

# Comparative bioaccumulation of trace elements between Nautilus pompilius and N. macromphalus (Cephalopoda: Nautiloidea) from Vanuatu and New-Caledonia

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27 **ABSTRACT:** The concentrations of sixteen trace elements were investigated and compared 28 for the first time in the digestive and excreting tissues of two Nautilus species (Cephalopoda: 29 Nautiloidea) from two geologically contrasted areas: 1) N. macromphalus from New 30 Caledonia, a region characterised by its richness in nickel ores and its lack of tectonic 31 activities and 2) N. pompilius from the Vanuatu archipelago showing high volcanic and 32 tectonic activities. In both *Nautilus* species, results clearly highlighted that the digestive gland 33 played a key role in the bioaccumulation and storage of Ag, Cd, Ce, Co, Cu, Fe, La, Nd, V 34 and Zn whereas As, Cr, Mn, Ni, Pb and Se were accumulated in a greater extent in the 35 excreting tissues (i.e. pericardial and renal appendages). Despite contrasting environments, no 36 significant difference (P<0.05) was found between the two Nautilus species in the 37 concentrations of most of the essential and non essential elements, including Ni and 38 associated metals in Ni ores (i.e. Co and Mn). As nautilus lives on the outer shelf of barrier reefs, these results strongly support the hypothesis that the New Caledonian lagoon traps the 39 40 major amount of the trace elements derived from natural erosion and the intense mining 41 activities conducted on land. In contrast, the concentrations of the rare earth elements (Ce, La, 42 and Nd) were significantly higher in N. pompilius than in N. macromphalus, probably as a 43 result of the local enrichment of Vanuatu waters by specific environmental processes, such as 44 volcanism or upwelling.

45

46 Keywords: metals, metalloids, cephalopods, detoxification, excretion, South Pacific

#### 47 **1. INTRODUCTION**

48

49 Marine organisms are exposed to trace elements that are present in their diet and dissolved in 50 seawater. This double exposure results in element accumulation in their different tissues 51 whether or not these elements are essential to the metabolism (Rainbow, 2002). This process, 52 widely described as bioaccumulation can vary greatly according to the specificities of the 53 organisms such as age/size, sex and lifestyle, and according to the relative bioavailability of 54 the metal in diet and seawater (Rainbow and Wang, 2001). Comparative analysis of trace element concentrations between closely related species living in different geographical areas 55 56 can therefore allow at assessing the influence of the environment on their bioaccumulation.

57 Among the marine species, cephalopods belong to a molluscan group comprising ca. seven 58 hundred species (Boyle and Rodhouse, 2005), in which high concentrations of trace elements 59 such as Ag, Cd, Cu or Zn have generally been recorded (Martin and Flegal, 1975; Finger and 60 Smith, 1987; Miramand and Bentley, 1992; Yamada et al., 1997; Bustamante et al., 1998; 61 Dorneles et al., 2007). Indeed, several studies have demonstrated that the digestive and the 62 excretory organs of cephalopods play a major role in the bioaccumulation of trace elements, 63 as these organs are deeply involved in the assimilation processes, detoxification and storage 64 of both essential and non-essential metals (Miramand and Guary, 1980; Miramand and 65 Bentley, 1992; Bustamante et al., 2000, 2002a,b, 2004).

In the present work, we focused on the bioaccumulation of trace elements in the *Nautilus* genus. Nautiluses are scavenger cephalopods, inhabiting the outer shelf of barrier reefs in the Indo-Pacific area, generally at a depth of 300 to 500 meters (O'Dor et al., 1993; Norman, 2000). In spite of its evolutionary interest (Nautilus is the last representative of ectocochleate cephalopods, i.e. considered as a "living fossil"; Boyle and Rodhouse, 2005) and its physiological specificities (slow digestion and most of the excretory processes taking place in 72 the pericardial appendages; Schipp et al., 1985; Westermann et al., 2002), only very few studies deal with the bioaccumulation of trace elements in this ancestral genus. High 73 74 concentrations of several metals and metalloids were reported in the digestive and excretory 75 tissues of Nautilus macromphalus (Bustamante et al., 2000), a species endemic to the New 76 Caledonian waters. These authors suggested that the high metal concentrations may be due to 77 a high exposure in the metal-rich conditions in the surrounding New Caledonian waters. Indeed, the largest resources of Ni as laterites in the world are present in New Caledonia 78 where open-cast mining is conducted since the 20<sup>th</sup> century, leading to a dramatic increase of 79 80 metal concentrations in coastal waters and, subsequently, the metal burdens in marine biota 81 (Labrosse et al., 2000; Metian et al., 2008). However, it is not clear whether metal 82 accumulation observed in N. macromphalus reflected the impact of metal contamination 83 through human activities on the outer shelf waters of New Caledonia or either if it is more due 84 to a natural progressive bioaccumulation during nautilus long life span (viz. 10-15 years).

85

86 The aim of the present study was therefore to determine the bioaccumulation of a large range 87 of elements in the genus Nautilus comparing two species from geologically contrasted areas: 88 1) N. macromphalus from New Caledonia which is characterised by its richness in nickel ores 89 and its lack of tectonic activities and 2) N. pompilius from the Vanuatu archipelago which 90 shows high volcanic and tectonic activities (Harrison et al, 1996). The analysed elements 91 comprised 11 metals (Ag, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, V and Zn), 2 metalloids (As and 92 Se) and 3 rare earth elements (Ce, La and Nd). These elements were selected for comparison 93 with other species and other published works concerning Nautilus (e.g. Miramand et al. 1992, 94 Bustamante et al. 2000, Ichihashi et al. 2001). The concentrations and relative contents of 95 these different trace elements were determined in the digestive and the excretory tissues (i.e. pericardial and renal appendages) and compared between both nautilus species. The 96

97 specificities and the potential influence of the environment on bioaccumulation of these trace98 elements in the *Nautilus* genus are discussed.

