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**Carbon dynamics in a  
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# Aggregate structure and stability linked to carbon dynamics in a south Chilean Andisol

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## Abstract

The extreme vulnerability of soil organic carbon to climate and land use change emphasizes the need for further research in different terrestrial ecosystems. We have studied the aggregate stability and carbon dynamics in a chronosequence of three different land uses in a south Chilean Andisols: a second growth *Nothofagus obliqua* forest (SGFOR), a grassland (GRASS) and a *Pinus radiata* plantation (PINUS). The aim of this study was to investigate the role of Al as soil organic matter stabilizing agent in this Andisol. In a case study, we linked differences in carbon dynamics between the three land use treatments to physical protection and recalcitrance of the soil organic matter (SOM). In this study, C aggregate stability and dynamics were studied using size and density fractionation experiments of the SOM,  $\delta^{13}\text{C}$  and total carbon analysis of the different SOM fractions, and mineralization measurements. The results showed that electrostatic attractions between and among Al-oxides and clay minerals are mainly responsible for the stabilization of soil aggregates and the physical protection of the enclosed soil organic carbon. Whole soil C mineralization rate constants were highest for SGFOR and PINUS, followed by GRASS. In contrast, incubation experiments of isolated macro organic matter fractions showed that the recalcitrance of the SOM decreased in another order: PINUS > SGFOR > GRASS. We concluded that physical protection of soil aggregates was the main process determining whole soil C mineralization. Land use changes affected soil organic carbon dynamics in this south Chilean Andisol by altering soil pH and consequently available Al.

## 1. Introduction

Andisols develop from volcanic ash and are characterised by a high soil organic matter (SOM) content, good physical properties, and a high anion- and cation exchange capacity (AEC and CEC) (Galindo and Bingham, 1977). Their clay fraction consists mainly of poorly crystalline allophane, imogolite, and oxides and oxyhydroxides of Fe

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and Al (Besoain, 1958). Allophane is a group of clay minerals made up of silica, aluminum, and water in a chemical combination (Parfitt, 1990). In southern Chile (35°–49° S lat), Andisols cover a large part of the surface area of the Central Valley and Andes mountain ranges (FAO-UNESCO, 1971).

5 General overviews of changes in C-stocks after land use conversions are given by Paul et al. (2002) and Guo and Gifford (2002). De Koning et al. (2003) concluded more specific that in Al-rich soils, changes in C-stocks follow a pattern according to land use history and accompanying changes in soil characteristics. Temperate native forests covered most of southern Chile for the last 3000 years, remaining almost intact until  
10 the arrival of the Europeans, 450 years ago (Lara et al., 1999). Significant land use changes have taken place in this region (Lara et al., 1999). Today, only 56% (or 10.3 million ha) of this original forest cover remains. Most affected are the *Sclerophyllus* and the *Nothofagus* forest types, with 3 and 30% of the original forest cover remaining, respectively (Lara et al., 1999). On the other hand, grasslands and shrublands have  
15 doubled in area, from 2.5 to 5.5 million ha (Lara et al., 1999). New land use types, such as urban areas (55 000 ha), arable lands (2.8 million ha) and forestry plantations (2.1 million ha), now occupy a significant area as well.

Soil organic matter (SOM) has a very complex and heterogeneous composition and is generally mixed or associated with the mineral soil constituents to form soil aggregates (Del Galdo et al., 2003). Soil organic matter dynamics are mainly influenced by its recalcitrance and accessibility, and interactions between SOM and soil components (Sollins et al., 1996). Recalcitrance is the inherent molecular level resistance of a molecule to microbial and enzymatic breakdown. Accessibility is the extent to which the location of substrates controls access by microbes, fungi or enzymes. Interactions  
20 of organic substrates with organic or inorganic molecules may alter the degradation rate of the substrates (Swanston et al, 2002). Several studies have shown that in aluminum rich soils, such as Andisols, chemical stabilisation is the determining process for carbon accrual (Zunino et al., 1982; Percival et al., 2000). However, the exact role of SOM and Al in the process of chemical stabilization, and which kind of interactions  
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occurs between the different soil components is not yet fully understood.

The objective of this study was to evaluate the role of Al on carbon dynamics in Chilean Andisols and to investigate C stabilization by electrostatic Al-C binding, and occlusion within soil aggregates. As a case study, we tried to assess the effect of changes in certain physical and chemical soil characteristics – which resulted from the land use conversions – on SOC, physical protection of SOC in aggregates, quality of SOC, and organo-mineral associations. A chronosequence of three different land uses were studied: a second growth *Nothofagus obliqua* forest (SGFOR), a grassland (GRASS), and a plantation of *Pinus radiata* (PINUS).

## 2. Materials and methods

### 2.1. Site description and soil sampling

Soil samples were taken in August 2002 near Paillaco, in the Central Valley of southern Chile (40°07 S, 72°51 W). The mean annual precipitation varies from 1600–1800 mm, while mean annual temperature is 10.8°C. All soils were classified as *trumao's* or Andisols, Typic Dysandrepts (Tosso, 1985). They have typical variable electrical charge characteristics (i.e. pH dependent) as a result of their particular clay mineral and humus content (Radcliffe and Gillman, 1985; Nanzyo et al., 1993). A mixture of 1:1 and 2:1 clay minerals (e.g. halloysite, vermiculite, etc.) has been documented in Chilean Andisols as well as the generally observed poorly crystalline minerals (Besoin, 1958, 1969; Galindo and Bingham, 1977; Besoin et al., 2000).

The three sites are all located in close proximity (100 m apart) and were originally all under the same forest vegetation dominated by *Nothofagus obliqua*. Nowadays, only the SGFOR site is still under this original vegetation. The land use of the GRASS and PINUS sites changed 50–100 years ago from second growth *N. obliqua* forest to grassland for agricultural livestock activity. Finally, 16 years ago, a *Pinus radiata* plantation was established on a part of the GRASS site.

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The second growth *N. obliqua* forest (SGFOR) has a mean tree height of 35 m, a stand density of 757 ha<sup>-1</sup> and a canopy cover of 70% during the growing season. The average tree age is 120 years, while mean diameter at breast height equals 42.5 cm. In this forest, the overstorey is dominated by *N. obliqua* (32 m) with a cover of 40%.

5 The understorey (25 m) consists of *Aextoxicon punctatum*, *Persea linque*, *Nothofagus obliqua*, *Eucriphia cordifolia*, *Laurelia sempervirens*, *Nothofagus dombeyi* and *Gevuina avellana* with a cover of 70%. The lower understorey (10 m) is dominated by *Aextoxicon punctatum*. In the shrub stratum (<4 m) *Chusquea quila* and *Raphithamnus spinosus* are dominant.

10 Since the establishment of the pasture, agricultural activities (cattle breeding) have taken place on the GRASS and PINUS sites with an animal density of 1.5–2 cows ha<sup>-1</sup>. The grassland has a species composition consisting of *Lolium perenne* L. and *Trifolium repens* L., and is ploughed and resown every two years. A mixture of various forms of acidifying fertilizers has been applied yearly on the GRASS site at high levels, though  
15 no data were available on the exact fertilizer type or application rate.

On the PINUS site, fertilization was stopped with the establishment of the plantation, 16 years ago. This *P. radiata* monoculture has a mean tree height of 15–18 m, while stand density is around 1500 ha<sup>-1</sup>.

20 Sampling was done after selection of representative plots in the three ecosystem sites (land use treatments). In each land use treatment, three sampling plots were selected based on pedological, forestry and ecological experience. Samples originating from a different sampling plot in the same land use treatment were considered as replicates, and were kept separate for later analysis. In each sampling plot, one soil sample has been taken at 0–10, 10–20 and 20–30 cm depth. Thus, in total 27 soil  
25 samples were taken. After transport to the laboratory, the recognisable plant material was removed. All soil samples were dried, homogenised and sieved (2 mm) for later analyses. Soil samples were dried in open air to a gravimetric water content of approximately 20%. Volcanic-ash soils can granulate irreversibly when completely dried (Kubota, 1972), and this can affect the subsequent fractionation of SOM. We will refer

to these partly dried samples using the term “air dried samples”.

