



HAL
open science

Near-infrared reflectance spectroscopy (NIRS): a practical tool for the assessment of soil carbon and nitrogen budget

Lauric Cécillon, Jean-Jacques Brun

► To cite this version:

Lauric Cécillon, Jean-Jacques Brun. Near-infrared reflectance spectroscopy (NIRS): a practical tool for the assessment of soil carbon and nitrogen budget. R. Jandl & M. Olsson. COST Action 639: Greenhouse-gas Budget of Soils Under Changing Climate and Land Use (BurnOut)., Federal Research and Training Centre for Forests, Natural Hazards and Landscape (BFW), Vienna, pp.103-110, 2007, Diverse Veröffentlichungen Nr. 40. hal-00396317

HAL Id: hal-00396317

<https://hal.science/hal-00396317>

Submitted on 17 Jun 2009

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Near-infrared reflectance spectroscopy (NIRS): a practical tool for the assessment of soil carbon and nitrogen budget

Lauric Cécillon^{1,2,*} & Jean-Jacques Brun¹

Addresses:

¹ Cemagref Grenoble, Mountain Ecosystems Research Unit, 2 rue de la papeterie BP 76, 38402 Saint Martin d'Hères Cedex, France

² French Agency for Environment and Energy Management (ADEME) 2 square La Fayette BP 90406, 49004 Angers Cedex 01, France

* corresponding author: L. Cécillon, Phone: + 33 (0)130 799 564, E-Mail: cecillon@cetiom.fr, Webpage: <http://lauric.cecillon.free.fr/>

Abstract

The assessment of greenhouse gas emissions from soils requires an accurate knowledge on the fate of carbon (C) and nitrogen (N) in soils. This knowledge shouldn't be limited to C and N stocks in soils but should incorporate the quantification of functional soil organic matter (SOM) fractions with different turnover rates. Conventional analysis can't be used to assess such functional properties of SOM over large geographical areas. Near-infrared reflectance spectroscopy (NIRS) is a rapid and cheap method that could achieve this goal. This technology has proven its efficiency to discriminate among functional SOM fractions and to predict C, N on large sets of soil samples at a large regional scale. NIRS can be used for the detailed analysis of decomposition and aggregation of SOM, two key ecosystem processes that control C and N budget in soils. This review points up the improvement of NIR prediction models by the technique of local calibration and the potential of NIR analysis to assess the ontogeny and heterogeneity of detritus in soils. Mapping soil C, N and SOM fractions at the European level is a challenge that NIRS can help to take up.

Keywords: NIRS; Soil organic matter pools; Soil carbon sequestration; Soil mapping; Ontogeny of detritus; Decomposition; Soil aggregation; Local calibration

Introduction

When trying to assess greenhouse gas budget of soils (e.g. Cost Action 639), it is important to get accurate information on carbon stocks (CS) in soils at a global scale. Increasing carbon stocks in soils is possible through changes in land use management practices (Lal, 2004), but enhancing CS in soils can increase emissions of N₂O and CH₄ (Lal, 2004) which are other important greenhouse gases. Monitoring changes in soil CS with precision requires the determination of a baseline value (Lark *et al.*, 2006). At European level, a map of organic carbon in topsoils (Jones *et al.*, 2004) is already available from the European Soil Bureau based at the Joint Research Center (Ispra, Italy). This map defines the baseline status of soil organic carbon (SOC) in 1990. Although this map generated by pedo-transfer rules presents good correlations with measured values, its confidence level remains quite heterogeneous across large areas.

Current models of soil organic matter (SOM) dynamics need more detailed information describing the heterogeneity of SOM: the quantification of SOM pools with different turnover rates (Six *et al.*, 2002). Measuring SOM pools is time-consuming and seems unfeasible with a fine-resolution at a global scale. Thus, there is an urgent need for developing methods for determining SOC and SOM fractions that would allow the analysis of large number of samples necessary for monitoring purposes over large geographical areas.

Near-infrared reflectance spectroscopy (NIRS) is a rapid and non-destructive analytical technique that involves diffuse-reflectance measurement in the near-infrared region (1000 – 2500 nm). Near-infrared spectra (Figure 1) depend on the number and type of chemical bonds in the material being analysed (Foley *et al.*, 1998). NIR spectra can be used to predict various soil properties including C and N content in soils (e.g. Chang & Laird, 2002, Ludwig & Khanna, 2001). It has been recognized as a practical indicator and tool to measure soil health and their basic characteristics (Bunning & Jimenez, 2003). With the recent development of *in situ* NIR analysers, NIRS is now considered as a promising emerging technology for measurement of soil C (Gehl & Rice, 2007) in the field.

