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Multiple Phosphorylations in the C-terminal Tail of Plant Plasma Membrane Aquaporins

ROLE IN SUBCELLULAR TRAFFICKING OF AtPIP2;1 IN RESPONSE TO SALT STRESS[§]

Sodana Prak[‡], Sonia Hem^{§¶}, Julie Boudet[‡], Gaëlle Viennois[‡], Nicolas Sommerer[§], Michel Rossignol[§], Christophe Maurel[‡], and Véronique Santoni^{‡||}

Aquaporins form a family of water and solute channel proteins and are present in most living organisms. In plants, aquaporins play an important role in the regulation of root water transport in response to abiotic stresses. In this work, we investigated the role of phosphorylation of plasma membrane intrinsic protein (PIP) aquaporins in the *Arabidopsis thaliana* root by a combination of quantitative mass spectrometry and cellular biology approaches. A novel phosphoproteomics procedure that involves plasma membrane purification, phosphopeptide enrichment with TiO₂ columns, and systematic mass spectrometry sequencing revealed multiple and adjacent phosphorylation sites in the C-terminal tail of several AtPIPs. Six of these sites had not been described previously. The phosphorylation of AtPIP2;1 at two C-terminal sites (Ser²⁸⁰ and Ser²⁸³) was monitored by an absolute quantification method and shown to be altered in response to treatments of plants by salt (NaCl) and hydrogen peroxide. The two treatments are known to strongly decrease the water permeability of *Arabidopsis* roots. To investigate a putative role of Ser²⁸⁰ and Ser²⁸³ phosphorylation in aquaporin subcellular trafficking, AtPIP2;1 forms mutated at either one of the two sites were fused to the green fluorescent protein and expressed in transgenic plants. Confocal microscopy analysis of these plants revealed that, in resting conditions, phosphorylation of Ser²⁸³ is necessary to target AtPIP2;1 to the plasma membrane. In addition, a NaCl treatment induced an intracellular accumulation of AtPIP2;1 by exerting specific actions onto AtPIP2;1 forms differing in their phosphorylation at Ser²⁸³ to induce their accumulation in distinct intracellular structures. Thus, the present study documents stress-induced quantitative changes in aquaporin phosphorylation and establishes for the first time a link with plant aquaporin subcellular localization. *Molecular & Cellular Proteomics* 7:1019–1030, 2008.

Aquaporins form a family of channel proteins that mediate the transport across membranes of water, small neutral solutes, and occasionally ions (1–3). Aquaporins are present in all living kingdoms and in plants. Aquaporins exhibit a characteristically high multiplicity of forms with for instance 35 members in *Arabidopsis* (4, 5). Based upon their amino acid sequence homology, plant aquaporins can be classified into four subfamilies (4–6). One of these corresponds to the plasma membrane intrinsic proteins (PIPs).¹ The PIPs with 13 members in *Arabidopsis* represent the most abundant aquaporins in the plasma membrane (PM) and can be further divided into two sequence homology groups (AtPIP1 and AtPIP2). Aquaporins are 25–35-kDa proteins that share a typical organization with six transmembrane α -helices interrupted by five connecting loops (loops A–E) (7, 8). In PM aquaporins, the N and C termini as well as loops B and D are exposed in the cytosol, whereas loops A, C, and E face the cell wall.

Plants need to continuously adjust their water status in response to changing environmental conditions, and aquaporins play an important role in these processes (3, 9, 10). In particular, physiological and genetics studies have provided compelling evidence for a role of aquaporins in the regulation, in response to abiotic stresses, of root water transport, *i.e.* root hydraulic conductivity (L_p) (10, 11). For instance, exposure of *Arabidopsis* plants to salt (100 mM NaCl) induced a rapid (half-time, 45 min) and significant decrease (–70%) in L_p that was maintained for at least 24 h (12). Whereas the long term effect of this NaCl stress can be accounted for by an overall transcriptional down-regulation of aquaporins, the molecular mechanisms involved in the early inhibition of L_p by NaCl are not fully understood yet. These mechanisms involve a slight decrease in overall abundance of AtPIP1 proteins as soon as 30 min after exposure to NaCl and a trafficking of AtPIP1 and AtPIP2 isoforms between the PM and intracellular compartments that may contribute to reducing the abundance of AtPIPs at the PM and therefore the hydraulic conductivity of salt-stressed root cells (12). Chilling is another stress that leads to inhibition of L_p , and a relationship

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¹ The abbreviations used are: PIP, plasma membrane intrinsic protein; GFP, green fluorescent protein; L_p , root hydraulic conductivity; PM, plasma membrane; TiO₂, titanium dioxide; WT, wild type; ER, endoplasmic reticulum.

between aquaporin regulation and reactive oxygen species was established in this context (13). In cucumber for instance, hydrogen peroxide (H_2O_2) accumulated in response to chilling, and treatment of roots with exogenous H_2O_2 inhibited L_p , to the same extent as chilling. In *Arabidopsis* a rapid decrease in L_p can also be observed in response to 2 mM H_2O_2 .² Because of its amplitude (>70%) and rapidity (half-time, ≈ 8 min) this decrease is undoubtedly due to a down-regulation of root aquaporins.