## 2.2. Size and density fractionation of SOM

The size fractionation procedure was performed by separately wet sieving a soil sample of 100 g over three sieves (250  $\mu\text{m}$ , 150  $\mu\text{m}$  and 50  $\mu\text{m}$ ) with tap water. The suspension passing the bottom size was collected and left about 48 h at room temperature to settle. After settling, the clear solution was removed and the size fraction <50  $\mu\text{m}$  was collected. Four size fractions (2000–250  $\mu\text{m}$ , 250–150  $\mu\text{m}$ , 150–50  $\mu\text{m}$  and <50  $\mu\text{m}$ ) were obtained, dried at 40°C for 48 h and homogenised using a planetary ball mill (PM400, Retsch, Germany) for subsequent analysis.

The size fractionation was performed at two different times, using the same procedure: (1) starting from “air dried” soil (slaking) and (2) after an 80 day laboratory incubation experiment (wet sieving of field moist soil) (see below). In the incubation experiment, the soil was exposed to several supplementary disruption processes such as rewetting and mixing of the soils. Using this procedure, the susceptibility of the soil aggregates against disruption processes could be evaluated. The distribution of the soil weight among the different size fractions was expressed using the weighted average diameter (WAD) and could thus be quantified as one single parameter for further statistical analyses. The difference in WAD between the two fractionation experiments can be related to the stability of the soil aggregates. A smaller difference in WAD indicates a higher stability of the soil aggregates.

Density fractionation of the soil organic matter was performed using the method of Meijboom et al. (1995) (Fig. 1). Air dried soil samples (400 g) were wet sieved over two sieves with tap water (top sieve mesh size 250  $\mu\text{m}$ , bottom sieve mesh size 150  $\mu\text{m}$ ), until the passing water became clear in order to destroy all macroaggregates >150  $\mu\text{m}$ . Next, the two size fractions retained on the sieves were washed into a bucket and swirled with a jet of water to separate the organic matter (OM) from the mineral fraction by decantation. Swirling and decanting were repeated until no more floating organic matter appeared. The obtained macro organic matter (MOM) from both size frac-

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tions (250–2000 and 150–250  $\mu\text{m}$ ) was recombined to be further separated into a light (LF, density  $<1.13 \text{ Mg m}^{-3}$ ), intermediate (IF,  $1.13 \text{ Mg m}^{-3} < \text{density} <1.37 \text{ Mg m}^{-3}$ ) and heavy (HF, density  $>1.37 \text{ Mg m}^{-3}$ ) fraction by subsequent submersion in two colloidal silica suspensions (Ludox, Dupont, USA) with a respective density of  $1.13 \text{ Mg m}^{-3}$  and  $1.37 \text{ Mg m}^{-3}$ . The obtained density fractions were washed with distilled water and dried at  $40^\circ\text{C}$  for 48 h.

The C concentrations increase with decreasing density of the fractions (LF > IF > LF). The IF and HF show a higher association of the SOM of these heavier fractions with soil minerals (Hassink, 1995). The LF consists mainly of recognisable plant material in an early stage of decomposition and serves as readily decomposable substrate for the microbial biomass (Six et al., 1999; Solomon et al., 2000).

### 2.3. Carbon mineralization experiments

The C mineralization experiments consisted out of two different experiments. In a first experiment, whole soil C mineralization was measured. In a second experiment, isolated macro organic matter fractions (LF and HF) were incubated to determine the specific C mineralization of these fractions.

Before starting the whole soil C mineralization experiment, the air dried soil was rewetted and pre-incubated during six days at a temperature of  $15^\circ\text{C}$ . Air dried bulk samples were brought to a gravimetric water content of 46% for SGFOR and PINUS, respectively, and 32% for GRASS, corresponding to a water filled pore space of 25% at the bulk density in the field (using a bulk density for all the layers of  $0.45 \text{ Mg m}^{-3}$  for SGFOR and PINUS, and  $0.60 \text{ Mg m}^{-3}$  for GRASS (see Table 1) and a soil particle density of  $2.65 \text{ Mg m}^{-3}$ ). As such, a realistic microbial soil activity could be restored. An exact quantity of soil was put in PVC tubes (4.6 cm diameter, 13 cm high) and compressed manually to the corresponding bulk density. The tubes were covered with parafilm, which permits sufficient gas exchange and aeration, but reduced moisture loss. During the entire experiment, the moisture content and temperature were kept constant. Carbon dioxide emission was measured on eight different time points (day 1,

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3, 7, 11, 17, 35, 43 and 62). This was done by putting the PVC-tubes into glass jars, which were closed airtight. The headspace of the glass jars was sampled after 0, 27, 54 and 81 min through a rubber septum. Before sampling, the CO<sub>2</sub> concentration in the headspace was homogenised by repeatedly sucking and re-injecting the headspace.

5 A gas sample of 1 mL was injected into a gas chromatograph (GC-14B, Shimadzu, Japan) (Porapak Q, mesh size 80/100 column specification, electron capture detector (ECD)). The CO<sub>2</sub> concentration was calculated via a linear calibration using a certified CO<sub>2</sub> concentration (1% CO<sub>2</sub> in dry air, Air Liquide, France). Linear regression between the measuring points at 0, 27, 54 and 81 min gave the CO<sub>2</sub> mineralization flux.

10 The incubation of the isolated HF and LF was only performed for SOM originating from the 0–10 cm soil layer of each land use treatment, following the method described by Swanston et al. (2002). For LF, 2.00 g substrate was mixed with 18.00 g inert quartz sand (acid washed, 580–650 μm diameter) in a glass flask. For HF, 5.00 g substrate was mixed with 15.00 g inert quartz sand. A microbial inoculum was added at the beginning of the incubation. A water extract from the mineral soil originating from the same soil layer was used as inoculum source. The inoculum was made by adding distilled water to a wet soil in a 1/1.5 ratio, followed by placing the mixture on a rotary shaker for 30 min. After settling down, 4 mL of the supernatant was added to each flask and the substrate was pre-incubated for two days. During the entire experiment, the inoculated flasks were stored in an incubation room at a constant temperature of 15°C. The CO<sub>2</sub> concentration was measured through a septum in the cap of the flasks after 0, 27, 54 and 81 min and on six different time points (day 1, 3, 7, 11, 17 and 35) with a gas chromatograph (GC-14B, Shimadzu, Japan), using the same method as described above.

#### 25 2.4. Physico-chemical analyses

Measurement of <sup>13</sup>C and total C (TC) content in the different soils and SOM fractions was performed using an elemental analyser (ANCA-SL, PDZ-Europa, UK) coupled to an Isotope Ratio Mass Spectrometer (20–20, PDZ Europa, UK). The measured

$^{13}\text{C}/^{12}\text{C}$  ratios are expressed as  $\delta^{13}\text{C}$  values (‰) vs. the international Vienna Pee Dee Belemnite (VPDB) standard:

$$\delta^{13}\text{C} = \frac{R_{\text{sample}} - R_{\text{standard}}}{R_{\text{standard}}} \cdot 1000(\text{‰}) \quad (1)$$

$R_{\text{sample}}$  and  $R_{\text{standard}}$  refer to the  $^{13}\text{C}/^{12}\text{C}$  ratio of the measured sample and the standard, respectively. The working standard was flour with a  $\delta^{13}\text{C}$  value of  $-27.01 \pm 0.04\text{‰}$  vs. VPDB and TC of  $39.95 \pm 0.40\text{‰}$  (certified by Iso-Analytical, UK).

A modified procedure of the method described by Schwertmann (1964), using a mixture of ammonium oxalate (0.2 M) and oxalic acid as extractant, was applied for the determination of the extractable aluminum oxalate ( $\text{Al}_{\text{ox}}$ ), and silicon oxalate ( $\text{Si}_{\text{ox}}$ ) content. In the absence of light, a soil:solution (1:10) mixture was placed on a rotary shaker for 2 h. A pyrophosphate reagent (0.1 M) was used in soil:solution ratio of 1:10 (shaking time 16 h) for the determination of the pyrophosphate extractable Al content ( $\text{Al}_{\text{py}}$ ). All supernatants were centrifuged at 3000 rpm for 10 min, filtrated (folded filters, Schleicher & Schüell, Germany) and the Al and Si content of the solutions were determined with, respectively, ICP-OES (Inductive Coupled Plasma – Optical Emission Spectrometry) (VISTA-MPX CCD Simultaneous ICP-OES, Varian, USA), and AAS. Allophane contents were calculated using the formula proposed by Parfitt and Wilson (1985), and later simplified by Mizota and van Reeuwijk (1989):

$$\text{Allophane} = 100 \text{Si}_{\text{ox}} \cdot \left[ 23.4 - 5.1 \left( \frac{\text{Al}_{\text{ox}} - \text{Al}_{\text{py}}}{\text{Si}_{\text{ox}}} \right) \right] (\%) \quad (2)$$

## 2.5. Statistical analyses

Statistical analyses were performed using SYSTAT version 10.0 (SPSS, USA). Statistical differences of means (95% significance level) were distinguished using ANOVA (3 replicates) followed by a Duncan's multiple range post-hoc test. Paired samples T-tests were used to determine differences in C mineralization rates between LF and HF

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fractions. Log transformation was used to improve normality and homoscedasticity in the data when necessary. In case no homogeneity of variance was reached after log transformation, Tamhane's T2 test was used as post-hoc test. Correlations were determined by the "Pearson's correlation factor".