99

#### 100 2. MATERIALS AND METHODS

#### 101 a) Biological material

Three *Nautilus pompilius* and five *Nautilus macromphalus* specimens were collected respectively off Santo island coast (Vanuatu, 15° 40' 522 S, 167° 02' 838 E, depth: 334m) and on the outer shelf of New-Caledonia (22° 20' 500 S, 166° 13' 400 E, depth: 300m) in May 2006 (Figure 1). Each individual was measured and weighted prior to dissection. The main features of the eight specimens sampled are reported in Table 1.

All specimens were dissected aseptically and for each specimen one pericardial appendage,
one renal appendage and a fraction of the digestive gland were immediately frozen in liquid
nitrogen and stored at -80°C for metal analyses.

110

#### 111 b) Analytical procedure

Tissue samples were freeze-dried. Aliquots of the samples ranging from 20 to 300 mg were digested using a 3:1 v:v nitric-hydrochloric acid mixture with 65% ultrapure HNO<sub>3</sub> and ultrapure 70% HCl. The acidic digestion was performed overnight under ambient temperature and then heated in a microwave during 30 minutes, with increasing temperature until 105°C, and 15 minutes at 105°C (1200 Watts). After the mineralization process, each sample was diluted to 30 ml or 50 ml with milli-Q quality water, according to the volume of acid added to the mineralization (3 ml and 4.5 ml. respectively).

Ag, As, Cd, Ce, Co, Cr, Cu, Fe, La, Mn, Nd, Ni, Pb, Se, V and Zn were analysed either by
ICP-OES (Varian<sup>®</sup> Vista-Pro) and ICP-MS (Varian<sup>®</sup> Ultra Mass 700). Reference tissues,
dogfish liver DOLT-3 (NRCC), lobster hepatopancreas TORT-2 (NRCC) and Bush Branches

122 and Leaves (GBW 07602) were treated and analysed in the same way as the samples. Results 123 were in good agreement with the certified values, and the standard deviations were low, 124 proving good repeatability of the method. The results for standard reference materials 125 displayed recoveries of the elements ranging from 81% to 119% (n=10). For each set of analyses, blanks were included in each analytical batch. The detection limits ( $\mu g.g^{-1}$  dry wt) 126 127 for ICP-OES were 8.3 (As, Fe, Zn), 3.3 (Ag, Se), 1.67 (Pb, V), 0.83 (Cd, Co, Cr, Cu, Mn, Ni), 128 and for ICP-MS, they were 0.150 (Ni, V), 0.065 (Cd, Co, Cr, Cu, Mn, Pb), 0.033 (Ag), 0.017 129 (Nd), 0.008 (Ce, La). Average element concentrations are presented with standard deviations. All trace element concentrations are given on a dry weight basis ( $\mu g.g^{-1}$  dry wt). 130

131

#### 132 Data analyses

133 Statistical analyses were performed using MINITAB 13.2 Software. Comparison of trace 134 element concentrations in nautilus tissues between sites was tested by 1-way ANOVA. 135 Hypothesis of normal distribution was tested using the Anderson-Darling test and equality of 136 variance by the Bartlett test. Pearson coefficients were calculated between trace elements in 137 each tissue. The significance level for statistical analyses was always set at  $\alpha = 0.05$ .

138

#### 139 **3. RESULTS**

140

#### 141 Trace element concentrations in the tissues

Metal concentrations in the digestive gland, the renal appendages and the pericardial appendages of *N. macromphalus* and *N. pompilius* are reported in Table 2. La, Ce, and Nd concentrations fell below the detection limit in the renal appendages. Among the three tissues analysed, the digestive gland was the major site of accumulation for Ag, Cd, Ce, Co, Cu, Fe, La, Nd, V and Zn (Table 2). The digestive gland also concentrated As and Ni at 147 concentrations close to the highest ones recorded in the other tissues. As, Cr, Mn, Ni, Pb, and 148 Se concentrations were remarkable in excreting tissues. More specifically, the renal 149 appendages exhibited the highest Mn and Se concentrations, whereas As, Ni, and Pb were 150 concentrated at similar concentrations within both renal and pericardial appendages. Finally, 151 the pericardial appendages were the major site of accumulation for Cr.

152

#### 153 *Comparison between species*

Generally, the pattern of trace element concentrations was the same in both *N. macromphalus* and *N. pompilius* tissues. However, there were few exceptions and more particularly in the digestive gland: firstly, Ce, Fe, La, and Nd concentrations were significantly (P<0.05) higher in *N. pompilius;* secondly, Cd and Cr concentrations were significantly (P<0.05) higher in *N. macromphalus* (Table 3).

In the case of the pericardial appendages, As, La, and Nd concentrations were significantly higher (P<0.05) in *N. pompilius* (Table 3). For the renal appendages, Cu was the only element for which a significant difference was detected, *N. pompilius* having significantly higher concentrations than *N. macromphalus* (Table 3).

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#### 164 *Relative proportion of trace elements between tissues*

Among the three tissues considered, the digestive gland contained the largest proportions of most of the trace elements with 73 to 81% of Ag, 90 to 91% of Cd, 61 to 79% of Ce, 80 to 86% of Co, 67 to 72% of Cu, 90 to 93% of Fe, 72 to 81% of La, 55 to 75% of Nd, 56 to 67 of V, 81% to 82% of Zn in both *Nautilus* species (Fig. 2). The only exception were As, Cr, Mn, Ni, Se, and Pb which were mainly contained in the excreting tissues, i.e. renal and pericardial appendages (Fig. 2). The renal appendages were the major site of accumulation of Mn, Ni, and Se (i.e. 74 to 89% for Mn, 49 to 54% for Ni; 61 to 73% for Se) while the pericardial appendages accumulated the highest relative proportion of Cr (i.e. 63 to 74%). Although the
concentrations of some of the trace elements were high in the excretory organs, these tissues
contained in fact low amounts of metals because of their small masses.

