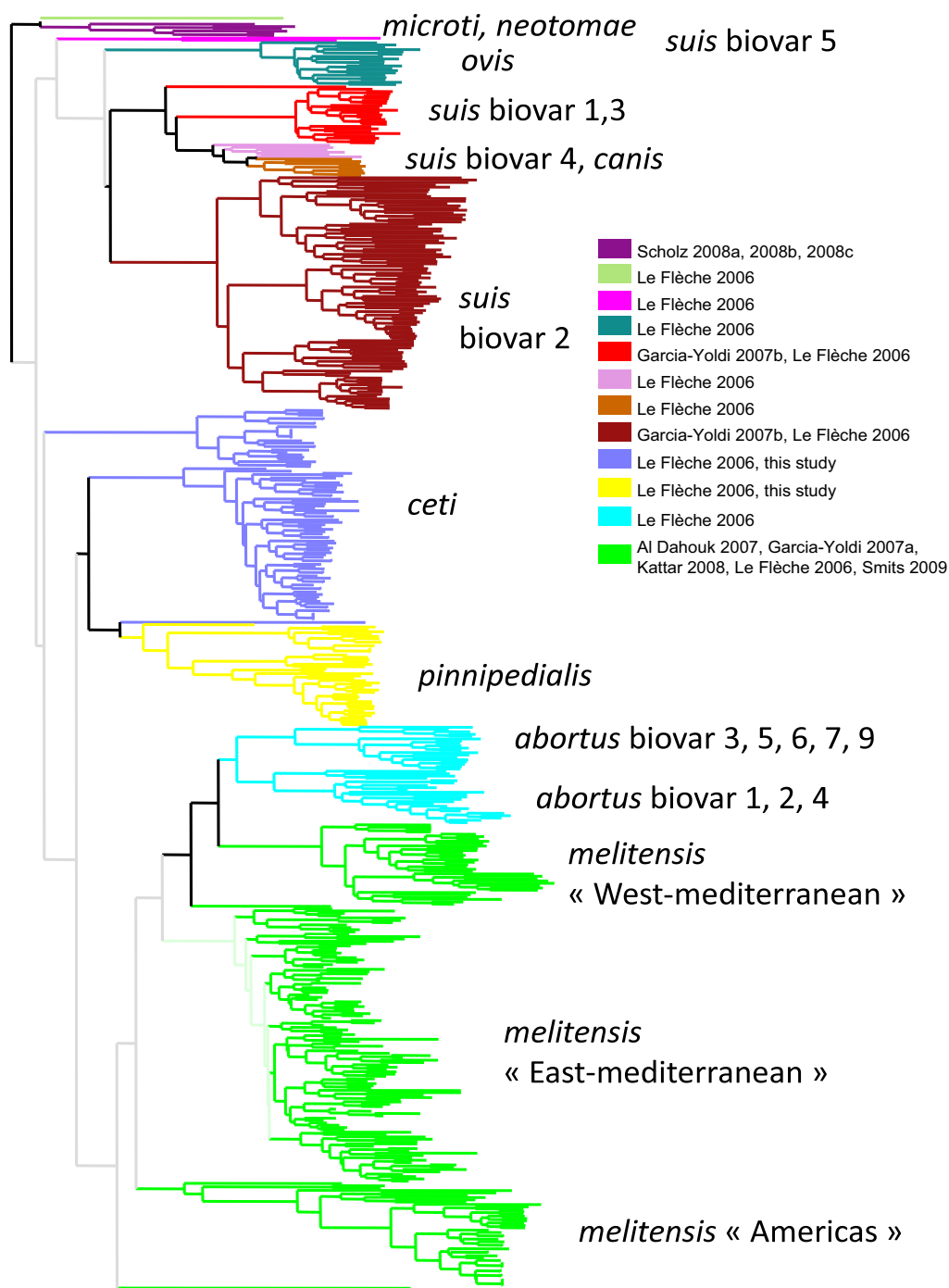


Figure 4
Current view of the global population structure of the *Brucella* genus. Clustering was done using the Neighbor Joining (NJ) algorithm. The microti/neotomae cluster was used to root the tree. The dendrogram is based upon more than 500 genotypes, observed by typing more than 750 strains [see Additional file 1]. The terrestrial mammal strains data were compiled from [5,17,19-23,37]. The colour code reflects the *Brucella* species (or some highly specific biovars). The publications from which the data were derived are indicated. The long blue branch close to the *B. pinnipedialis* cluster represents the human isolate from New Zealand (MLST ST27).