### 3. Results

#### 3.1. Physico-chemical soil properties

The chemical and physical properties of the soils are listed in Table 1. Averaged over the three soil layers, land use changes significantly affected soil C stocks with +31 and -42% for the conversion from SGFOR to GRASS and GRASS to PINUS, respectively. The N content was highest in GRASS, followed by SGFOR and PINUS. Nitrogen concentrations varied from 0.53 to 0.18 kg m<sup>-2</sup>. Fertilizer additions on the GRASS site are responsible for the higher N contents in this site. The bulk density was higher in GRASS than in SGFOR and PINUS and is as well a result of agricultural livestock activities. Soil texture was categorized as sandy loam (USDA texture class) for all the land use treatments. The soils in SGFOR and PINUS showed an average pH<sub>H2O</sub> between 6.1 and 6.7. In contrast, GRASS showed a lower pH<sub>H2O</sub> (around 5.5). The extractable Al<sub>ox</sub> and Al<sub>py</sub> concentrations in the GRASS site were higher than for SGFOR and PINUS, though differences between GRASS and PINUS were not always significant for Al<sub>ox</sub>. Soil pH was negatively correlated with Al<sub>ox</sub> ( $r=-0.45$ ;  $p=0.02$ ), and Al<sub>py</sub> ( $r=-0.83$ ;  $p=0.00$ ). Allophane contents were for the three soil layers significantly lower for GRASS compared to SGFOR and PINUS.

#### 3.2. Size and density fractionation

The highest weight percentage after the first size fractionation was found in the size fraction 250–2000 μm (Fig. 2). The other size fractions contain smaller amounts of

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soil weight, varying from 23.1 to 7.0% for all land use treatments. The WAD was not significantly different between the different land use treatments, except for PINUS 20–30 cm where a significant lower value was found (Table 2). TC concentration (%) was not significantly different among the different size fractions in all the sites, with exception of the 0–10 cm layer of SGFOR (Table 3).

The results of the weight distribution of the second size fractionation experiment (after an 80 day incubation period, starting from field moist soil) are shown in Fig. 3. A considerable shift in soil weight towards the finer size fractions could be noticed, which results in a smaller WAD compared to the first fractionation procedure (Table 2). Between land use treatments, GRASS showed the coarsest aggregate distribution but due to relative large standard deviations (on average  $50 \mu\text{m}$ ), differences in WAD were only significant for the 20–30 cm layer. As in the first fractionation procedure, the soil carbon concentration did not differ significantly among the different size classes for all land uses, though a few exceptions were observed here (data not shown).

Averaged over the three soil layers, the aggregate stability decreased in the following order: GRASS > PINUS > SGFOR, however only significant differences were observed for the 20–30 cm layer between SGFOR and GRASS (Table 2). The difference in WAD was negatively correlated with the extractable  $\text{Al}_{\text{ox}}$  content of the soil ( $r=-0.50$ ;  $p=0.01$ ), but not with the TC content ( $r=-0.18$ ;  $p=0.38$ ), extractable  $\text{Al}_{\text{py}}$  content ( $r=-0.23$ ;  $p=0.25$ ) or allophane content ( $r=0.11$ ;  $p=0.60$ ) of the soil.

The amounts of MOM are expressed relative to the TC content (i.e. as % of total C) (Table 4). The differences between land use treatments diminished with depth and were only significant between SGFOR, GRASS and PINUS for the 0–10 cm and 10–20 cm layer. The quantities of MOM for the three soil layers, expressed as % of total C, were negatively correlated with the extractable  $\text{Al}_{\text{ox}}$  content of the soil ( $r=-0.74$ ,  $p<0.01$ ).

Table 5 shows the complete mass and carbon balance for the 0–10 cm. It was found that the C content of the MOM is only a small fraction of the total C content (2.4%–8.3%) in the soil. Though the concept of the fractionation procedure assumes that

macroaggregates ( $>150 \mu\text{m}$ ) are broken and the enclosed organic matter is released, still 56–74% of the TC content of the soil was found in the remaining mineral fraction (i.e. the fraction 150–2000  $\mu\text{m}$ , from which the MOM is isolated).

### 3.3. $\delta^{13}\text{C}$ signature in size and density fractions

5 The  $\delta^{13}\text{C}$  values of the whole soil C and the different SOM fractions from the three land use treatments and soil layers are presented in Table 6. Within the different size and density fractions of a certain land use, no significant differences in  $\delta^{13}\text{C}$  value were found. However, a significant difference of 1-2‰ was found between the  $\delta^{13}\text{C}$  value of the size classes and density classes for the different land use treatments, with  
10 exception for PINUS 0–10 cm.

### 3.4. Mineralization experiments

#### 3.4.1. Whole soil C mineralization

The cumulative C mineralization curves were calculated for all land use treatments as shown in Fig. 4 for SGFOR. The linear part of each curve (from day 17 to 62) was used  
15 to estimate the potential C mineralization rate via linear regression (Table 7). The differences in potential C mineralization rates between the different land use treatments were significant in the 0–10 cm soil layer: SGFOR > PINUS > GRASS. However, differences between the three land use treatments disappeared gradually with depth. In the 10–20 cm and 20–30 cm layer SGFOR still showed the highest potential mineralization rate, but differences were no longer significant. For all land use treatments, potential  
20 C mineralization rates decreased with soil depth. The C mineralization rate constant, which can be used as an indicator of the degradability of the SOM, was calculated by dividing the observed potential C mineralization rate (expressed per gram soil) by the corresponding TC content in each soil layer (Table 7). For the 0–10 cm soil layer, these  
25 values varied from 0.029 for SGFOR to 0.023 for PINUS and  $0.011 \text{ yr}^{-1}$  for GRASS. In

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all soil layers, the decomposability of the SOM was significantly lower for GRASS than for PINUS and SGFOR. The SOM from GRASS showed the highest stability against microbial breakdown.

### 3.4.2. Incubation of the isolated MOM density fractions HF and LF

In this procedure, the MOM was isolated by the density separation procedure, through which the physical protection of SOM in soil aggregates was removed (Fig. 1). Especially, the accessibility of the HF has been increased, as this fraction is more associated with soil minerals and components compared to the LF. The LF is mainly present in the soil as free MOM. Only a small fraction is bounded in soil aggregates (Hassink, 1995; Swanston et al., 2002). Potential C mineralization rates and C mineralization rate constants of the isolated density fractions were calculated using the same method as the whole soil C mineralization experiment (Table 8). There, the mineralization rate constants are an indicator of the recalcitrance of the macro organic matter fractions. Statistical analyses showed that the potential C mineralization rates of the LF and HF were significantly different for all land use treatments. The C mineralization rate constants of the LF and HF are significantly different for GRASS, but not for SGFOR and PINUS.

The C mineralization rate constants showed that the MOM from GRASS decomposes fastest. The recalcitrance of the LF decreased in the following order: PINUS > SGFOR > GRASS. For the HF, PINUS showed a higher recalcitrance than GRASS, whereas the value of SGFOR was not significantly different from PINUS and GRASS (Table 8).

## 4. Discussion

In a first section of the discussion, we will focus on the interaction of AI, SOM and soil minerals. AI-humus complexes, electrostatic-C binding and aggregate structure

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will be evaluated in this Andisol. In a second part, we will make a link between aggregate structure, physical and chemical soil properties and C mineralization experiments. Though land use changes certainly affect C dynamics by differences in C input, and direct effects of accompanying soil disturbances (Guo and Gifford, 2002), we will focus on differences in soil properties between land use treatments, which resulted of changes in land use and land management.