The aim of this short paper is to briefly review the capacity of NIRS to quantify soil C, N and functional SOM fractions from the field level to a global scale. First, we describe the methodology of NIRS for the analysis of soil samples. Then, we present most recent NIRS researches on the detailed study of two key soil processes controlling C and N stocks in soil: decomposition and aggregation of SOM. In a third part, we review the use of NIR data for soil C and N mapping. We finally list a number of key issues for future development of NIRS in routine analysis of soil C and N budget.

1. Implementation of NIRS in Soil Science

Sample processing for NIR analysis of soil is rather simple. Soil samples can be either air-dried or oven-dried at 40°C. Particle size and moisture content influence NIR spectra (Barthès *et al.*, 2006). Thus, soil samples are dry-sieved (2 mm), ground (0.25 mm) to obtain a homogeneous powder and stored in polypropylene boxes. The day before NIR analysis, powders are oven-dried at 40°C during the night to homogenize moisture and kept in a desiccator before analysis. From 1 g to 25 g of soil is required for NIR analysis, depending on the NIR analyser. Spectra consist of absorbance (absorbance = $\log [1 / \text{reflectance}]$) values from 1000 to 2500 nm. From 100 to 300 samples per day can be scanned.

The diffuse-reflectance spectra of soil result from bending, twisting, stretching or scissoring of different chemical bounds (*e.g.* C–H, O–H, N–H) under near infrared radiation (Ludwig & Khanna, 2001). Absorptions in the near-infrared are weak since they consists of overtones or combinations of fundamentals (Wetzel, 1983). Direct interpretation of NIR spectra is almost impossible : each NIR analysis require a statistical treatment. First, spectral data should be normalized with a mean centering technique and can be variance scaled. Then, the first or second derivative of spectra can be computed to reveal information hidden in the spectra, and also to reduce baseline drift. Smoothing the optical signal with derivative filters (mainly Norris or Savitzky-Golay) can help to lessen spectral noise as well as scattering corrections like multiplicative scatter

correction (MSC) or standard normal variate (SNV) correction. After pretreatment, statistical tools relevant to the aim of the analysis are implemented to analyse spectral data.

NIR analysis of soil can be run in two main ways (Figure 2): (i) qualitative or (ii) quantitative.

(i) Qualitative analysis of soil with NIRS can be used to discriminate among soil samples from a given population with principal component analysis of soil spectra (PCA) or redundancy analysis (RDA), build classes of soil samples with discriminant analysis (DA) or to study time series of soil samples under controlled conditions.

(ii) Quantitative analysis allows the prediction of various components and soil properties after a calibration of NIR spectra against reference values. Quantification with NIRS usually involves multivariate techniques (e.g. partial least square regression, PLS) that consist of regression method used to build prediction models (Dardenne *et al.*, 2000). Recently, a local approach for calibration of NIR prediction models (Brunet *et al.*, 2007, Dardenne *et al.*, 2000, Genot *et al.*, 2007, van Waes *et al.*, 2005) has attracted much attention. Local calibrations are carried out within spectral classes defined either on statistical distances among spectra (Mahalanobis distances) or directly upon soil types, texture groups or land use.

The quality of NIR prediction models is assessed with various indices (Fearn, 2002, Williams, 2001):

- SECV: standard error of cross-validation performed on the calibration data set
- SEP: standard error of prediction which is the standard deviation (SD) of differences between NIR reflectance and reference values, that should be calculated on real independent data set, but is usually calculated on a non independent validation data set
- RPD: the ratio of performance to deviation, (SD of reference values) / SEP
- R²: the coefficient of determination

2. Use of NIRS to study decomposition and aggregation of SOM

The amount of carbon returned to the atmosphere by decomposition of dead organic matter (OM) is a key component of the global carbon budget (Aerts, 1997). SOM decline is one of the major threat regarding soil degradation in Europe (Bellamy *et al.*, 2005, SEC(2006)1165). During soil formation, organisms create and manipulate detritus into complex SOM pools that contribute to OM protection against decomposition (*e.g.* Lavelle & Spain, 2001, Lützow *et al.*, 2006). Biological and physical aggregation of OM is one of the main process responsible for the creation of such SOM pools with different turnover rates (*e.g.* Blanco-Canqui & Lal, 2004). This heterogeneous nature of detritus that results from variation in sources, and the changes it undergoes during decomposition has recently been incorporated in a conceptual model of ontogeny of detritus by Moore *et al.* (2004). Finding an operational expression of the heterogeneity and ontogeny of detritus (Moore *et al.*, 2004) that could easily quantify functional SOM pools (Blanco-Canqui & Lal, 2004, Robert *et al.*, 2004) is a challenge for soil scientists.