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#### 176 *Correlations between metals*

Table 4 shows the correlation between trace elements within the three tissues. It is noteworthy that the rare earth elements were correlated between each other within the digestive gland (r> 0.985 and P<0.001 in all cases) and the pericardial appendages (r> 0.922 and P<0.003 in all cases). In the digestive gland, rare earth elements varied positively with Fe and Mn (r> 0.918 and r>0.807 and P<0.004 and P<0.028, respectively) but negatively with As (r> -0.726 and P=0.047). Cu correlated positively with Zn (r= 0.834 and P=0.020 in all cases) but negatively with Se in this tissue (r=-0.809 and P= 0.028 in all cases).

In the renal appendages, correlations were weak, Mn varied positively with Cd (r= 0.833, P=0.010) and Zn (r= 0.735, P=0.038), Fe with Zn (r= 0.833, P=0.010) and Pb with Cr (r=0.936, P=0.001) but negatively with V (r= -0.753, P=0.031; Table 3).

In the pericardial appendages, Cd concentrations correlated positively with Cr (r = 0.897, P=0.006), Ni (r= 0.965, P<0.001), and V (r= 0.931, P=0.002).

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#### 190 4. DISCUSSION

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192 The present extensive analysis of sixteen trace element concentrations in *N. macromphalus* 193 and *N. pompilius* digestive and excreting tissues clearly highlighted that these organs 194 constitute important tissues for the bioaccumulation of rare earth elements, metals and 195 metalloids. Such results support the hypothesis of efficient regulation processes for non essential metals in Nautilidae (Bustamante et al., 2000) even if, among the eight studiedspecimens, the variability of metal concentrations was relatively high (Table 2).

198 In cephalopods, the digestive system plays a major role in both the digestive and the 199 detoxification processes: on the one hand, the digestive gland synthesizes the digestive 200 enzymes and is a place of absorption and assimilation of nutrients (Boucaud-Camou and 201 Boucher-Rodoni, 1983; Boucher-Rodoni and Boucaud-Camou, 1987); on the other hand, this 202 organ is involved in the storage and detoxification of trace elements (Bustamante et al., 203 2002ab, 2004). Our investigations on trace element concentrations also confirmed the key role 204 of the digestive gland of both N. macromphalus and N. pompilius in the metabolism of most 205 of the analysed trace elements (i.e. Ag, Cd, Ce, Co, Cu, Fe, La, Nd, V and Zn), as it is also the 206 case in more recent cephalopods, i.e. belonging to the Coleoidae (e.g. Ghiretti-Magaldi et al., 207 1958; Martin and Flegal, 1975; Miramand and Guary, 1980; Smith et al., 1984; Finger and 208 Smith, 1987; Miramand and Bentley, 1992; Seixas et al., 2005a; Pierce et al., 2008).

209 Our results also stressed the potential of excreting tissues (i.e. renal and pericardial 210 appendages) to accumulate several elements such as As, Cr, Mn, Ni, Pb, and Se. In contrast to 211 the Coleoidae, in nautiluses the renal appendages are specialized in the storage of calcium and 212 the pericardial appendages play a major role in the evacuation of the metabolic wastes 213 (Schipp and Martin, 1987). These organs are known to harbour a dual bacterial symbiosis 214 (Pernice et al., 2007) involving spirochaetes and betaproteobacteria. The exact mechanisms 215 by which these bacterial symbionts are involved in nautilus excretion are still enigmatic. 216 Nevertheless, it is widely admitted that micro-organisms can play an important role in the 217 incorporation of trace elements (Worms et al., 2006). Accumulation of Co, Fe and Ni by 218 betaproteobacterial symbionts of the pericardial appendages of *N. macromphalus* has already 219 been observed by electronic microscopy (Pernice et al., in preparation). However, it remains 220 unclear if this accumulative process is only passive (viz. through adsorption processes) or active (viz. effective absorption of trace elements). Indeed, the betaproteobacterial symbionts present in nautilus tissues could play a role in the detoxification and storage of some metals such as Co, Fe and Ni (Table 2). Such processes could involve enzymatic reduction and/or complexation with organic compounds (Pernice et al., 2006), leading to the storage of these metals at high concentrations in the excreting tissues, independently of the nautilus species. The role of this bacterial symbiosis on trace element bioaccumulation and detoxification in nautilus excreting tissues clearly deserves further investigations.

228 Although Nautilidae have been previously reported for their special abilities to accumulate heavy metals at relatively elevated concentrations in their tissues (Bustamante et al., 2000), 229 230 this study is the first comparative approach aiming to compare trace element concentrations in 231 two Nautilus species from two geographically close areas (viz. distant from less than 1000 232 kms). Both species potentially share common trophic features but, at the same time, they live 233 in geologically different environments (richness in nickel ores and lack of tectonic activities 234 in New Caledonia and high volcanic and tectonic activities in Vanuatu), potentially resulting 235 in a different degree of "natural" contamination. New Caledonia is currently the third largest 236 producer of Ni in the world (Dalvi et al., 2004) and mining activities associated to a strong 237 natural erosion have resulted in the enrichment of Ni and associated metals in Ni ores (i.e. Co, 238 Cr, Cu, Fe and Mn) and some other extracted metals (i.e. Ag and Pb) in the coastal waters and 239 consequently in the lagoon food webs (Monniot et al., 1994; Fernandez et al., 2006; Hédouin 240 et al., 2006; Metian et al., 2008). Conversely, it is likely that the seawater concentrations of most of these metals are very low in Santo islands waters and around Vanuatu in general 241 242 according to the absence of industrial and mining activities in this area (Harrison et al., 1996). 243 Surprisingly, in the present study, very little difference appeared in the sixteen trace element concentrations between N. macromphalus from New Caledonia and N. pompilius from 244 Vanuatu. Interestingly, there was no significant difference between both *Nautilus* species for 245