Discussion

Since 1994, marine mammal *Brucella* strains have been isolated and characterized, both phenotypically and by means of different molecular typing methods. This led to the division of the marine mammal *Brucella* strains in 2 species i.e. *B. ceti* on one side and *B. pinnipedialis* on the other side defined by oxidative metabolism patterns and CO₂ requirement for growth, and a number of subclusters defined by complementary molecular analysis methods.

This MLVA-16 study is, to date, the most important one in terms of number of strains analysed and number of animal species from which these strains have been isolated. These strains were isolated from animals stranded, caught or killed for scientific purposes in the waters surrounding Europe, from the Barents Sea, above the Arctic Circle to the Atlantic coast of Spain. For the 295 strains analysed, using the MLVA-16 assay, 117 genotypes were resolved and seven clusters were identified, (i) two clusters almost exclusively composed of dolphin isolates, (ii) the predominantly porpoise cluster of strains (which also includes several strains isolated from dolphins), (iii) two main seal species clusters, (iv) the hooded seal cluster, and (v) the human isolate. The last cluster might correspond to Pacific Ocean isolates [29-31], which are under-represented in the present collection. The hooded seal cluster of strains was composed of strains from Scotland and Norway. The low level of genetic diversity between the hooded seal isolates from Scotland and from Norway could indicate that all the investigated hooded seals originated from the same population of animals. The population that was sampled between Svalbard and Greenland have their breeding area in the pack ice north of Jan Mayen (West Ice), but except for the few weeks on ice during birth, mating and moulting, the hooded seal is a typical pelagic and a migratory species with a huge geographical range [27]. It is thus likely that the hooded seals stranded in Scotland were originally from this area rather than from the more western bound populations of hooded seals that breed in the Davis Strait and at the east coast of Canada.

Groussaud *et al.* [25] analysed the diversity of marine mammal isolates by MLVA using another selection of VNTRs, including all 8 loci defining the HOOF-prints MLVA assay described by Bricker *et al.* [16] and 13 additional loci characterised by Le Flèche *et al.* [17] and Whatmore *et al.* [18]. This panel of 21 VNTR loci corresponded to a 21-locus MLVA scheme sharing 9 loci with MLVA-16 and also provides a high degree of diversity. In this previous study, multilocus sequence types (STs) were determined, allowing the clustering of marine mammal isolates in five groups labelled ST23 to ST27. The closely related ST24 and ST25 were composed of the pinniped isolates, forming the cluster C. The hooded seal isolates

define subcluster C3. ST26 was exclusively composed of dolphin isolates and formed the cluster A. The other cetacean isolates all clustered in the cluster B (ST23) and consisted of strains isolated from porpoises and dolphins. ST27 was represented by only one isolate from an aborted bottlenose dolphin foetus originating from the Western coast of the United States (strain F5/99) [28]. Our results are thus in excellent accordance with those published by Groussaud *et al.* [25] showing that the previously identified population structure of marine mammal *Brucella* strains is not significantly modified by the inclusion of a large number of strains from European waters. MLVA-16 results are also in accordance with the recently reported genomic structures of 24 marine mammal *Brucella* isolates for which three subgroups were identified [32]. In that study, one separate group was identified for the *B. pinnipedialis* strains, another subgroup included dolphin isolates and a third subgroup comprised dolphin and porpoise isolates. The only hooded seal isolate analysed in that study clustered in the *B. pinnipedialis* group but revealed a separate pattern with a 62 kb missing fragment, specific for this group and relevant for a distinct genetic background [32].

MLVA-16 classification in the present report revealed some exceptions like the M490/95/1 strain, isolated from a common seal, which was clustered in the *B. ceti* group of strains. This exception suggests that transmission from cetaceans to pinnipeds may occur. Although the currently recognized terrestrial mammal *Brucella* species also have a preferred host, they can be isolated from different hosts in regions where brucellosis is endemic, e.g. *B. melitensis* which has been isolated from cattle in the southern part of France [33].