2.1 Decomposition process

The suitability of NIRS in decomposition studies has been demonstrated by Joffre *et al.* (1992) for plant litter materials. Litter decomposability can be predicted with NIRS (Gillon *et al.*, 1999). It is also possible to discriminate among soil organic layers, predict their biological and chemical properties (Chodak *et al.*, 2002, Ludwig *et al.*, 2002) like C, N and SOM fractions including microbial biomass at different decomposition stages with this technology (Coûteaux *et al.*, 2003). It is also possible to predict the relative amount and the composition of litter and root mixtures with NIRS (M.C. Gruselle, unpublished PhD, Roumet *et al.*, 2006). Recently, Cécillon *et al.* (2007) have shown with NIRS that a holistic approach of decomposition is possible in order to obtain real-world decay constants for organic horizons of mix-stands. With a PCA of NIR spectra, it is possible to follow decomposition trajectories of detritus under different experimental conditions. The effect of soil mesofauna on litter decomposition can be quantified with PCA (Figure 3) which is the first example of a preliminary exploration of the ontogeny of detritus with NIRS.

2.2 Aggregation process

Soil invertebrates produce compact organic and organo-mineral faecal pellets (Lavelle & Spain, 2001). These animal faeces, which are major components of humic material can form stable SOM pools (Rawlins *et al.*, 2007). Hedde *et al.* (2005) have shown with PCA of NIR spectra that each soil biogenic structure has a specific NIRS signature, different from the surrounding soil. A change in decomposability of organic matter from leaves to faecal pellets of soil fauna has been revealed by NIRS (Gillon & David, 2001). Recently, Velasquez *et al.* (2007) have classified soil aggregates upon their origin (biogenic, physicogenic ; Figure 4) with PCA analysis of NIR spectra. A current research work at Cemagref Grenoble has shown a change in the NIR spectra of soil and biogenic structures of earthworms after forest wildfires (Master thesis of S. Figarol, IRISE project, Forest Focus). Spectral differences between soil and biogenic structures are reduced in plots impacted by recent wildfires.

All these NIRS studies show the potential of this technology for the study of macroscopic functional SOM pools. But current SOM models need the quantification of microscopic functional SOM pools, as a recalcitrant SOM pool as been identified in microaggregates within soil macroaggregates (Six *et al.*, 2002). These microscopic functional SOM fractions can be quantified with NIRS (Cozzolino & Moron, 2006) as well as their respective C, N content and mineralization rates (Mutuo *et al.*, 2006). However, prediction of these functional SOM pools with NIRS have to be confirmed on larger data sets, at the regional scale.

Finally, the power of NIRS in SOM studies have been demonstrated by Joffre *et al.* (2001) who managed to link experimental NIR spectra to SOM theoretical quality, q (Q model, Ågren & Bosatta, 1998).

3. Mapping of soil C and N with NIRS

Mapping soil C and N budget of soil at a regional scale implies analysing large number of samples. It has been estimated that monitoring soil C in all US farms, with existing analytical techniques would cost more than \$1 billion for the first 4-yr period (Subak, 2000). A reduction in per-sample

cost analysis for soil C and N, coupled with a quicker sampling and sample preparation method would be welcome (Martin *et al.*, 2002). That's why NIRS has recently been tested for mapping soil C and N at the field and regional scale.

At the field level, Martin *et al.* (2002) found useful correlations for SOC using PCA and PLS regression of NIR spectra. However, they failed at predicting soil organic nitrogen with the same experimental design. Their conclusion is that NIRS has potential to be a suitable method for SOC mapping for the purposes of monitoring C sequestration. A study of Yong *et al.* (2005) has shown good correlations between a map of SOM predicted by NIRS and a map of SOM measured with a conventional method. This research found also a good prediction of total soil N with NIRS. Odlare *et al.* (2005), used a NIR-PCA strategy to determine soil spatial variation in a field. They compared maps of soil properties with maps of PCA scores of NIR spectra and concluded that this technique can be useful in precision farming.

At the regional scale, a recent study of Genot *et al.* (2007) used NIRS to predict soil properties including SOC on a large data set of 946 samples representative of the soil diversity of the Wallonia region (Belgium). They tested various calibration method (global and two local calibration techniques) and found that local calibrations usually improve the quality of NIR prediction models (reduction of SECV).

Prediction of SOM at the global level could be facilitated by the use of portable NIR analyser. Gehl and Rice (2007) have classified NIRS among emerging technologies for *in situ* assessment of soil carbon. But the quality of NIR measurements in the field is still in debate, and NIR laboratory measurements are very rapid and simple. Maybe linking NIR field measurements to NIR remote sensing measurements from aircraft could be a suitable way of mapping soil properties over large geographical areas (Lagacherie *et al.*, 2007). Another promising approach for mapping soil properties is to use predictions made with spectral data in inference systems to predict and map other important soil properties (McBratney *et al.*, 2006).