Ag, Co, Mn, Ni, Pb, Se, V and Zn (Table 3) indicating that the geographical origin does not 246 247 seem to be a major factor contributing to interspecific variations of these trace element 248 concentrations. As nautilus lives on the outer shelf of barrier reefs and rarely penetrates into 249 lagoons (Saunders and Ward, 1987; Norman, 2000), our results strongly supported the 250 hypothesis that the New Caledonian lagoon traps the major amount of the trace elements 251 derived from the island (Ambatsian et al., 1997). To the best of our knowledge, there are no 252 published data in the open literature concerning trace elements in organisms from the oceanic 253 waters off New Caledonia, except for two marine mammal species (Bustamante et al., 2003). 254 These authors reported very low concentrations of Co, Cr and Ni in the marine mammal tissues which suggested that mining activity in New Caledonia does not seem to be a 255 256 significant source of contamination in the oceanic domain. Overall, the global geologic 257 contagion due to the mining activity in New Caledonia seems to be limited to the shoreline 258 waters and to the inner lagoon (Fernandez et al., 2006).

259 More interestingly, the concentrations of rare earth elements were significantly different 260 between the digestive gland and the pericardial appendages of N. macromphalus and N. 261 pompilius. F-transition elements of rare earths have similar physicochemical properties, and it 262 is therefore not surprising that they correlate to each other (Table 4). Our results suggested 263 that rare earth elements would reflect the characteristics of the surrounding environment. 264 Very little is known about the concentrations of Ce, La and Nd in cephalopods, which varied from 0.045 to 1.95  $\mu$ g.g<sup>-1</sup> dwt in the digestive gland of the squid *Stenoteuthis oualaniensis* 265 266 (Ichihashi et al., 2001). In fact, rare earth elements have chemistry and sorption/complexation 267 characteristics analogous to transuranic elements such as americium or plutonium (Choppin 268 Torres and Chopin, 1984; Kim et al., 1989; Buckau et al., 1992). In Coleoid cephalopods, 269 transuranic elements also accumulate in the digestive and excreting tissues, in particular in the branchial heart and their appendages (Guary et al., 1981; Guary and Fowler, 1982; 270

Bustamante et al., 2006). It is therefore not surprising that they accumulate at relatively highconcentrations in nautilus digestive gland and pericardial appendage.

273 Besides Ce, La and Nd, the concentrations of As, Cu and Fe were also higher in N. pompilius 274 than in N. macromphalus. It is difficult to ascertain the reasons of such a difference but this 275 might also be linked to geological features reflecting a potential enrichment of Vanuatu 276 waters by local environmental processes, such as volcanism or upwelling (Malinovsky and 277 Markevich, 2007). In fact, only two trace elements, Cd and Cr, displayed significantly higher 278 concentrations in N. macromphalus than in N. pompilius tissues (Table 2). Therefore, the 279 reasons of such interspecific differences in bioaccumulation, i.e. higher concentrations of 280 trace elements in one species, remain unclear whereas an absence of difference could be 281 explained by the similar physiology of these two species (Ward, 1988; Norman, 2000).

282 As for other marine organisms, in cephalopods, trace element concentrations may vary with 283 biological and environmental factors such as temperature, salinity, season, age (size), sex, 284 lifestyle and geographical origin (e.g. Monteiro et al., 1992; Bustamante et al., 1998; 285 Raimundo et al., 2004; Seixas et al., 2005b; Pierce et al., 2008). Further investigations are 286 needed to understand how exactly these factors affect their concentrations in the studied 287 species. However, among these factors, individual size/age is known to be of primary 288 importance in invertebrates (e.g. Warnau et al., 1995; Bustamante and Miramand, 2005; 289 Hédouin et al., 2007). In the present study, the size as well as the two others main features 290 measured (sex and weight) were globally homogenous among the eight studied specimens. 291 Because nautiluses are long lived animals, it would be striking to determine the influence of 292 the size/age on trace elements bioaccumulation.

Sources of trace elements to nautilus can be (1) seawater, as it passes through the skin and through the gills during respiration, and (2) food, which is now considered as a major route for metal accumulation in marine animals, including both invertebrates and fish (Koyama et

296 al., 2000; Wang, 2002; Bustamante et al., 2002a, 2004). Knowledge of trace element speciation in the direct environment of nautilus is a key factor to ascertain the relative 297 298 importance of each uptake pathways. Concerning the first pathway (i.e. seawater), further 299 aquatic toxicity studies are needed to determine the fraction of total metal dissolved in 300 seawater (the 'toxic fraction') that react with, and is transported across biological membranes 301 such as nautilus gills. Concerning the second pathway (i.e. food), our comparative analysis 302 suggested that the metal discharges due to intense mining activities conducted on land in New 303 Caledonia are not easily transferred in the food web of nautilus. Therefore, to assess the 304 impact of mining activities on cephalopods in the region of New Caledonia, further studies should be conducted on more coastal species known to be resident in the impacted areas, such 305 306 as Sepiidae, Sepiolidae or Octopodidae spending all of their life in the waters of the New 307 Caledonian lagoon.

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310

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318

#### 319 **6. REFERENCES**

321	Ambatsian, P., Fernex, F., Bernat, M., Parron, C., Lecolle, J. 1997. High metal inputs to
322	closed seas: the New Caledonian lagoon. J. Geochem. Explor. 59, 59-74.
323	

Boucaud-Camou, E., Boucher-Rodoni, R. 1983. Feeding and digestion in Cephalopods. In:
The Mollusca, Part 2, physiology (Wilbur, K M ed). Academic press, New York, 1983, pp.
149-180.

327

Boucher-Rodoni, R., Boucaud-Camou, E. 1987. Fine structure and absorption of ferritin in the
digestive organs of *Loligo vulgaris* and *L. forbesi* (Cephalopoda, Teuthoidea). J. Morph. 193,
173-184.

331

Boyle, P., Rodhouse, P. 2005. Cephalopods: Ecology and Fisheries Blackwell publishing,Ames. 452 pp.