The pH differences between GRASS and both SGFOR and PINUS were significant for all soil layers, and most probably a result of the acidifying fertilizer applications. Under lower pH conditions, Al is more soluble and available for complexation with soil components (Wesselink et al., 1996). Consequently, the extractable  $Al_{ox}$  and  $Al_{py}$  concentrations in the GRASS site were higher than for SGFOR and PINUS. This might be a result of differences in soil pH between the sites, but this is not certain as the extraction procedure changes the soil pH. In the literature (Bertsch and Bloom, 1996), the extractable  $Al_{py}$  content is often related to organically bound Al, while the quantity of poorly ordered amorphous or noncrystalline Al phases plus the organically bound Al would be proportional to  $Al_{ox}$ . Though, as stated by Bertsch and Bloom (1996), the extractants are not completely selective for specific phases. Considerable overlap between the forms of Al solubilized by a given extractant exists. Pyrophosphate reagent, for instance, forms strong complexes with Al, that, coupled with the high pH (10), can also remove poorly ordered or non-crystalline Al phases associated with SOM (Bascomb, 1968; McKeague and Schluppi, 1982). This non-crystalline Al may be largely coassociated with organic matter that is released via dispersion of the organic material during the pyrophosphate extraction (McKeague and Schluppi, 1982).

The presence of poorly crystalline Fe- and Al-oxides in Andisols is known to have a stabilizing effect on aggregation upon drying (Schahabi and Schwertmann, 1970). Elliott (1986) and Beare et al. (1994) indicated the disrupting effect of physical disturbance such as dry-wet cycles and variations in moisture content on soil aggregates. This explains the decreased resistance of soil aggregates against applied dispersive forces in the second size fractionation procedure.

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The results of the size and density fractionation procedures results showed that the extractable  $Al_{ox}$  content in the soil is positively related to the stability of soil aggregates. Thus, if less extractable  $Al_{ox}$  is present in the soil, less stable soil aggregates can be formed and relatively more SOM will remain in the more labile MOM pool. Though that the GRASS site is ploughed every two years, the aggregates appear to be most stable in this site. The stronger binding forces involved in the soil aggregation process in GRASS overcome the disruptive effect of ploughing on soil aggregates (Beare et al., 1994; Six et al., 1999).

The MOM fractions that were isolated during the fractionation procedure had a significant lower  $\delta^{13}C$  value than the SOM that remained in the mineral fraction. It was indicated that the SOM in the mineral fraction has been more processed than the recent incorporated macro organic matter. The  $\delta^{13}C$  value of the OM namely increases as decomposition proceeds (O'Brien and Stout, 1978; Accoe et al., 2002). As the OM enters the soil, it is at first found in the more labile MOM pool. Later, after a certain time of processing, it is shifted towards the more stable fractions, where it is enclosed in stable aggregates. Our results are in contrast to Accoe et al. (2002) where a more gradual shift in  $\delta^{13}C$  value among the different density and size classes could be observed, indicating a gradual shift of the SOM towards, firstly, a heavier density and, secondly, finer size class as decomposition occurs. In this study, we could only distinguish two different SOM pools: the MOM and the mineral soil pool. The non-significant differences in the MOM fractions for a soil layer can be explained by the absence of major isotopic enrichments of bulk plant material during the early steps of decomposition in forest soils (Balasdent and Mariotti, 1996; Nadelhoffer and Fry, 1988).

#### 4.1. Aggregate structure in south Chilean Andisols

It is generally accepted that SOM is stabilised by Al in Andisols (e.g. Martin et al., 1982; Zunino et al., 1982). In the literature, two different mechanisms can be found for SOM stabilisation by Al. Firstly, organo-Al complexes would act as binding agents between the aggregates or soil particles, through which the enclosed SOM is protected against



microbial breakdown. In organo-Al complexes, SOM is sorbed by allophane through ligand exchange between the carboxyl functional groups of the SOM and the surface hydroxyl groups bonded to Al (Parfitt et al., 1977, 1999; Skjemstad, 1992). Since surface hydroxyls are characteristic for variable charge surfaces, sorption of negatively charged organics by ligand exchange should be most important in oxide-rich and allophonic soils, such as Andisols (Sollins et al., 1996). In this first case, the mechanism of aggregate hierarchy (Tisdall and Oades, 1982) would be adequate to describe carbon dynamics as SOM (bonded to Al) acts as principal binding agent between soil components.

Secondly, electrostatic attractions could occur between and among Al, Fe and their oxides and clay minerals due to the simultaneous existence of positive and negative charges at field pH (Schofield and Samson, 1954; El-Swaify, 1980; Six et al., 2000b, c; Deneff et al., 2002). Thus, here aggregate formation is partly induced by electrostatic interactions and aggregate hierarchy should be less pronounced (Oades and Waters, 1991).

We conceptually tested the occurrence of aggregate hierarchy for the different land use treatments. Three consequences of this theory are: (1) a gradual breakdown of macroaggregates into microaggregates before they dissolve into primary particles, as increasing dispersive energy is applied to the soil (Oades and Waters, 1991), (2) an increase in percentage C concentration with increasing aggregate-size class because large aggregate-size classes are composed of small aggregate-size classes plus organic binding agents (Elliott, 1986), and (3) younger and more labile organic matter is contained in macroaggregates rather than in microaggregates (Elliott, 1986; Puget et al., 1995; Jastrow et al., 1996). Firstly, the fractionation of the soils at two different periods allowed us to test the aggregate stability. A gradual breakdown of macroaggregates into microaggregates could be observed (Figs. 2, 3). It was shown that this breakdown could not be explained by the TC content, allophane content or extractable  $Al_{py}$  content of the soil, but by the extractable  $Al_{ox}$  content of the soil. In the Al-extraction procedure, only amorphous and poorly crystalline Al-oxides and Al

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bound to organic complexes could be isolated (Bertsch and Bloom, 1996). Secondly, the soil C concentration did not differ among the different size classes for a certain land use treatment (Table 3). Thirdly, there were no significant differences in  $\delta^{13}\text{C}$  values among the different size classes for a certain soil (Table 6). The latter indicates that the SOM in the different size fractions is at the same degree of decomposition.

The results of these experiments indicated that organo-Al complexes are probably not the main binding agents between soil aggregates in these Andisols. In small soil particles, organic compounds could be enclosed between soil particles or small aggregates because polyvalent-organic matter complexes form bridges between negative charged 2:1 clay platelets (Six et al., 2000c). However, a further increase in aggregate size would be mainly a result of electrostatic attractions between and among Al and Al-oxides and clay minerals, as documented as well by Schofield and Samson (1954), Violante and Huang (1992) and Deneff et al. (2002). Organic matter would only play a minor role in these strong bindings between larger aggregates. As such, large and stable aggregates (cf. mineral fraction) are formed and the enclosed organic matter would experience a high degree of physically protection in these Al-mineral-SOM complexes. These results are in accordance to Deneff et al. (2002), who investigated to role of Al and OM in oxide rich soils with a different clay mineralogy. These authors also suggested that the presence of non-crystalline oxides, vermiculites and 1:1 clay minerals in soils leads to very strong interactions between 2:1 and 1:1 clay minerals, oxides and organic matter compounds, and hence high levels of stable macroaggregation with high inputs of OM.

Allophane contents were significantly lower for GRASS than for SGFOR and PINUS. It was indicated that as a result of a lower pH, aluminum was released from allophanic compounds, and resulted in a higher Al content associated with the mineral-SOM complexes. Statistical evidence for this conclusion was found by a strong significant correlation between soil pH and allophane ( $r=0.74$ ,  $p=0.00$ ). Increased aluminum release from allophane at decreasing pH was also found by Farmer and Lumsdon (2002). Dahlgren and Saigusa (1994) found in Andisols evidence for the precipitation of re-

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leased Al from allophanic components on soil surfaces, while Walker et al. (1988) found more specifically that soluble Al was adsorbed extremely rapid on common soil minerals such as vermiculite. Violante and Huang (1992) found in OM rich soils that aggregation between soil particles was promoted by increasing Al precipitation. Tama and El-Swaify (1978) stated that soil pH affects the ability of soil particles to aggregate in another important way in variable charge soils. The electrostatic contribution of oxides to aggregate stability is pH dependent and affects the extent of aggregate breakdown at different pH values. In lowering soil pH, the amount of positive charges of the Al-oxides and allophane increases (Espinoza et al., 1975). As such, more electrostatic forces are involved in aggregating soil particles. This indicates that the soil pH could affect aggregate stability. However, in our statistic analyses of the aggregate stability test no significant correlation was found between soil pH and difference in WAD between the two fractionation procedures. Possibly, the sigmoid retention curves, relating the effect of soil pH and positive charges, documented in south Chilean Andisols (Espinoza et al., 1975) might be responsible for the absence of a linear correlation.