A drawback of most studies is that prediction capacity of NIRS is usually not tested on a real independent data set of soil samples. A research project carried out within the framework of Cost Action 639 – BurnOut will test the suitability of NIRS for SOC mapping on independent data at

the field scale (Short Term Scientific Mission – STSM of L. Cécillon at Cranfield University, UK, with P. Bellamy). Another promising way of mapping SOM could be the use of NIR spectra (PCA scores) of topsoil as secondary information to improve CS maps (e.g. Simbahan *et al.*, 2006). This approach will also be tested during the same forthcoming STSM within Cost Action 639.

4. Future directions for NIRS studies in Soil Science

Diffuse reflectance spectroscopic techniques have a real potential for analysing simultaneously various soil properties on large number of samples (Viscarra Rossel *et al.*, 2006). In this review and another recent paper of Reeves *et al.* (2006), the quality of NIR prediction models is compared to mid-infrared technology (2500 – 25000 nm). These two studies found that both technologies have potential for measuring soil properties but they underlined that MIR predictions were sometimes better. However, these researches didn't use the promising local calibration technique that can be decisive for the improvement of NIR calibration models (Genot *et al.*, 2007).

Although we pointed out the need for an assessment of prediction capacity of NIRS on independent data, we strongly believe that one of the most important topic in NIRS studies is the generalization of local calibration. That implies building large database of spectral data representative of soil diversity in Europe that could be shared between laboratories. To achieve this goal properly, the processing of samples (sieving, grinding, oven-drying and scanning) should be standardized to ensure that spectral data are comparable among laboratories. All promising techniques described in this paper suffer from the small number of samples used for calibration and validation of NIR prediction models, the lack of studies at a regional or European level and the use of the global calibration method which is generally not the best way in soil studies. Thus, a key issue would be to test their suitability over large areas using local calibration.

Regarding the assessment of greenhouse gas emissions from European soils which is the aim of Cost Action 639 – BurnOut, a challenge of NIRS studies is the quantification of functional SOM pools. Such NIR prediction models should be validated over large geographical areas, with a fine-resolution and directly interoperable with global models of C and N cycles.

Acknowledgments

This work was supported by the French Agency for Environment and Energy Management (ADEME), Cemagref and the Cost Office. We would like to thank Richard Joffre (CEFE-CNRS Montpellier, France, IRD) for introducing us to NIR technology. Robert Jandl, Patricia Bellamy and the Cost Office are thanked for their help in obtaining a STSM grant on NIRS within Cost Action 639. We also thank Cemagref for developing NIRS in the Mountain Ecosystems Research Unit within the INGECO project of Envirhonalp.

References

- Aerts, R. (1997) Climate, Leaf Litter Chemistry and Leaf Litter Decomposition in Terrestrial Ecosystems: A Triangular Relationship. *Oikos* **79**: 439-449.
- Ågren, G.L. & Bosatta, E. (1998) Theoretical ecosystem ecology: understanding element cycles. Cambridge University Press.
- Barthès, B.G., Brunet, D., Ferrer, H., Chotte, J.L. & Feller, C. (2006) Determination of total carbon and nitrogen content in a range of tropical soils using near infrared spectroscopy: influence of replication and sample grinding and drying. *Journal of Near Infrared Spectroscopy* **14**: 341-348.
- Bellamy, P.H., Loveland, P.J., Bradley, R.I., Lark, R.M. & Kirk, G.J.D. (2005) Carbon losses from all soils across England and Wales 1978-2003. *Nature* **437**: 245-248.
- Blanco-Canqui, H & Lal, R. (2004) Mechanisms of carbon sequestration in soil aggregates. *Critical Reviews in Plant Sciences* **23**: 481-504.
- Brunet, D., Barthès, B., Chotte, J.L. & Feller, C. (2007) Determination of carbon and nitrogen contents in Alfisols, Oxisols and Ultisols from Africa and Brazil using NIRS analysis: Effects of sample grinding and set heterogeneity. *Geoderma*, *in press*.