334

Buckeau, G., Kim, J.I., Klenze, R., Rhee, D.S., Wimmer, H. 1992. A comparative study of the
fulvate complexation of trivalent transuranic ions. Radiochim. Acta 57, 105-112.

337

Bustamante, P., Cherel, Y., Caurant, F., Miramand, P. 1998. Cadmium, copper and zinc in
octopuses from Kerguelen Islands, southern Indian Ocean. Polar Biol. 19, 264-271.

340

Bustamante, P., Grigioni, S., Boucher-Rodoni, R., Caurant, F., Miramand, P. 2000.
Bioaccumulation of 12 trace elements in the tissues of the nautilus *Nautilus macromphalus*from New Caledonia. Mar. Pollut. Bull. 40, 688-696.

345	Bustamante, P., Cosson, R.P., Gallien, I., Caurant, F., Miramand, P. 2002a. Cadmium
346	detoxification processes in the digestive gland of cephalopods in relation to accumulated
347	cadmium concentrations. Mar. Environ. Res. 53, 227-241.

349 Bustamante, P., Teyssié, J-L., Fowler, S.W., Cotret, O., Danis, B., Miramand, P., Warnau, M.

350 2002b. Biokinetics of zinc and cadmium accumulation and depuration at different stages in

the life cycle of the cuttlefish *Sepia officinalis*. Mar. Ecol. Prog. Ser. 231, 167–177.

352

Bustamante, P., Garrigue, C., Breau, L., Caurant, F., Dabin, W., Greaves, J., Dodemont, R.
2003. Trace elements in two odontocetes species (*Kogia breviceps* and *Globicephala macrorhynchus*) stranded in New Caledonia (South Pacific). Environ. Pollut. 124, 263-271.

356

Bustamante, P., Teyssié, J-L., Danis, B., Fowler, S.W., Miramand, P., Cotret, O., Miramand,
P., Warnau, M. 2004. Uptake, transfer and distribution of silver and cobalt in tissues of the
common cuttlefish *Sepia officinalis* at different stages of its life cycle. Mar. Ecol. Prog. Ser.
269, 185-195.

361

Bustamante, P., Miramand, P. 2005. Evaluation of the variegated scallop *Chlamys varia* as a
biomonitor of temporal trends of Cd, Cu, and Zn in the field. Environ. Pollut. 138, 109-120.

Bustamante, P., Teyssié, J-L., Fowler, S.W., Warnau, M. 2006. Assessment of the exposure
pathway in the uptake and distribution of americium and cesium in cuttlefish (*Sepia officinalis*) at different stages of its life cycle. J. Exp. Mar. Biol. Ecol. 331, 198-207.

368

Choppin Torres, R.A., Choppin, G.R. 1984. Europium (III) and americium (III) stability
constants with humic acid. Radiochim. Acta 34, 143-148.

5/1	
372	Dalvi, A.D., Bacon, W.G., Osborne, R.C. 2004. The past and the future of nickel laterites.
373	PDAC International Convention, Trade Show and Investors Exchange. March 7–10, 27 pp.
374	
375	Dorneles, P.R., Lailson-Brito, J., Aguiar dos Santos, R., Silva da Costa, P.A., Malm, O.,
376	Freitas Azevedo, A., Machado Torres, J.P. 2007. Cephalopods and cetaceans as indicators of
377	offshore bioavailability of cadmium off Central South Brazil Bight. Environ. Pollut. 148, 352-
378	359.
379	
380	Eisler, R. 1981. Trace metals concentration in marine organisms. Pergamon Press, New York,
381	685 pp.
382	
383	Fernandez, J.M., Ouillon, S., Chevillon, C., Douillet, P., Fichez, R., Legendre, R. 2006. A
384	combined modelling and geochemical study of the fate of terrigenous inputs from mixed
385	natural and mining sources in a coral reef lagoon (New Caledonia) Mar. Pollut. Bull. 52, 320-
386	331.
387	
388	Finger, J.M., Smith, J.D. 1987. Molecular association of Cu, Zn, Cd and <sup>210</sup> Po in the digestive
389	gland of the squid Nototodarus gouldi. Mar. Biol. 95, 87-91.
390	
391	Ghiretti-Magaldi, A., Giuditta, A., Ghiretti, F. 1958. Pathways of terminal respiration in
392	marine invertebrates. I. The respiratory system in cephalopods. J. Cell. Comp. Physiol. 52,
393	389-429.

- Guary, J-C., Higgo, J.J.W., Cherry, R.D., Heyraud, M. 1981. High concentrations of
  transuranic and natural radioactive elements in the branchial hearts of the cephalopods *Octopus vulgaris*. Mar. Ecol. Prog. Ser. 4, 123-126.
- 398
- Guary, J-C., Fowler, S.W. 1982. Experimental studies on the biokinetics of plutonium and
  americium in the cephalopod *Octopus vulgaris*. Mar. Ecol. Prog. Ser. 7, 327-335.
- 401
- 402 Harrison, N., Gangaiya, P., Morrison, R.J. 1996. Organochlorines in the Coastal Marine
  403 Environment of Vanuatu and Tonga. Mar. Pollut. Bull. 32, 575-579.
- 404
- 405 Hédouin, L., Metian, M., Teyssié, J-L., Fowler, S.W., Fichez, R., Warnau, M. 2006.
- 406 Allometric relationships in the bioconcentration of heavy metals by the edible tropical clam
  407 *Gafrarium tumidum*. Sci. Total Environ. 366, 154–163.
- 408
- Hédouin, L., Pringault, O., Metian, M., Bustamante, P., Warnau, M. 2007. Nickel
  bioaccumulation in bivalves from the New Caledonia lagoon: Seawater and food exposure.
  Chemosphere 66, 1449-1457.
- 412
- Hirano, S., Suzuki, K.T. 1996. Exposure, metabolism, and toxicity of rare earth and related
  compounds. Environ. Health Perspect. 104, 85-95.
- 415
- Kim, J.I., Buckeau, G., Bryant, E., Klenze, R. 1989. Complexation of americium (III) with
  humic acid. Radiochim. Acta 48, 135-143.
- 418
- Koyama, J., Nanamori, N., Segawa, S. 2000. Bioaccumulation of waterborne and dietary
  cadmium by oval squid *Sepioteuthis lessoniana*, and its distribution among organs. Mar.
  Pollut. Bull. 40, 961-996.
- 422