#### 4.2. C-mineralization experiments

For the 0–10 cm layer, the whole soil C mineralization rate constant decreased in the following order: SGFOR  $\approx$  PINUS > GRASS. Our values are in accordance to those found in Columbian Andisols by Boudot et al. (1986), who measured a mineralization rate constant of approximately  $0.01 \text{ yr}^{-1}$  in the 5–20 cm layer. For the incubation of the isolated MOM density fractions, the C mineralization rate constant of GRASS was highest. GRASS SOM showed thus the lowest recalcitrance against microbial breakdown (see further), but however, its SOM physical protection and macroaggregate turnover was highest.

In Andisols, it is known that metal-humus complexes are resistant to microbial decomposition (Juste et al., 1975), and offer as such physical protection against microbial breakdown. Gijsman and Sanz (1998) also investigated the physical protection of macroaggregates in Andisols. After crushing of macroaggregates, these authors ob-

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served a significant increase in CO<sub>2</sub> mineralization compared to intact soil samples. Plante and McGill (2002) indicated that macroaggregate turnover results in the exposure of labile organic matter. Jastrow (1996) found evidence for a relationship between macroaggregate formation and C accrual. Six et al. (2000a) underscored the importance of soil aggregation and more specifically the interactions of SOM and aggregate dynamics in controlling C-sequestration in soils.

In our study, the C mineralization rate constants (SGFOR  $\approx$  PINUS > GRASS) showed an opposite trend as the aggregate stability. It was indicated that aggregate stability affects the C mineralization process in Andisols. The latter was confirmed by the significant correlation between the whole soil C mineralization rate constants and the difference in WAD ( $r=0.39$ ,  $p=0.04$ ). The lower C mineralization rate constants in GRASS are probably a result of the stabilization of the SOM by aluminum in Andisols (see above). Further statistical evidence for this conclusion was found in a strong negative and significant correlation between the mineralization rate constant and Al<sub>ox</sub> content ( $r=-0.68$ ,  $p=0.00$ ). Percival et al. (2000) also indicated that the C stabilization by Al can lead to exceptionally slow soil C turnover.

## 5. Conclusions

It can be concluded that either by electrostatic bridges between Al-oxides and clay minerals, and dissolution of Al from allophanic compounds, physico-chemical stabilization of the SOM occurs. It is important to mention that Al compounds are mainly bound directly onto clay minerals via electrostatic attractions. However, as clay minerals are closely associated with organic matter compounds, stable Al-mineral-humus complexes can be formed. It was also indicated that this process of aggregate stabilization has an important effect on C mineralization.

In this study, land use conversion from a second growth forest (SGFOR) to a grassland (GRASS) showed, indirectly, a positive effect on C stabilization in the soil. In Andisols, soil management practices may affect the stability of Al-mineral-humus com-

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plexes via the soil pH and consequently the available Al content. Due to the reduced pH, more stable soil aggregates could be formed in GRASS and a higher physical protection of its SOC was reached. This was indicated by a smaller difference in WAD between the two fractionation experiments, and a smaller MOM fraction. Apparently, this positive pH effect on soil aggregation was even able to overcome the disruptive effect of frequent ploughing in GRASS. Moreover, though the recalcitrance of the SOM in GRASS was lowest, physical protection in soil aggregates resulted in a reduced C mineralization in this site. Percival et al. (2000) also indicated that land use changes from forest to grassland in Andisols can alter C dynamics, especially if a change in the stability of the Al-mineral-humus complexes occurs.

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**Table 1.** Chemical and physical properties of the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS) for the 0–10, 10–20 and 20–30 cm soil layer (standard deviations between brackets). Significant differences ( $p < 0.05$ ) between land use treatments within one soil depth layer are indicated by different letters.

| Soil layer | Land use class | pH <sub>H2O</sub> | $\rho_b$           | Al <sub>ox</sub>  | Al <sub>py</sub>  | Si <sub>ox</sub> | Allophane         | TC                   |                   | TN                   | C/N               |      |
|------------|----------------|-------------------|--------------------|-------------------|-------------------|------------------|-------------------|----------------------|-------------------|----------------------|-------------------|------|
| cm         |                |                   | Mg m <sup>-3</sup> |                   | %                 |                  |                   | kg C m <sup>-2</sup> | %                 | kg N m <sup>-2</sup> | %                 |      |
| 0–10       | SGFOR          | 6.1 <sup>a</sup>  | 0.42               | 2.03 <sup>b</sup> | 0.64 <sup>b</sup> | 0.96             | 6.00 <sup>a</sup> | 5.8 <sup>a,b</sup>   | 13.9 <sup>a</sup> | 0.31 <sup>b</sup>    | 0.74 <sup>b</sup> | 18.9 |
|            | GRASS          | 5.7 <sup>b</sup>  | 0.60               | 2.12 <sup>a</sup> | 1.11 <sup>a</sup> | 0.81             | 5.22 <sup>b</sup> | 6.7 <sup>a</sup>     | 11.1 <sup>b</sup> | 0.53 <sup>a</sup>    | 0.89 <sup>a</sup> | 12.5 |
|            | PINUS          | 6.2 <sup>a</sup>  | 0.45               | 2.23 <sup>a</sup> | 0.63 <sup>b</sup> | 0.91             | 6.46 <sup>a</sup> | 4.3 <sup>b</sup>     | 9.4 <sup>c</sup>  | 0.31 <sup>b</sup>    | 0.68 <sup>b</sup> | 13.8 |
| 10–20      | SGFOR          | 6.3 <sup>a</sup>  | 0.44               | 2.36 <sup>b</sup> | 0.54 <sup>b</sup> | 1.14             | 6.99 <sup>a</sup> | 4.0 <sup>b</sup>     | 9.3 <sup>b</sup>  | 0.23 <sup>b</sup>    | 0.55 <sup>b</sup> | 16.9 |
|            | GRASS          | 5.4 <sup>b</sup>  | 0.46               | 2.41 <sup>a</sup> | 1.05 <sup>a</sup> | 0.85             | 5.57 <sup>b</sup> | 4.8 <sup>b</sup>     | 10.5 <sup>a</sup> | 0.37 <sup>a</sup>    | 0.81 <sup>a</sup> | 13.0 |
|            | PINUS          | 6.4 <sup>a</sup>  | 0.46               | 2.44 <sup>a</sup> | 0.61 <sup>b</sup> | 0.91             | 6.63 <sup>a</sup> | 3.3 <sup>a</sup>     | 7.1 <sup>c</sup>  | 0.26 <sup>b</sup>    | 0.56 <sup>b</sup> | 12.7 |
| 20–30      | SGFOR          | 6.3 <sup>a</sup>  | 0.52               | 2.29 <sup>b</sup> | 0.43 <sup>b</sup> | 1.18             | 7.59 <sup>a</sup> | 3.4 <sup>b</sup>     | 6.4 <sup>b</sup>  | 0.21 <sup>b</sup>    | 0.40 <sup>b</sup> | 16.1 |
|            | GRASS          | 5.8 <sup>b</sup>  | 0.60               | 2.34 <sup>a</sup> | 1.07 <sup>a</sup> | 0.90             | 5.75 <sup>b</sup> | 5.8 <sup>a</sup>     | 9.8 <sup>a</sup>  | 0.44 <sup>a</sup>    | 0.73 <sup>a</sup> | 13.4 |
|            | PINUS          | 6.7 <sup>a</sup>  | 0.47               | 2.22 <sup>b</sup> | 0.39 <sup>b</sup> | 1.02             | 7.19 <sup>a</sup> | 2.4 <sup>b</sup>     | 5.1 <sup>b</sup>  | 0.18 <sup>b</sup>    | 0.39 <sup>b</sup> | 13.1 |