The human isolate from New Zealand formed a separate seventh MLVA-16 cluster. Whatmore *et al.* [28] have shown that the F5/99 strain, isolated from an aborted bottlenose dolphin fetus from the Western coast of the United States (together with three human isolates, one from New Zealand and two from Peru) shared the same MLST genotype (ST27). Interestingly, at the *omp2* level both strains were identical and also identical to isolates from the North Pacific minke whales. In all these strains the porin *omp2* genes were different from those from marine mammal strains isolated on European coasts [30]. Briefly, the *omp2* genes of these isolates from the Pacific share common features with both marine mammal (from Europe) and terrestrial mammal strains [29]. Another interesting observation is that all the Pacific isolates investigated so far (including the three reported human cases) carry fragment I identified by IRS-PCR which is part of a putative genomic island specific for *B. pinnipedialis* [12]. Since these cetacean isolates are quite distinct from European marine mammal isolates there might be a third

marine mammal *Brucella* species or subspecies found in Pacific waters. Owing to the simplicity of MLVA-16 typing, and in particular of panel 1 which can be typed on regular agarose gels and already provides a high informativity in classifying marine mammal strains (Figure 3), more typing information on Pacific Ocean strains (including the strains described in [29-31]) will likely be made available in a near future. The *Brucella*2009 genotyping database available at <http://mlva.u-psud.fr/> and based upon the data provided in Additional file 1 can be used for this purpose. Figure 4 shows the global population structure of the nine species currently constituting the *Brucella* genus, as can be revealed by MLVA-16 typing using this dataset (the extended data set provided here may provide new opportunities to evaluate additional methods for *Brucella* MLVA data clustering recently proposed [34]).

Conclusion

MLVA-16 proved to be useful for molecular classification of a high number of marine mammal *Brucella* strains and allows the typing of large populations, while providing a clustering in agreement with all previously reported methods, together with a much higher discriminatory power. From the clustering achieved, a few representative strains can be selected for whole genome sequencing.

Methods

Brucella strains

MLVA analysis was performed on 294 isolates from 173 marine mammals and one human patient. The strains essentially originate from the Northern Atlantic, from three main sources, Scotland (216 isolates from 116 animals), Germany (58 isolates from 42 animals) [35] and Norway (18 isolates from 13 animals) [27]. Six additional strains from various geographic origins were analysed. Two strains were obtained from France (one strain from a bottlenose dolphin (*Tursiops truncatus*) and one from a harbour porpoise (*Phocoena phocoena*)), one from Spain (from a striped dolphin (*Stenella coeruleoalba*)) [36] and two from The Netherlands (two strains from one harbour porpoise (*Phocoena phocoena*)). The sixth strain was a human isolate from New-Zealand (strain 02/611 genotype 117) [14]. Strains (one strain per genotype and animal) are listed in Figures 1 and 2 and in Additional file 1.

MLVA analysis

The method for selecting appropriate VNTRs has been described previously by Le Flèche *et al.* [17], adapted by Al Dahouk *et al.* (the initial MLVA-15 assay was completed by bruce19) [20]. The results were compared with the MLVA-16 results obtained for the 18 terrestrial mammal *Brucella* reference strains published previously by Le Flèche *et al.* [17] and additional published data [5,19-23,37]. The sixteen loci have been classified in 3 panels, called panel 1 (8 minisatellite loci), panel 2A (3 microsat-

ellite loci) and panel 2B (5 microsatellite loci) [20]. Panel 1 was composed of bruce06, bruce08, bruce11, bruce12, bruce42, bruce43, bruce45, bruce55, useful for species identification. Panel 2, showing a higher discriminatory power, was split into two groups, panel 2A and 2B, composed of three (bruce18, bruce19, bruce21) and five (bruce04, bruce07, bruce09, bruce16, bruce30) markers, respectively. Panel 2B contains the more variable loci, and this panel can be given a lower weight in clustering analysis, as described by Al Dahouk *et al.* [20] and Kattar *et al.* [21].

PCR amplification

Brucella DNA was prepared as previously described by Cloeckaert *et al.* [38]. PCR amplification was performed in a total volume of 15 µl containing 1 ng of DNA, 1× PCR reaction buffer, 1 U of Taq DNA polymerase (QBiogen, Illkirch, France), 200 µM of each deoxynucleotide triphosphate, and 0.3 µM of each flanking primer as described by Le Flèche *et al.* [17].

Amplifications were performed in a MJ Research PTC200 thermocycler. An initial denaturation step at 96°C for 5 minutes was followed by 30 cycles of denaturation at 96°C for 30 s, primer annealing at 60°C for 30 s, and elongation at 70°C for 1 min. The final extension step was performed at 70°C for 5 min.