- Bunning, S. & Jiménez, J.J. (2003) Indicators and assessment of soil biodiversity / soil ecosystem functioning for farmers and governments. Paper presented at the OECD Expert Meeting on indicators of soil erosion and soil biodiversity, 25-28 March 2003, Rome, Italy: 22 pp.
- Cécillon, L., Cassagne, N. & Brun, J.J. (2007) Décomposition de la litière: approche holistique et utilisation de la spectroscopie proche infrarouge (NIRS). (In French). Actes des 9^{èmes} Journées Nationales de l'Etude des Sols, AFES, 3-5 April 2007, Angers, France: 163-164.
- Chang, C.W. & Laird, D.A. (2002) Near-infrared reflectance spectroscopic analysis of soil C and N. *Soil Science* **167**: 110-116.
- Chodak, M., Ludwig, B., Khanna, P. & Beese, F. (2002) Use of near infrared spectroscopy to determine biological and chemical characteristics of organic layers under spruce and beech stands. *Journal of Plant Nutrition and Soil Science* **165**: 27-33.
- Coûteaux, M.M., Berg, B. & Rovira, P. (2003) Near infrared reflectance spectroscopy for determination of organic matter fractions including microbial biomass in coniferous forest soils. *Soil Biology and Biochemistry* **35**: 1587-1600.
- Cozzolino, D. & Moron, A. (2006) Potential of near-infrared reflectance spectroscopy and chemometrics to predict soil organic matter fractions. *Soil & Tillage Research* **85**: 78-85.
- Dardenne, P., Sinnaeve, G. & Baeten, V. (2000) Multivariate calibration and chemometrics for near infrared spectroscopy : Which method ? *Journal of Near Infrared Spectroscopy* **8**: 229-237.
- Fearn, T. (2002) Assessing calibrations : SEP, RPD, RER and R². *NIR news* **13**: 12-14.
- Foley, W.J., McIlwee, A., Lawler, I.R., Aragones, L., Woolnough, A. & Berding, N. (1998) Ecological applications of near-infrared spectroscopy - a tool for rapid, cost-effective prediction of the composition of plant and animal tissues and aspects of animal performance. *Oecologia* **116**: 293-305.
- Gehl, R.J. & Rice, C.W. (2007) Emerging technologies for in situ measurement of soil carbon. *Climatic Change* **80**: 43-54.
- Genot, V., Colinet, G., Dardenne, P., Bah, B., Legrain, X. & Bock, L. (2007) Etude de la performance de la spectroscopie proche infrarouge pour la mesure de caractéristiques du sol

nécessaires au conseil de fumure. (In French). Actes des 9^{èmes} Journées Nationales de l'Etude des Sols, AFES, 3-5 April 2007, Angers, France: 379-380.

Gillon, D., Joffre, R. & Ibrahima, A. (1999) Can litter decomposability be predicted by near infrared reflectance spectroscopy? *Ecology* **80**: 175-186.

Gillon, D. & David, J.F. (2001) The use of near infrared reflectance spectroscopy to study chemical changes in the leaf litter consumed by saprophagous invertebrates. *Soil Biology & Biochemistry* **33**: 2159-2161.

Hedde, M., Lavelle, P., Joffre, R., Jiménez, J.J. & Decaëns, T. (2005) Specific functional signature in soil macro-invertebrates biostructures. *Functional Ecology* **19**: 785-793.

Joffre, R., Gillon, D., Dardenne, P., Agneessens, R. & Biston, R. (1992) The use of near-infrared reflectance spectroscopy in litter decomposition studies. *Annales des Sciences Forestières* **49**: 481-488.

Joffre, R., Ågren, G.I., Gillon, D. & Bosatta, E. (2001) Organic matter quality in ecological studies: theory meets experiment. *Oikos* **93**: 451-458.

Jones, R.J.A., Hiederer, R., Rusco, E., Loveland, P.J. & Montanarella, L. (2004). The map of organic carbon in topsoils in Europe, Version 1.2, September 2003: Explanation of Special Publication Ispra 2004 No.72 (S.P.I.04.72). European Soil Bureau Research Report No.17, EUR 21209 EN, 26pp. and 1 map in ISO B1 format. Office for Official Publications of the European Communities, Luxembourg.

Lagacherie, P., Baret, F., Feret, J.B., Madeira-Netto, J., Robbez-Masson, J.M. (2007) Utilisation de spectres de reflectance pour estimer les taux d'argile et de carbonate de calcium de la surface de sol. Performances comparées des mesures de laboratoire, de terrain et par télédétection. (In French). Actes des 9^{èmes} Journées Nationales de l'Etude des Sols, AFES, 3-5 April 2007, Angers, France: 131-132.

Lal, R. (2004) Soil carbon sequestration impacts on global climate change and food security. *Science* **304**: 1623-1627.

Lark, R.M., Bellamy, P.H. & Kirk, G.J.D. (2006) Baseline values and change in the soil, and implications for monitoring. *European Journal of Soil Science* **57**: 916-921.