$\rho_b$  = Bulk density

Al<sub>ox</sub> = Ammonium oxalate extractable Al content

Al<sub>py</sub> = Pyrophosphate extractable Al content

Si<sub>ox</sub> = Ammonium oxalate extractable Si content

Allophane = Estimated allophane content

TC = Total carbon content

TN = Total nitrogen content

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**Table 2.** Distribution of the soil weight according to the size fraction in Fig. 2 and Fig. 3, expressed as weighted average diameter (WAD), of the first and second fractionation procedure, and difference in WAD between the two fractionation procedures for the three soil layers of the second growth forest (SGFOR), grassland (GRASS), and plantation (PINUS). Significant differences ( $p < 0.05$ ) between land use treatments are indicated by different letters.

|                                                      | 0–10 cm          |                  |                  | 10–20 cm         |                  |                  | 20–30 cm         |                  |                    |
|------------------------------------------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|--------------------|
|                                                      | SGFOR            | GRASS            | PINUS            | SGFOR            | GRASS            | PINUS            | SGFOR            | GRASS            | PINUS              |
| WAD first fractionation procedure ( $\mu\text{m}$ )  | 786 <sup>A</sup> | 788 <sup>A</sup> | 790 <sup>A</sup> | 794 <sup>A</sup> | 760 <sup>A</sup> | 758 <sup>A</sup> | 739 <sup>A</sup> | 785 <sup>A</sup> | 647 <sup>B</sup>   |
| WAD second fractionation procedure ( $\mu\text{m}$ ) | 511 <sup>A</sup> | 662 <sup>A</sup> | 572 <sup>A</sup> | 522 <sup>A</sup> | 590 <sup>A</sup> | 585 <sup>A</sup> | 565 <sup>B</sup> | 720 <sup>A</sup> | 526 <sup>B</sup>   |
| Difference in WAD ( $\mu\text{m}$ )                  | 275 <sup>A</sup> | 126 <sup>A</sup> | 218 <sup>A</sup> | 273 <sup>A</sup> | 170 <sup>A</sup> | 173 <sup>A</sup> | 174 <sup>A</sup> | 65 <sup>B</sup>  | 120 <sup>A,B</sup> |

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**Table 3.** Carbon concentrations (%) for the soil layers of the second growth forest (SGFOR), grassland (GRASS), and plantation (PINUS). Significant differences ( $p < 0.05$ ) between aggregate size classes for a certain land use treatment and soil depth are indicated by different letters.

| Aggregate size class                  | Carbon concentration (%) |                   |                  |                  |                   |                  |                  |                  |                  |
|---------------------------------------|--------------------------|-------------------|------------------|------------------|-------------------|------------------|------------------|------------------|------------------|
|                                       | 0–10 cm                  |                   |                  | 10–20 cm         |                   |                  | 20–30 cm         |                  |                  |
|                                       | SGFOR                    | GRASS             | PINUS            | SGFOR            | GRASS             | PINUS            | SGFOR            | GRASS            | PINUS            |
| 250 $\mu\text{m}$ –2000 $\mu\text{m}$ | 14.1 <sub>a,b</sub>      | 11.1 <sub>a</sub> | 9.3 <sub>a</sub> | 9.0 <sub>a</sub> | 10.5 <sub>a</sub> | 7.1 <sub>a</sub> | 6.6 <sub>a</sub> | 9.8 <sub>a</sub> | 5.2 <sub>a</sub> |
| 150 $\mu\text{m}$ –250 $\mu\text{m}$  | 14.6 <sub>a</sub>        | 11.1 <sub>a</sub> | 9.8 <sub>a</sub> | 8.8 <sub>a</sub> | 10.6 <sub>a</sub> | 6.9 <sub>a</sub> | 6.3 <sub>a</sub> | 9.9 <sub>a</sub> | 5.0 <sub>a</sub> |
| 50 $\mu\text{m}$ –150 $\mu\text{m}$   | 14.8 <sub>a</sub>        | 10.7 <sub>a</sub> | 9.9 <sub>a</sub> | 9.3 <sub>a</sub> | 10.5 <sub>a</sub> | 7.2 <sub>a</sub> | 6.7 <sub>a</sub> | 9.7 <sub>a</sub> | 5.2 <sub>a</sub> |
| <50 $\mu\text{m}$                     | 12.2 <sub>b</sub>        | 11.0 <sub>a</sub> | 8.4 <sub>a</sub> | 8.1 <sub>a</sub> | 10.6 <sub>a</sub> | 6.8 <sub>a</sub> | 6.1 <sub>a</sub> | 9.6 <sub>a</sub> | 4.8 <sub>a</sub> |

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**Table 4.** Percentage of total C of the macro organic matter fractions (MOM) for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS). Significant differences under the same soil depth are indicated by different letters.

| Soil depth<br>cm | % of total C     |                  |                  |
|------------------|------------------|------------------|------------------|
|                  | SGFOR            | GRASS            | PINUS            |
| 0–10             | 8.3 <sup>A</sup> | 2.4 <sup>C</sup> | 5.0 <sup>B</sup> |
| 10–20            | 2.8 <sup>B</sup> | 1.6 <sup>C</sup> | 3.6 <sup>A</sup> |
| 20–30            | 3.8 <sup>A</sup> | 1.1 <sup>B</sup> | 3.4 <sup>A</sup> |

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**Table 5.** Weight, C-concentration, and % of total C of the different SOM fractions for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS) for the 0–10 cm soil layer. Significant differences ( $p < 0.05$ ) between land use treatments are indicated by different letters.

|                                   | Weight<br>(g kg <sup>-1</sup> soil) |                  |                   | C-concentration<br>(g C kg <sup>-1</sup> fraction) |                  |                    | % of total C<br>(%) |                   |                    |
|-----------------------------------|-------------------------------------|------------------|-------------------|----------------------------------------------------|------------------|--------------------|---------------------|-------------------|--------------------|
|                                   | SGFOR                               | GRASS            | PINUS             | SGFOR                                              | GRASS            | PINUS              | SGFOR               | GRASS             | PINUS              |
| Whole soil                        | 1000                                | 1000             | 1000              | 139 <sup>A</sup>                                   | 111 <sup>B</sup> | 94 <sup>C</sup>    |                     |                   |                    |
| Size and density fractions        |                                     |                  |                   |                                                    |                  |                    |                     |                   |                    |
| MOM                               | 31.5 <sup>A</sup>                   | 8.6 <sup>C</sup> | 14.4 <sup>B</sup> | 366 <sup>A</sup>                                   | 309 <sup>B</sup> | 325 <sup>A,B</sup> | 8.3 <sup>A</sup>    | 2.4 <sup>C</sup>  | 5.0 <sup>B</sup>   |
| Mineral fraction<br>(150–2000 μm) | 738 <sup>A</sup>                    | 756 <sup>A</sup> | 749 <sup>A</sup>  | 108 <sup>A</sup>                                   | 109 <sup>A</sup> | 90 <sup>B</sup>    | 56.4 <sup>B</sup>   | 74.4 <sup>A</sup> | 71.5 <sup>A</sup>  |
| 50–150 μm                         | 152 <sup>A</sup>                    | 130 <sup>A</sup> | 143 <sup>A</sup>  | 148 <sup>A</sup>                                   | 107 <sup>B</sup> | 99 <sup>B</sup>    | 16.7 <sup>A</sup>   | 12.5 <sup>A</sup> | 15.0 <sup>A</sup>  |
| <50 μm                            | 91 <sup>A</sup>                     | 104 <sup>A</sup> | 93 <sup>A</sup>   | 122 <sup>A</sup>                                   | 110 <sup>B</sup> | 84 <sup>C</sup>    | 7.9 <sup>B</sup>    | 10.3 <sup>A</sup> | 8.3 <sup>A,B</sup> |
| Sum of the fractions              |                                     |                  |                   |                                                    |                  |                    | 89.5 <sup>A</sup>   | 99.6 <sup>A</sup> | 99.7 <sup>A</sup>  |

MOM = Macro organic matter (>150 μm)

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**Table 6.**  $\delta^{13}\text{C}$  values (‰) of the different density and size fractions for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS). Significant differences ( $p < 0.05$ ) between soil organic matter fractions are indicated by different letters.