Two to five microliters of the amplification product were loaded on a 3% standard agarose gel for analyzing tandem repeats with a unit length shorter than 10 bp (panel 2) and on a 2% standard agarose gel for all others (panel 1), and run under a voltage of 8 V/cm until the bromophenol blue dye had reached the 20 cm position. Gels were stained with ethidium bromide, visualized under UV light, and photographed (Vilber Lourmat, Marnes-la-Vallée, France). A 100-bp and a 20-bp ladder (EZ load 100 bp or 20 bp PCR Molecular Ruler, Biorad, Marnes-la-Coquette, France) were used as molecular size markers depending on the tandem repeat unit length. Gel images were managed using the BioNumerics software package (version 6.0, Applied-Maths, Belgium).

Data analysis

Band size estimates were converted to a number of units within a character dataset using the BioNumerics software and the previously published allele calling convention [17]. Clustering analyses used the categorical coefficient and the UPGMA (unweighted pair group method using arithmetic averages) or Neighbor Joining algorithm. The use of categorical parameter implies that the character states are considered unordered. The same weight is given to a large or a small number of differences in the number of repeats at each locus. Maximum parsimony was done using BioNumerics, running 200 bootstrap simulations

treating the data as categorical and giving the same weight to all loci.

Authors' contributions

JG and GV coordinated contributions by the different participants. IJ, MT, GF, BD, SAD, HN, FR, KW and JG isolated and/or maintained strains and/or produced DNA. PLF did the MLVA genotyping work. GV and PLF were in charge of the BioNumerics database, error checking, clustering analyses. MM, AC and GV wrote the report. IJ helped to draft the manuscript. All authors read, commented and approved the final manuscript.

Additional material

Additional file 1

MLVA-16 data. The repeat copy numbers at each locus are indicated for each strain.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2180-9-145-S1.xls>]

Acknowledgements

Work on the typing of dangerous pathogens is supported by the French "Délégation Générale pour l'Armement" (DGA) and by the European Defense Agency. GV, PLF, FR are members of the European Biodefense Laboratory Network (EBLN). We thank Vincent Ramière and Claudette Simoes from the Centre d'Etudes du Bouchet DNA bank for the provision of DNAs. We thank Bruno Garin-Bastuji, Clara M. Marin and Wendy McDonald for the gift of *Brucella* strains or DNA of marine mammal origin from France, Spain and New Zealand, respectively.