- 423 Ichihashi, H., Kohno, H., Kannan, K., Tsumura, A., Yamasaki, S. 2001. Multielemental
  424 analysis of purpleback flying squid using high resolution inductively coupled plasma-mass
  425 spectrometry (HR ICP-MS). Environ. Sci. Technol. 35, 3103-3108.
- 426
- Labrosse, P., Letourneur, Y., Kulbicki, M., Paddon, J.R. 2000. Fish stock assessment of the
  Northern New Caledonian lagoons: fishing pressure, potential yields and impact on
  management options. Aquat. Living Res. 13, 91-98.
- 430
- 431 Norman, M.D. 2000. *Cephalopods: a world guide*. IKAN publishing, Frankfurt, 320 pp.
- 432
- 433 Malinovsky, A.I., Markevich, P.V. 2007. Heavy Clastic Minerals from the Far East Island434 Arc Complexes. Russian J. Pacific Geol. 1, 71–81.
- 435
- 436 Martin, J.H., Flegal, A.R. 1975. High copper concentrations in squid livers in association with
  437 elevated levels of silver, cadmium, and zinc. Mar. Biol. 30, 51-55.
- 438
- 439 Metian, M., Bustamante, P., Hédouin, L., Warnau, M. 2008. Accumulation of trace elements
- 440 in the tropical scallop *Comptopallium radula* from coral reefs in New Caledonia. Environ.
- 441 Pollut. 152: 543-552.
- 442
- Miramand, P., Guary, J-C. 1980. High concentrations of some heavy metals in tissues of the
  Mediterranean octopus. Bull. Environ. Contam. Toxicol. 24, 783-788.
- 445

- 446 Miramand, P., Bentley, D. 1992. Concentration and distribution of heavy metals in tissues of
  447 two cephalopods, *Eledone cirrhosa* and *Sepia officinalis*, from the French coast of the English
  448 Channel. Mar. Biol. 114, 407-414.
- 449
- 450 Monniot, F., Martoja, R., Monniot, C. 1994. Cellular sites of iron and nickel accumulation in
- 451 ascidians related to the naturally and anthropic enriched New Caledonian environment.
- 452 Annal. Inst. Océanogr. 70, 205-216.
- 453
- Monteiro, L.R., Porteiro, F.M., Gonçalves, J.M. 1992. Inter- and intra-specific variation of
  mercury levels in muscle of cephalopods from the Azores. Arquipelago 10, 13-22.
- 456
- 457 O'Dor, R.K., Forsythe, J., Webber, D.M., Wells, J., Wells, M.J. 1993. Activity levels of458 Nautilus in the wild. Nature 362, 626-627.
- 459
- 460 Pernice, M., Wetzel, S., Gros, O., Boucher-Rodoni, R., Dubilier, N. 2007. Enigmatic dual
  461 symbiosis in the excretory organs of *Nautilus macromphalus* (Cephalopoda: Nautiloidea).
  462 Proc. Royal Soc. Lond. Ser. Biol. Sci. 274, 1143–1152.
- 463
- 464 Pernice, M., Boucher, J., Joannot, P., Boucher-Rodoni, R. 2006. Comparative study of
  465 bacteria diversity and bioaccumulation of trace elements in the excretory organs of Nautilus.
  466 In: CIAC symposium 2006, February 6-10, Hobart, Australia, 120 pp.
- 467
- 468 Pierce, G.J., Stowasser, G., Hastie, L.C., Bustamante, P. 2008. Geographic, seasonal and469 ontogenetic variation in cadmium and mercury concentrations in squid (Cephalopoda:

- 470 Teuthoidea) from UK waters. Ecotox. Environ. Safety in press
  471 (doi:10.1016/j.ecoenv.2007.07.007).
- 472
- 473 Rainbow, P.S., Wang, W-X. 2001. Comparative assimilation of Cr, Cr, Se, and Zn by the
  474 barnacle *Elminius modestus* from phytoplankton and zooplankton diets. Mar. Ecol. Prog. Ser.
  475 218, 239–248.
- 476
- 477 Rainbow, P.S. 2002. Trace metal concentrations in aquatic invertebrates: why and so what?
  478 Environ. Pollut. 120, 497–507.
- 479
- Raimundo, J., Caetano, M., Vale, C. 2004. Geographical variation and partition of metals in
  tissues of *Octopus vulgaris* along the Portuguese coast. Sci. Total Environ. 325, 71-81.
- 482
- 483 Saunders, W., Ward, P.D. 1987. Ecology, distribution and population characteristics of
  484 Nautilus. In: Nautilus, the biology and paleobiology of a living fossil Saunders, WB &
  485 Landman, NH. (eds). New-York & London, Plenum press, 137-162 pp.
- 486
- 487 Schipp, R., Martin, A.W., Liebermann, H., Magnier, Y. 1985. Cytomorphology and function
  488 of pericardial appendages of Nautilus (Cephalopoda, Tetrabranchiata). Zoomorph. 105, 16-29.
  489
- Schipp, R., Martin, A.W. 1987. The excretory system of Nautilus. In: *Nautilus, the biology and Paleobiology of a living fossil* (WB Sauders and NH. Landman, eds). Plenum Press, New
  York, 281-304 pp.
- 493