- Lavelle, P. & Spain, A.V. (2001) *Soil Ecology*. Kluwer Academic Publishers.
- Ludwig, B. & Khanna, P.K. (2001) Use of near infrared spectroscopy to determine inorganic and organic carbon fractions in soil and litter. *In: Assessment Methods for Soil Carbon*. Lal, R., Kimble, J.M, Follet, R.F. & Stewart, B.A. (eds.). Lewis Publ., Boca Raton, FL. Chapter 25: 361-370.
- Ludwig, B., Khanna, P., Bauhus, J. & Hopmans, P. (2002) Near infrared spectroscopy of forest soils to determine chemical and biological properties related to soil sustainability. *Forest Ecology and Management* **171**: 121-132.
- Lützw, M., Kögel-Knabner, I., Ekschmitt, K., Matzner, E., Guggenberger, G., Marschner, B. & Flessa, H. (2006) Stabilization of organic matter in temperate soils: mechanisms and their relevance under different soil conditions - a review. *European Journal of Soil Science* **57**: 4: 426-445.
- Martin, P.D., Malley, D.F., Manning, G. & Fuller, L. (2002) Determination of soil organic carbon and nitrogen at the field level using near-infrared spectroscopy. *Canadian Journal of Soil Science* **82**: 4: 413-422.
- McBratney, A.B., Minasny, B. & Viscarra Rossel, R.A. (2006) Spectral soil analysis and inference systems: a powerful combination for solving the soil data crisis. *Geoderma* **136**, 272-278.
- Moore, J.C., Berlow, E.L., Coleman, D.C., de Ruiter, P.C., Dong, Q., Hastings, A., Collins Johnson, N., McCann, K.S., Melville, K., Morin, P.J., Nadelhoffer, K., Rosemond, A.D., Post, D.M., Sabo, J.L., Scow, K.M., Vanni, M.J. & Wall, D.H. (2004) Detritus, trophic dynamics and biodiversity. *Ecology Letters* **7**: 584-600.
- Mutuo, P.K., Shepherd, K.D., Albrecht, A. & Cadisch, G. (2006) Prediction of carbon mineralization rates from different soil physical fractions using diffuse reflectance spectroscopy. *Soil Biology and Biochemistry* **38**: 1658-1664.
- Odlare, M., Svensson, K. & Pell, M. (2005) Near infrared reflectance spectroscopy for assessment of spatial soil variation in an agricultural field. *Geoderma* **126**: 193-202.
- Rawlins, A.J., Bull, I.D., Ineson, P. & Evershed, R.P. (2007) Stabilisation of soil organic matter in invertebrate faecal pellets through leaf litter grazing. *Soil Biology and Biochemistry* **39**: 1202-1205.

Reeves III, J.B., Follett, R.F., McCarty, G.W., Kimble, J.M. (2006) Can near- or mid-infrared diffuse reflectance spectroscopy be used to determine soil carbon pools? *Communications in Soil Science and Plant Analysis* **37**: 2307-2325.

Robert, M., Nortcliff, S., Yli-Halla, M., Pallière, C., Baritz, R., Leifeld, J., Bannick, C.G., Chenu, C. (2004). Organic matter and biodiversity. Task Group 1 on functions, roles and changes in SOM. *In*: Reports of the technical working groups established under the thematic strategy for soil protection. Van-Camp, L., Bujarrabal, B., Gentile, A.R., Jones, R.J.A., Montanarella, L., Olazabal, C. & Selvaradjou, S. (eds). European commission, JRC, European Environment Agency.

Roumet, C., Picon-Cochard, C., Dawson, L.A., Joffre, R., Mayes, R., Blanchard, A. & Brewer, M.J. (2006) Quantifying species composition in root mixtures using two methods: near-infrared reflectance spectroscopy and plant wax markers. *New Phytologist* **170**: 631-638.

SEC(2006)1165 (2006) Commission staff working document. Accompanying document of the communication from the commission to the Council, the European parliament, the European economic and social committee and the committee of the regions. Thematic strategy for soil protection. Summary of impact assessment, 8 pp.

Simbahan, G.C., Dobermann, A., Goovaerts, P., Ping, J. & Haddix, M.L. (2006). Fine-resolution mapping of soil organic carbon based on multivariate secondary data. *Geoderma* **132**: 471-489.

Six, J., Conant, R.T., Paul, E.A. & Paustian, K. (2002) Stabilization mechanisms of soil organic matter: Implications for C-saturation of soils. *Plant and Soil* **241**: 155-176.

Subak, S. (2000) Agricultural soil carbon accumulation in North America: considerations for climate policy. *Global Environmental Change* **10**: 185-195.

Viscarra Rossel, R.A., Walvoort, D.J.J., McBratney, A.B., Janik L.J. & Skjemstad, J.O. (2006). Visible, near-infrared, mid-infrared or combined diffuse reflectance spectroscopy for simultaneous assessment of various soil properties. *Geoderma* **131**: 59-75.

Velasquez, E., Pelosi, C., Brunet, D., Grimaldi, M., Martins, M., Rendeiro, A.C., Barrios, E. & Lavelle, P. (2007) This ped is my ped: Visual separation and near infrared spectra allow determination of the origins of soil macroaggregates. *Pedobiologia* **51**: 75-87.

van Waes, C., Mestdagh, I., Lootens, P. & Carlier, L. (2005) Possibilities of near infrared reflectance spectroscopy for the prediction of organic carbon concentrations in grassland soils. *Journal of Agricultural Science* **143**: 487-492.