References

- Corbel MJ, Brinley Morgan WJ: **Genus *Brucella* Meyer and Shaw 1920, 173AL.** In *Bergey's Manual of Systematic Bacteriology Volume 1*. Edited by: Krieg NR, Holt JG. Baltimore: Williams and Wilkins; 1984:377-390.
- Moreno E, Cloeckaert A, Moriyón I: ***Brucella* evolution and taxonomy.** *Vet Microbiol* 2002, **90**:209-227.
- Alton GG, Jones LM, Angus RD, Verger JM: *Techniques for the brucellosis laboratory* Paris, France: INRA; 1988.
- Foster G, Osterman BS, Godfroid J, Jacques I, Cloeckaert A: ***Brucella ceti* sp. nov. and *Brucella pinnipedialis* sp. nov. for *Brucella* strains with cetaceans and seals as their preferred hosts.** *Int J Syst Evol Microbiol* 2007, **57**:2688-2693.
- Scholz HC, Hubálek Z, Sedláček I, Vergnaud G, Tomaso H, Al Dahouk S, Melzer F, Kämpfer P, Neubauer H, Cloeckaert A, et al.: ***Brucella microti* sp. nov., isolated from the common vole *Microtus arvalis*.** *Int J Syst Evol Microbiol* 2008, **58**:375-382.
- Jahans KL, Foster G, Broughton ES: **The characterisation of *Brucella* strains isolated from marine mammals.** *Vet Microbiol* 1997, **57**:373-382.
- Jacques I, Grayon M, Verger JM: **Oxidative metabolic profiles of *Brucella* strains isolated from marine mammals: contribution to their species classification.** *FEMS Microbiol Lett* 2007, **270**:245-249.
- Cloeckaert A, Verger JM, Grayon M, Paquet JY, Garin-Bastuji B, Foster G, Godfroid J: **Classification of *Brucella* spp. isolated from marine mammals by DNA polymorphism at the omp2 locus.** *Microbes Infect* 2001, **3**:729-738.
- Bricker BJ, Ewalt DR, MacMillan AP, Foster G, Brew S: **Molecular characterization of *Brucella* strains isolated from marine mammals.** *J Clin Microbiol* 2000, **38**:1258-1262.
- Clavareau C, Wellemans V, Walravens K, Tryland M, Verger JM, Grayon M, Cloeckaert A, Letesson JJ, Godfroid J: **Phenotypic and molecular characterization of a *Brucella* strain isolated from a minke whale (*Balaenoptera acutorostrata*).** *Microbiology* 1998, **144**(Pt 12):3267-3273.
- Cloeckaert A, Grayon M, Grepinet O, Boumedine KS: **Classification of *Brucella* strains isolated from marine mammals by infrequent restriction site-PCR and development of specific PCR identification tests.** *Microbes Infect* 2003, **5**:593-602.
- Maquart M, Fardini Y, Zygmunt MS, Cloeckaert A: **Identification of novel DNA fragments and partial sequence of a genomic island specific of *Brucella pinnipedialis*.** *Vet Microbiol* 2008, **132**:181-189.
- Sohn AH, Probert WS, Glaser CA, Gupta N, Bollen AW, Wong JD, Grace EM, McDonald WC: **Human neurobrucellosis with intracerebral granuloma caused by a marine mammal *Brucella* spp.** *Emerg Infect Dis* 2003, **9**:485-488.
- McDonald WL, Jamaludin R, Mackereth G, Hansen M, Humphrey S, Short P, Taylor T, Swingle J, Dawson CE, Whatmore AM, et al.: **Characterization of a *Brucella* sp. strain as a marine-mammal type despite isolation from a patient with spinal osteomyelitis in New Zealand.** *J Clin Microbiol* 2006, **44**:4363-4370.
- Brew SD, Perrett LL, Stack JA, MacMillan AP, Staunton NJ: **Human exposure to *Brucella* recovered from a sea mammal.** *Vet Rec* 1999, **144**:483.
- Bricker BJ, Ewalt DR, Halling SM: ***Brucella* 'Hoof-Prints': strain typing by multi-locus analysis of variable number tandem repeats (VNTRs).** *BMC Microbiol* 2003, **3**:15.
- Le Flèche P, Jacques I, Grayon M, Al Dahouk S, Bouchon P, Denoëud F, Nöckler K, Neubauer H, Guilloteau LA, Vergnaud G: **Evaluation and selection of tandem repeat loci for a *Brucella* MLVA typing assay.** *BMC Microbiol* 2006, **6**:9.
- Whatmore AM, Shankster SJ, Perrett LL, Murphy TJ, Brew SD, Thirlwall RE, Cutler SJ, MacMillan AP: **Identification and characterization of variable-number tandem-repeat markers for typing of *Brucella* spp.** *J Clin Microbiol* 2006, **44**:1982-1993.
- García-Yoldi D, Le Flèche P, De Miguel MJ, Muñoz PM, Blasco JM, Cvetnic Z, Marin CM, Vergnaud G, López-Goñi I: **Comparison of multiple-locus variable-number tandem-repeat analysis with other PCR-based methods for typing *Brucella suis* isolates.** *J Clin Microbiol* 2007, **45**:4070-4072.
- Al Dahouk S, Le Flèche P, Nöckler K, Jacques I, Grayon M, Scholz HC, Tomaso H, Vergnaud G, Neubauer H: **Evaluation of *Brucella* MLVA typing for human brucellosis.** *J Microbiol Methods* 2007, **69**:137-145.
- Kattar MM, Jaafar RF, Araj GF, Le Flèche P, Matar GM, Abi Rached R, Khalife S, Vergnaud G: **Evaluation of a multilocus variable-number tandem-repeat analysis scheme for typing human *Brucella* isolates in a region of brucellosis endemicity.** *J Clin Microbiol* 2008, **46**:3935-3940.
- Scholz HC, Hofer E, Vergnaud G, Le Flèche P, Whatmore AM, Al Dahouk S, Pfeffer M, Krüger M, Cloeckaert A, Tomaso H: **Isolation of *Brucella microti* from Mandibular Lymph Nodes of Red Foxes, *Vulpes vulpes*, in Lower Austria.** *Vector Borne Zoonotic Dis* 2009, **9**:153-156.
- Scholz HC, Hubálek Z, Nesvadbová J, Tomaso H, Vergnaud G, Le Flèche P, Whatmore AM, Al Dahouk S, Krüger M, Lodri C, Pfeffer M: **Isolation of *Brucella microti* from soil.** *Emerg Infect Dis* 2008, **14**:1316-1317.
- Whatmore AM, Perrett LL, MacMillan AP: **Characterisation of the genetic diversity of *Brucella* by multilocus sequencing.** *BMC Microbiol* 2007, **7**:34.
- Groussaud P, Shankster SJ, Koylass MS, Whatmore AM: **Molecular typing divides marine mammal strains of *Brucella* into at least three groups with distinct host preferences.** *J Med Microbiol* 2007, **56**:1512-1518.
- Foster G, MacMillan AP, Godfroid J, Howie F, Ross HM, Cloeckaert A, Reid RJ, Brew S, Patterson IA: **A review of *Brucella* sp. infection of sea mammals with particular emphasis on isolates from Scotland.** *Vet Microbiol* 2002, **90**:563-580.
- Tryland M, Sørensen KK, Godfroid J: **Prevalence of *Brucella pinnipediae* in healthy hooded seals (*Cystophora cristata*) from the North Atlantic Ocean and ringed seals (*Phoca hispida*) from Svalbard.** *Vet Microbiol* 2005, **105**:103-111.
- Whatmore AM, Dawson CE, Groussaud P, Koylass MS, King AC, Shankster SJ, Sohn AH, Probert WS, McDonald WL: **Marine mam-**