- 494 Seixas, S., Bustamante, P., Pierce, G.J. 2005a. Interannual patterns of variation in
  495 concentrations of trace elements in arms of *Octopus vulgaris*. Chemosphere 59, 1113-1124.
  496
- 497 Seixas, S., Bustamante, P., Pierce, G.J. 2005b. Accumulation of mercury in the tissues of the
  498 common octopus *Octopus vulgaris* (L.) in two localities on the Portuguese coast. Sci. Total
  499 Environ. 340, 113-122.
- 500
- Smith, J.D., Plues, L., Heyraud, M., Cherry, R.D. 1984. Concentrations of the elements Ag,
  Al, Ca, Cd, Cu, Fe, Mg,, Pb and Zn, and the radionuclides <sup>210</sup>Pb and <sup>210</sup>Po in the digestive
  gland of the squid *Nototodarus gouldi*. Mar. Environ. Res. 13, 55-68.
- 504
- Wang, W-X. 2002. Cd and Se aqueous uptake and exposure of green mussels *Perna viridis*:
  influences of seston quantity. Mar. Ecol. Prog. Ser. 226, 211–221.
- 507
- 508 Ward, P.D. 1988. In search of Nautilus "A New York Academy of Sciences book", Simon &509 Schuster eds, New-York.
- 510
- Warnau, M., Ledent, G., Temara, A. 1995. Allometry of heavy metal bioconcentration in the
  echinoid *Paracentrotus lividus*. Arch. Environ. Contam. Toxicol. 29, 393–399.
- 513
- Westermann, B., Ruth, P., Litzbauer, H.D., Beck, I., Beuerlin, K., Schimdtberg, H., Kleta,
  E.F., Schipp, R. 2002. The digestive tract of *Nautilus pompilius* (Cephalopoda,
  Tetrabranchiata): an X-ray analytical and computational tomography study on the living
  animal. J. Exp. Biol. 205, 1617-1624.
- 518

- 519 Worms, I., Simona, D.F., Hasslera, C.S., Wilkinson, K.J. 2006. Bioavailability of trace metals
  520 to aquatic microorganisms: importance of chemical, biological and physical processes on
  521 biouptake. Biochimie 88, 1721–1731.
- 522
- 523 Yamada, H., Takayanagi, K., Tateishi, M., Tagata, H., Ikeda, K. 1997. Organotin compounds
- and polychlorinated biphenyls of livers in squid collected from coastal waters and open ocean.
- 525 Environ. Pollut. 96(2), 217-226.

Species	Sex	Length (mm)	Weight (g)	
N. pompilius				
1	Male	177	825	
2	Male	151	660	
3	Male	171	796	
N. macromphalus				
1	Male	164	766	
2	Female	162	676	
3	Male	173	870	
4	Male	160	758	
5	Male	140	480	

Table 1. Characteristics of the sampled specimens of *Nautilus pompilius* and *N*. *macromphalus*.

Table 2: Trace elements concentrations (Mean  $\pm$  SE;  $\mu$ g g<sup>-1</sup> dry wt) in the tissues of Nautiloidae from this study and from the literature. The highest values are in bold. dl: detection limit; NA: not analysed; # only one value.

531

	Rare earth elements		Non essential metals			Essential metals and metalloids										
Tissue	La	Ce	Nd	Ag	Cd	Pb	As	Co	Cr	Cu	Fe	Mn	Ni	Se	V	Zn
Nautilus macromphalus <u>(</u> Busta	amante et al. 2	2000)														
Digestive gland	NA	NA	NA	4.45±2.68	45.1±13.2	0.46±0.17	166±26	7.8±3.6	4.4±1.1	106±46	666±231	10.1±1.7	16.3±7.8	NA	8.8±2.0	672±208
Pericardial appendages	NA	NA	NA	20.3±6.2	2.16±1.63	1.00±0.82	260±111	1.31±0.80	8.25±1.55	27.2±33.4	78±46	25.1±23.0	5.64±3.54	NA	0.18±0.09	22.4±26.6
Renal appendages	NA	NA	NA	1.80±2.22	1.87±1.23	1.06±0.88	112±65	0.43±0.37	2.54±2.88	7.1±10.2	343±160	77.1±54.1	22.7±13.9	NA	1.02±0.72	11.0±16.8
N. macromphalus (Present stu	udy)															
Digestive gland	0.30±0.07	0.25±0.11	0.19±0.07	9.42±3.91	51.5±3.5	0.98±0.15	257±19	4.69±1.51	2.95±0.35	263±78	353±63	10.9±1.3	$14.0{\pm}1.04$	2.65±0.90	14.8±2.28	388±61
Pericardial appendages	0.11±0.02	0.17±0.05	$0.15 \pm 0.05$	1.82±0.54	1.93±0.76	$1.57 \pm 0.11$	169±19	0.65±0.09	9.59±3.35	73.7±9.9	19±1	12.7±2.9	6.93±1.37	3.72 #	5.62±2.26	56.5±1.8
Renal appendages	<dl< td=""><td><dl< td=""><td><dl< td=""><td>0.44±0.12</td><td>3.39±1.16</td><td>2.11±0.51</td><td>152±17</td><td>0.18±0.03</td><td>2.57±0.36</td><td>27.6±4.2</td><td>20±3</td><td>68.2±6.2</td><td>19.7±2.39</td><td>9.84±0.94</td><td>1.54±0.13</td><td>35.2±2.8</td></dl<></td></dl<></td></dl<>	<dl< td=""><td><dl< td=""><td>0.44±0.12</td><td>3.39±1.16</td><td>2.11±0.51</td><td>152±17</td><td>0.18±0.03</td><td>2.57±0.36</td><td>27.6±4.2</td><td>20±3</td><td>68.2±6.2</td><td>19.7±2.39</td><td>9.84±0.94</td><td>1.54±0.13</td><td>35.2±2.8</td></dl<></td></dl<>	<dl< td=""><td>0.44±0.12</td><td>3.39±1.16</td><td>2.11±0.51</td><td>152±17</td><td>0.18±0.03</td><td>2.57±0.36</td><td>27.6±4.2</td><td>20±3</td><td>68.2±6.2</td><td>19.7±2.39</td><td>9.84±0.94</td><td>1.54±0.13</td><td>35.2±2.8</td></dl<>	0.44±0.12	3.39±1.16	2.11±0.51	152±17	0.18±0.03	2.57±0.36	27.6±4.2	20±3	68.2±6.2	19.7±2.39	9.84±0.94	1.54±0.13	35.2±2.8
N. pompilius (Present study)																
Digestive gland	1.00±0.14	1.58±0.32	0.92±0.10	5.65±1.04	74.3±6.8	$0.62 \pm 0.07$	210±16	3.85±1.04	1.37±0.24	311±61	625±51	15.4±1.7	9.77±0.07	2.35±0.84	8.42±0.57	470±52
Pericardial appendages	0.24±0.03	0.42±0.05	0.31±0.01	1.75±0.07	2.39±0.37	1.24±0.24	323±53	0.79±0.07	9.30±3.87	99.7±2.4	21±3	24.1±4.9	7.02±0.79	2.49 #	4.94±0.74	58.2±2.7
Renal appendages	<dl< td=""><td><dl< td=""><td><dl< td=""><td>0.35±0.03</td><td>5.93±0.31</td><td>0.99±0.21</td><td>292±80</td><td>0.18±0.03</td><td>1.98±0.08</td><td>53.3±4.6</td><td>24±3</td><td>307±83</td><td>19.6±0.97</td><td>13.3±6.12</td><td>1.82±0.02</td><td>42.7±1.5</td></dl<></td></dl<></td></dl<>	<dl< td=""><td><dl< td=""><td>0.35±0.03</td><td>5.93±0.31</td><td>0.99±0.21</td><td>292±80</td><td>0.18±0.03</td><td>1.98±0.08</td><td>53.3±4.6</td><td>24±3</td><td>307±83</td><td>19.6±0.97</td><td>13.3±6.12</td><td>1.82±0.02</td><td>42.7±1.5</td></dl<></td></dl<>	<dl< td=""><td>0.35±0.03</td><td>5.93±0.31</td><td>0.99±0.21</td><td>292±80</td><td>0.18±0.03</td><td>1.98±0.08</td><td>53.3±4.6</td><td>24±3</td><td>307±83</td><td>19.6±0.97</td><td>13.3±6.12</td><td>1.82±0.02</td><td>42.7±1.5</td></dl<>	0.35±0.03	5.93±0.31	0.99±0.21	292±80	0.18±0.03	1.98±0.08	53.3±4.6	24±3	307±83	19.6±0.97	13.3±6.12	1.82±0.02	42.7±1.5