Wetzel, D.L. (1983) Near-infrared reflectance analysis. Sleeper among spectroscopic techniques. *Analytical Chemistry* **55**, **12**: 1165 A-1176 A.

Williams, P.C. (2001) Implementation of near-infrared technology. In: Near infrared technology in the agricultural and food industries (second edition), Williams, P.C. & Norris, K. (eds). Chapter 8: 145-169.

Yong, H., Song, H., Pereira, A.G., Gomez, A.H. (2005) Measurement and analysis of soil nitrogen and organic matter content using near-infrared spectroscopy techniques. *Journal of Zhejiang University Science*, **6B**(11): 1081-1086.

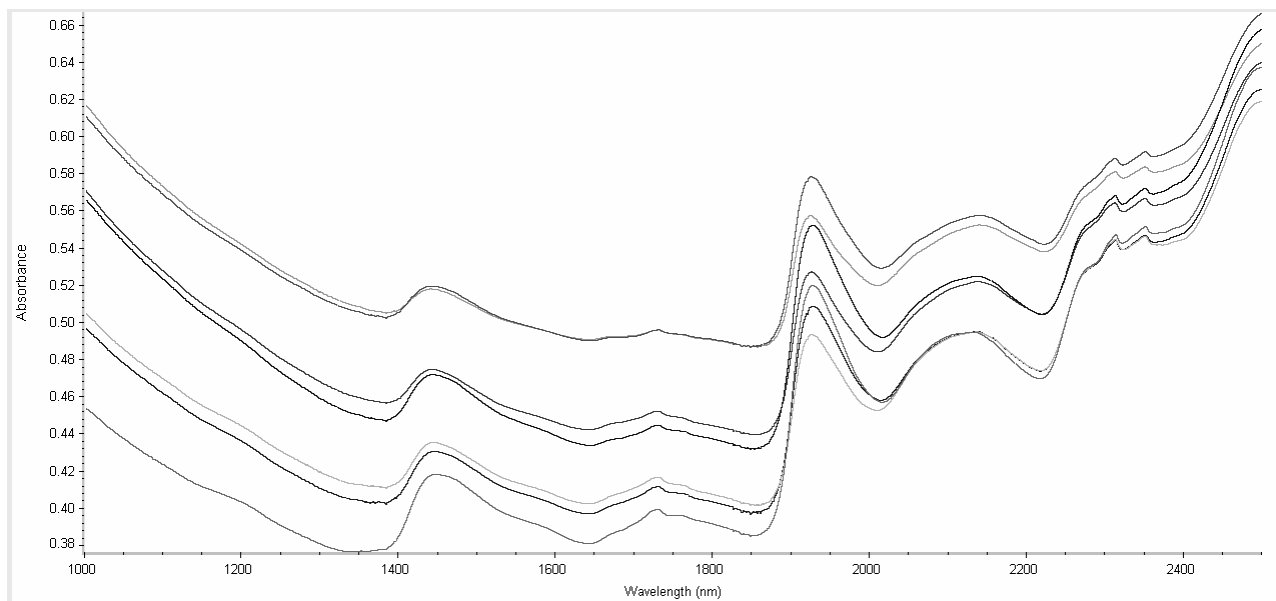


Figure 1: Examples of NIR spectra of forest topsoils –
Data acquisition with an Antaris II NIR Analyser (Thermo electron corporation)