- mal *Brucella* genotype associated with zoonotic infection. *Emerg Infect Dis* 2008, **14**:517-518.
29. Ohishi K, Takishita K, Kawato M, Zenitani R, Bando T, Fujise Y, Goto Y, Yamamoto S, Maruyama T: **Chimeric structure of omp2 of *Brucella* from Pacific common minke whales (*Balaenoptera acutorostrata*).** *Microbiol Immunol* 2005, **49**:789-793.
 30. Ohishi K, Takishita K, Kawato M, Zenitani R, Bando T, Fujise Y, Goto Y, Yamamoto S, Maruyama T: **Molecular evidence of new variant *Brucella* in North Pacific common minke whales.** *Microbes Infect* 2004, **6**:1199-1204.
 31. Hernández-Mora G, González-Barrientos R, Morales JA, Chaves-Olarte E, Guzmán-Verri C, Barquero-Calvo E, De-Miguel MJ, Marín CM, Blasco JM, Moreno E: **Neurobrucellosis in stranded dolphins, Costa Rica.** *Emerg Infect Dis* 2008, **14**:1430-1433.
 32. Bourg G, O'Callaghan D, Boschirolu ML: **The genomic structure of *Brucella* strains isolated from marine mammals gives clues to evolutionary history within the genus.** *Vet Microbiol* 2007, **125**:375-380.
 33. Verger JM, Garin-Bastuji B, Grayon M, Mahe AM: **[Bovine brucellosis caused by *Brucella melitensis* in France].** *Ann Rech Vet* 1989, **20**:93-102.
 34. Almendra C, Silva TL, Beja-Pereira A, Ferreira AC, Ferrão-Beck L, de Sá MI, Bricker BJ, Luikart G: **"HOOF-Print" genotyping and haplotype inference discriminates among *Brucella* spp. isolates from a small spatial scale.** *Infect Genet Evol* 2009, **9**:104-107.
 35. Prenger-Berninghoff E, Siebert U, Stede M, König A, Weiss R, Baljer G: **Incidence of *Brucella* species in marine mammals of the German North Sea.** *Dis Aquat Organ* 2008, **81**:65-71.
 36. Muñoz PM, García-Castrillo C, López-García P, González-Cueli JC, De Miguel MJ, Marín CM, Barberán M, Blasco JM: **Isolation of *Brucella* species from a live-stranded striped dolphin (*Stenella coeruleoalba*) in Spain.** *Vet Rec* 2006, **158**:450-451.
 37. Smits HL, Espinosa B, Castillo R, Hall E, Guillen A, Zevaleta M, Gilman RH, Melendez P, Guerra C, Draeger A, et al.: **MLVA genotyping of human *Brucella* isolates from Peru.** *Trans R Soc Trop Med Hyg* 2009, **103**:399-402.
 38. Cloeckert A, Verger JM, Grayon M, Grepinet O: **Restriction site polymorphism of the genes encoding the major 25 kDa and 36 kDa outer-membrane proteins of *Brucella*.** *Microbiology* 1995, **141**(Pt 9):2111-2121.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