| Fraction                              | $\delta^{13}\text{C}$ |                    |                      |                    |                    |                      |                    |                    |                    |
|---------------------------------------|-----------------------|--------------------|----------------------|--------------------|--------------------|----------------------|--------------------|--------------------|--------------------|
|                                       | 0–10 cm               |                    |                      | 10–20 cm           |                    |                      | 20–30 cm           |                    |                    |
|                                       | SGFOR                 | GRASS              | PINUS                | SGFOR              | GRASS              | PINUS                | SGFOR              | GRASS              | PINUS              |
|                                       | ‰                     |                    |                      |                    |                    |                      |                    |                    |                    |
| Whole soil                            | -27.2                 | -26.7              | -27.8                | -26.4              | -26.8              | -26.7                | -26.2              | -26.7              | -26.4              |
| Macro organic matter                  | -28.3 <sup>b</sup>    | -28.7 <sup>b</sup> | -29.2 <sup>b</sup>   | -27.7 <sup>b</sup> | -27.8 <sup>b</sup> | -28.2 <sup>b,c</sup> | -27.7 <sup>b</sup> | -27.5 <sup>b</sup> | -28.2 <sup>b</sup> |
| HF                                    | -28.4 <sup>b</sup>    | -28.8 <sup>b</sup> | -29.1 <sup>b,c</sup> | -27.8 <sup>b</sup> | -27.9 <sup>b</sup> | -27.9 <sup>b</sup>   | -27.7 <sup>b</sup> | -27.5 <sup>b</sup> | -28.0 <sup>b</sup> |
| IF                                    | -28.3 <sup>b</sup>    | -28.8 <sup>b</sup> | -29.4 <sup>b,c</sup> | -27.9 <sup>b</sup> | -27.7 <sup>b</sup> | -28.4 <sup>c</sup>   | -27.9 <sup>b</sup> | -27.6 <sup>b</sup> | -28.3 <sup>b</sup> |
| LF                                    | -28.3 <sup>b</sup>    | -28.6 <sup>b</sup> | -29.2 <sup>b</sup>   | -27.7 <sup>b</sup> | -27.8 <sup>b</sup> | -28.2 <sup>b,c</sup> | -27.7 <sup>b</sup> | -27.5 <sup>b</sup> | -28.3 <sup>b</sup> |
| Size fractions                        |                       |                    |                      |                    |                    |                      |                    |                    |                    |
| 250 $\mu\text{m}$ –2000 $\mu\text{m}$ | -27.2 <sup>a</sup>    | -26.7 <sup>a</sup> | -27.9 <sup>a</sup>   | -26.4 <sup>a</sup> | -26.8 <sup>a</sup> | -26.7 <sup>a</sup>   | -26.3 <sup>a</sup> | -26.7 <sup>a</sup> | -26.4 <sup>a</sup> |
| 150 $\mu\text{m}$ –250 $\mu\text{m}$  | -27.2 <sup>a</sup>    | -26.7 <sup>a</sup> | -27.9 <sup>a,c</sup> | -26.4 <sup>a</sup> | -26.8 <sup>a</sup> | -26.7 <sup>a</sup>   | -26.2 <sup>a</sup> | -26.7 <sup>a</sup> | -26.3 <sup>a</sup> |
| 50 $\mu\text{m}$ –150 $\mu\text{m}$   | -27.1 <sup>a</sup>    | -26.6 <sup>a</sup> | -27.7 <sup>a,c</sup> | -26.4 <sup>a</sup> | -26.8 <sup>a</sup> | -26.7 <sup>a</sup>   | -26.2 <sup>a</sup> | -26.7 <sup>a</sup> | -26.3 <sup>a</sup> |
| <50 $\mu\text{m}$                     | -27.2 <sup>a</sup>    | -26.7 <sup>a</sup> | -27.7 <sup>a</sup>   | -26.4 <sup>a</sup> | -26.8 <sup>a</sup> | -26.7 <sup>a</sup>   | -26.1 <sup>a</sup> | -26.6 <sup>a</sup> | -26.2 <sup>a</sup> |

HF = Heavy density fraction (density  $< 1.13 \text{ Mg m}^{-3}$ )

IF = Intermediate density fraction ( $1.13 \text{ Mg m}^{-3} < \text{density} < 1.37 \text{ Mg m}^{-3}$ )

LF = Light density fraction fraction (density  $> 1.37 \text{ Mg m}^{-3}$ )

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**Table 7.** Potential C mineralization rates and decomposition rate constants for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS) for 0–10 cm, 10–20 cm and 20–30 cm soil layer. Significant differences between land use treatments ( $p < 0.05$ ) are indicated by different letters.

| Land use class | Potential mineralization rate                        |                    |                    | Mineralization rate constant |                    |                    |
|----------------|------------------------------------------------------|--------------------|--------------------|------------------------------|--------------------|--------------------|
|                | 0–10 cm                                              | 10–20 cm           | 20–30 cm           | 0–10 cm                      | 10–20 cm           | 20–30 cm           |
|                | g CO <sub>2</sub> -C m <sup>-2</sup> d <sup>-1</sup> |                    |                    | yr <sup>-1</sup>             |                    |                    |
| SGFOR          | 0.495 <sup>a</sup>                                   | 0.192 <sup>a</sup> | 0.142 <sup>a</sup> | 0.029 <sup>a</sup>           | 0.017 <sup>a</sup> | 0.018 <sup>a</sup> |
| GRASS          | 0.196 <sup>c</sup>                                   | 0.092 <sup>a</sup> | 0.087 <sup>a</sup> | 0.011 <sup>b</sup>           | 0.005 <sup>b</sup> | 0.005 <sup>b</sup> |
| PINUS          | 0.266 <sup>b</sup>                                   | 0.135 <sup>a</sup> | 0.087 <sup>a</sup> | 0.023 <sup>a</sup>           | 0.015 <sup>a</sup> | 0.014 <sup>a</sup> |

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**Table 8.** Potential mineralization rates and mineralization rate constants of the light (LF) and heavy (HF) density fraction for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS). Significant differences ( $p < 0.05$ ) are indicated by different letters (upper case letters between LF and HF, lower case letters between land use treatments).

| Land use class | Potential mineralization rate                                   |                                  | Mineralization rate constant    |                                   |
|----------------|-----------------------------------------------------------------|----------------------------------|---------------------------------|-----------------------------------|
|                | LF                                                              | HF                               | LF                              | HF                                |
|                | mg CO <sub>2</sub> -C kg <sup>-1</sup> fraction d <sup>-1</sup> |                                  | yr <sup>-1</sup>                |                                   |
| SGFOR          | 131.2 <sub>a</sub> <sup>A</sup>                                 | 59.7 <sub>a,b</sub> <sup>B</sup> | 0.120 <sub>b</sub> <sup>A</sup> | 0.090 <sub>a,b</sub> <sup>A</sup> |
| GRASS          | 137.4 <sub>a</sub> <sup>A</sup>                                 | 73.0 <sub>a</sub> <sup>B</sup>   | 0.151 <sub>a</sub> <sup>A</sup> | 0.106 <sub>a</sub> <sup>A</sup>   |
| PINUS          | 74.7 <sub>b</sub> <sup>A</sup>                                  | 37.6 <sub>b</sub> <sup>B</sup>   | 0.071 <sub>c</sub> <sup>A</sup> | 0.060 <sub>b</sub> <sup>A</sup>   |

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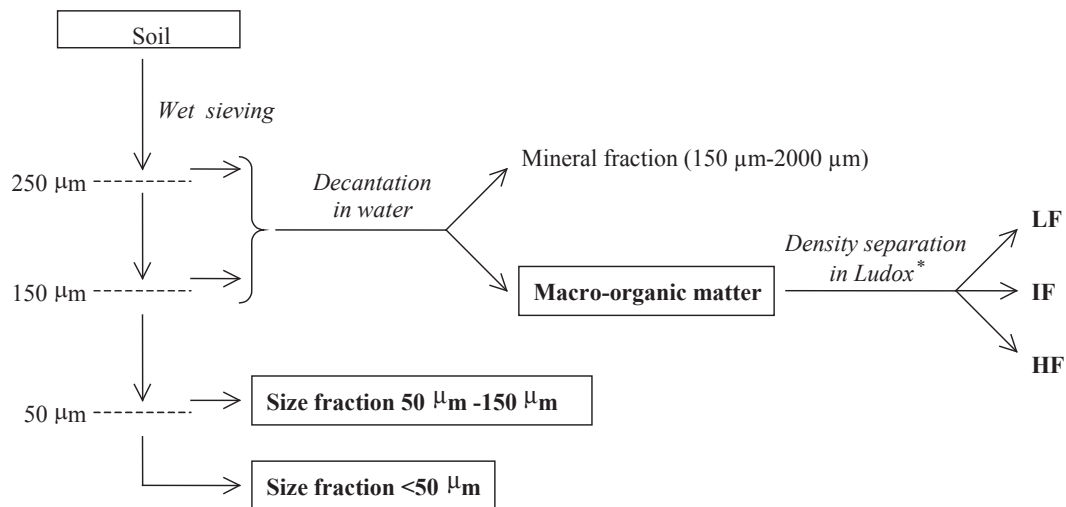
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\*Ludox : silica suspensions with different density