## 532 Table 3. One-way ANOVA comparison of trace element concentrations in *Nautilus pompilius*

533 (Np) and in *N. macromphalus* (Nm). NS: not significant.

Element	Digestive gland	Pericardial appendages	Renal appendages		
Rare earth elements					
Ce	Np > Nm (F= 19.58 ; p= 0.007)	NS	-		
La	Np > Nm (F= 23.40; p= 0.005)	Np > Nm (F= 0.609; p= 0.025)	-		
Nd	Np > Nm (F= 25.13 ; p= 0.004)	Np > Nm (F= 6.54 ; p= 0.051)	-		
Essential elements					
As	NS	Np > Nm (F= 9.58 ; p= 0.027)	NS		
Co	NS	NS	NS		
Cr	Nm > Np (F= 11.34 ; p= 0.020)	NS	NS		
Cu	NS	NS	Np > Nm (F= 15.58; p=0.008)		
Fe	Np > Nm (F= 10.09 ; p= 0.025)	NS	NS		
Mn	NS	NS	NS		
Ni	NS	NS	NS		
Se	NS	NS	NS		
V	NS	NS	NS		
Zn	NS	NS	NS		
Non essential elements					
Ag	NS	NS	NS		
Cd	Nm > Np (F= 0.740 ; p= 0.023)	NS	NS		
Pb	NS	NS	NS		

- 535 Table 4: Correlations between trace elements within the different tissues of nautilus.
- 536 Not underlined: p<0.05; underlined: p<0.001.
- 537

Elements	Digestive gland	Renal appendages	Pericardial appendages
La	<u>+ Ce, + Nd</u> , - As, <u>+ Fe, + Mn</u>	<dl< td=""><td><u>+ Ce, + Nd</u>, + Mn</td></dl<>	<u>+ Ce, + Nd</u> , + Mn
Ce	+ Nd, $-$ As, $+$ Fe, $+$ Mn,	<dl< td=""><td>+ Nd, <math>+</math> Co, <math>+</math> La</td></dl<>	+ Nd, $+$ Co, $+$ La
Nd	<u>+ Fe, + Mn</u>	<dl< td=""><td><u>+ Ce</u>, +Co, <u>+ La</u></td></dl<>	<u>+ Ce</u> , +Co, <u>+ La</u>
Cu	+ Zn, -Se	-	-
Fe	+ Mn, -V, $+$ <u>La</u> , $+$ <u>Ce</u> , $+$ <u>Nd</u> $+$	- Zn	+ Ag, <u>+ Mn</u> , + Cr
Pb	+ V -	V, <u>+ Cr</u>	-
Mn	+ La, $+$ Ce, $+$ Nd, $+$ Fe $+$	- Cd, +Zn	+ La, <u>+ Fe</u>
Cd	- <u>+</u>	- <u>Mn</u>	+ Ni, $+$ V, $+$ Cr

539 <dl: concentrations below the detection limit.

### **<u>Caption to figures</u>**

542 Figure 1: Location of *Nautilus pompilius* and *N. macromphalus* specimens' collection.

Figure 2: Relative distribution of the trace elements between the digestive gland (DG), pericardial appendages (PA) and renal appendages (RA) of Nautilus pompilius from Vanuatu (black) and N. macromphalus from New Caledonia (white). Scale bars represent 1 standard error.



Figure 1.