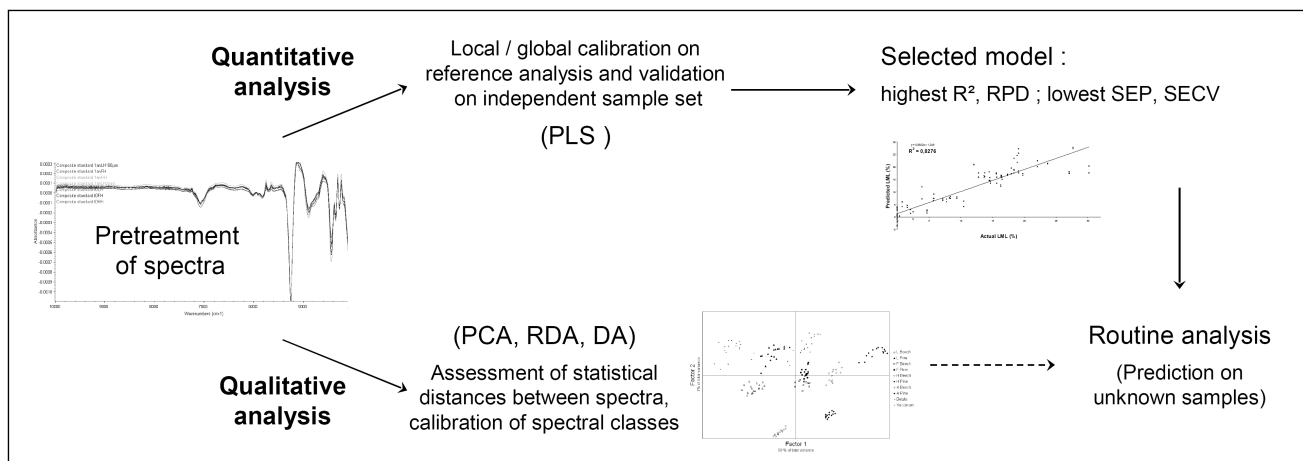


Figure 2: Quantitative and qualitative analysis of NIR spectra

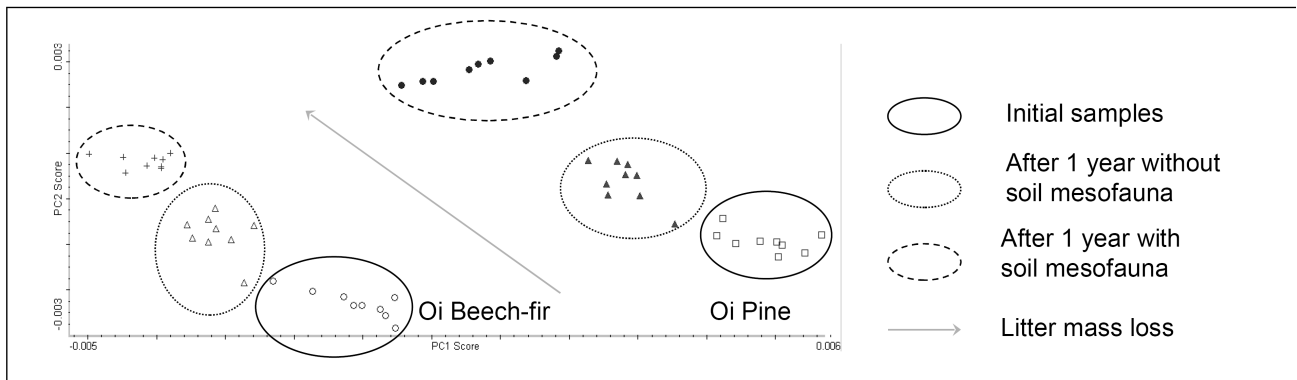


Figure 3: Impact of soil mesofauna on the ontogeny of forest litter (PCA of NIR spectra; PC1: 72% of total variance, PC2: 25% of total variance)

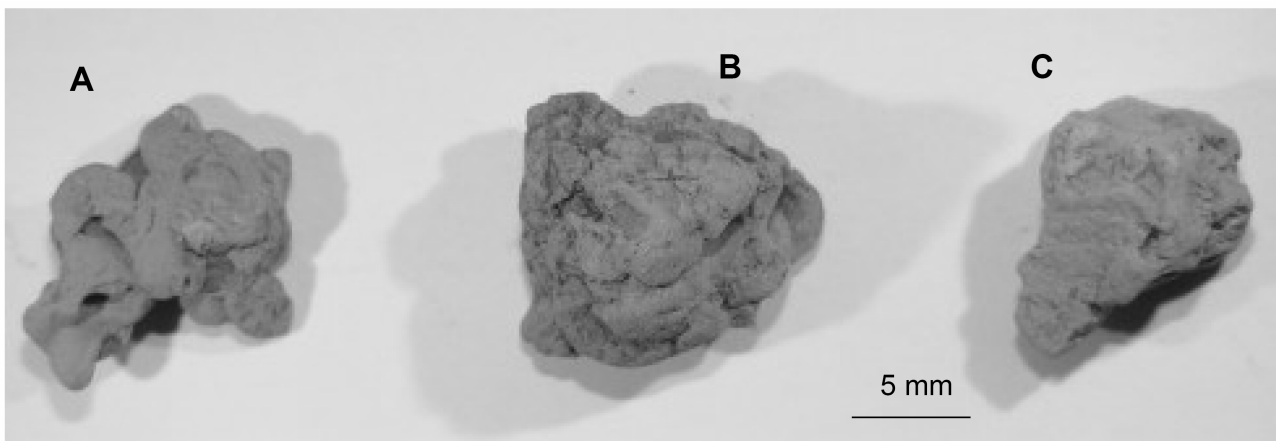


Figure 4: Soil macroaggregates
 (A) Biogenic
 (B) Intermediate
 (C) Physcogenic