**Fig. 1.** Scheme of the density fractionation of SOM; LF = light density fraction (density  $<1.13 \text{ Mg m}^{-3}$ ), IF = intermediate density fraction ( $1.13 \text{ Mg m}^{-3} < \text{density} < 1.37 \text{ Mg m}^{-3}$ ), HF = heavy density fraction (density  $>1.37 \text{ Mg m}^{-3}$ ). (Adapted from Accoe et al., 2002.)

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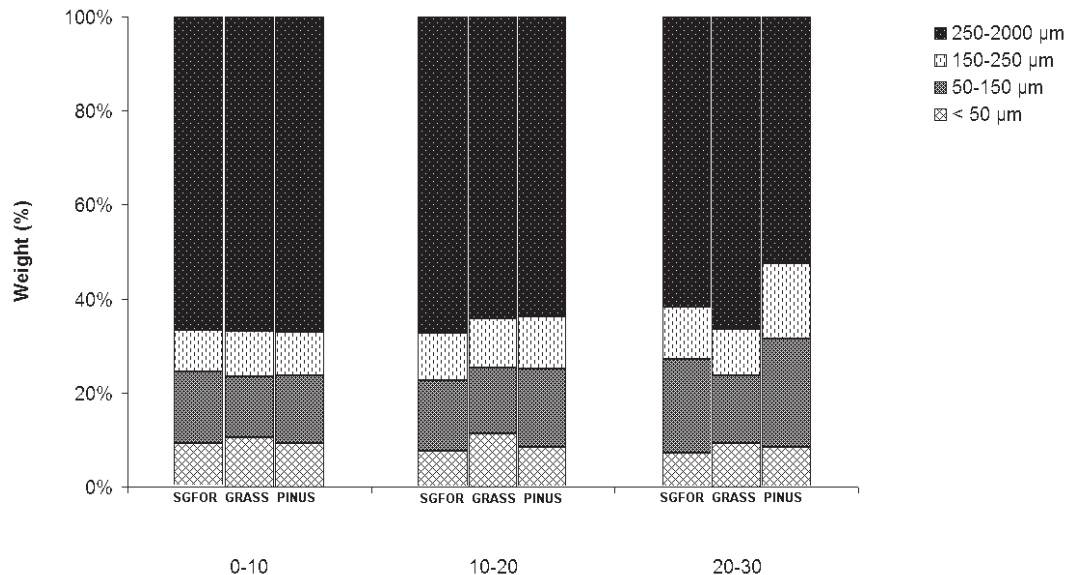
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**Fig. 2.** Distribution of the soil weight over the different size classes (2000–250 μm, 250–150 μm, 150–50 μm, and <50 μm) after the first fractionation procedure (starting from “air-dried” soil) for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS) and for the three soil layers (0–10 cm, 10–20 cm, and 20–30 cm).

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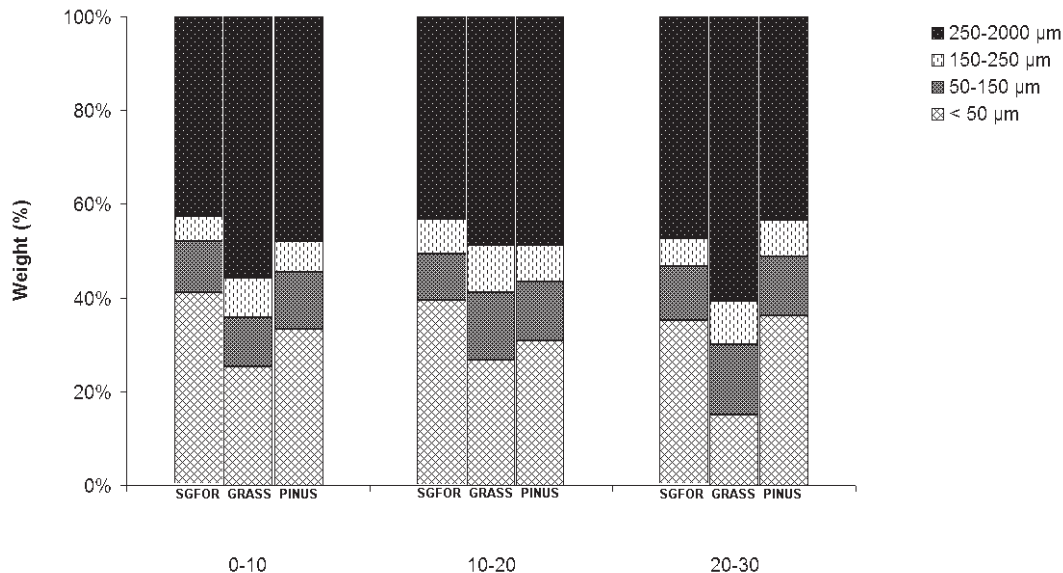
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**Fig. 3.** Distribution of the soil weight over the different size classes (2000–250 μm, 250–150 μm, 150–50 μm, and <50 μm) after the second fractionation procedure (starting from field moist soil) for the second growth forest (SGFOR), grassland (GRASS) and plantation (PINUS) and for the three soil layers (0–10 cm, 10–20 cm, and 20–30 cm).

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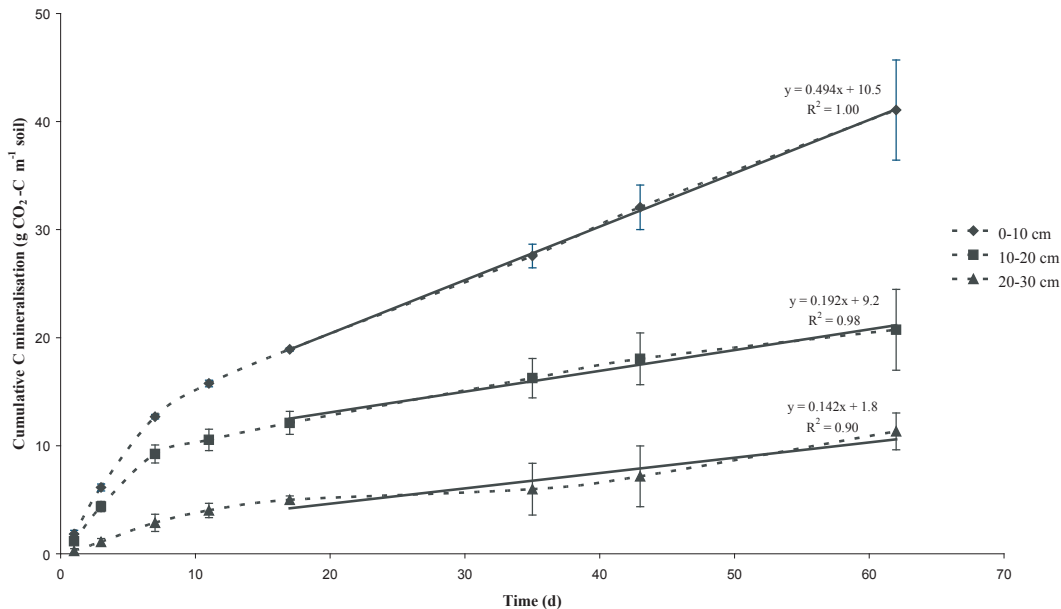
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**Fig. 4.** Evolution of the cumulative C mineralization for the second growth forest (SGFOR) for the 0–10 cm, 10–20 cm and 20–30 cm soil layer (day 1 to day 62) (error bars indicate plus/minus one standard deviation), and linear regression of the curves (from day 17 to day 62).

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