Post-translational modifications are central for regulating protein structure and function and thereby for modulating and controlling protein catalytic activity, subcellular localization, stability, and interaction with other partners. Qualitative and quantitative information about post-translational modifications and in particular measurements of their dynamic changes are now critically needed to understand the complexity of cell regulations. Protein phosphorylation is one of the most important and best characterized post-translational modifications. Virtually all cellular processes are regulated in one or multiple ways by reversible phosphorylation, and the identification of the protein kinases and phosphatases, their substrates, and the specific sites of phosphorylation involved is crucial for the understanding of cell signaling. Besides classical methods relying on *in vivo* and *in vitro* labeling or immunodetection of phosphorylated proteins, MS is now widely used for studies on protein phosphorylation (14, 15). Different instrumentations such as ESI- and MALDI-MS systems are now amenable to phosphoprotein analysis (16), and sample preparation procedures have been optimized to enhance phosphopeptide recovery and detection by MS (17). In particular, immobilized metal affinity chromatography (18) or titanium dioxide (TiO_2) microcolumns (19) have proved powerful for the selective enrichment of phosphorylated peptides.

Phosphorylated serine residues have been identified in the N-terminal and C-terminal tails of various plant aquaporins (20–25). In particular, two phosphorylation sites were identified in the C terminus of *Arabidopsis* AtPIP2;1 (26) and AtPIP2;6 (25) and spinach SoPIP2;1 (22) (Table I). AtPIP2;7 also shows double phosphorylation, but only one phosphosite was clearly identified (26) (see Table I). Also all plant AtPIPs show a conserved putative phosphorylation site in loop B (22, 23). Based on functional analyses in *Xenopus* oocytes, it was proposed that phosphorylation of SoPIP2;1 at this site and at Ser²⁶² (in the C terminus) was able to regulate its water transport activity (22). A molecular mechanism for phosphorylation-dependent gating of PIPs has recently been proposed from the atomic structures of SoPIP2;1 in its open and closed conformations (8). Mammalian aquaporins also carry multiple phosphorylation sites, and by contrast to plant aquaporins, phosphorylation of mammalian aquaporin-2 is not involved in gating but rather regulates the shuttling of the protein between the PM and intracellular compartments (27, 28).

The purpose of this work was to study the role of plant PM aquaporin phosphorylation in regulating the root water permeability in response to NaCl and H_2O_2 treatments. For this, a systematic inventory of phosphorylation sites in the C terminus of AtPIP aquaporins was performed, and novel phosphoresidues were discovered. Because of the emerging role of stimulus-dependent trafficking of plant aquaporins between the PM and intracellular compartments (12, 29, 30), the role of aquaporin phosphorylation in this process was investigated. The results point to a specific phosphorylated site in the C terminus of AtPIP2;1 that regulates the trafficking of this aquaporin in control conditions and in response to an NaCl treatment.

EXPERIMENTAL PROCEDURES

Reagents—Endoproteinase Lys-C was purchased from Calbiochem. Synthetic PIP2 peptides (²⁷⁷SLGSFRSAANV²⁹⁷), either unmodified or singly phosphorylated at Ser²⁸⁰, were isotopically labeled on Arg²⁸¹ with ¹³C and ¹⁵N to induce a 10-Da mass increment (Sigma). The same PIP2 peptide but diphosphorylated was isotopically labeled on Ala²⁸⁴ and Ala²⁸⁵ with ¹³C to induce a 6-Da mass increment (NeoMPS, Strasbourg, France). TiO_2 beads were obtained by disassembling TiO_2 guard columns purchased from GL Sciences Inc. (Tokyo, Japan). The 3M EmporeTM C₈ disks were from 3M Bioanalytical Technologies (St. Paul, MN). GELoader tips were from Eppendorf (Hamburg, Germany). 2,5-Hydroxybenzoic acid and α -cyano-4-hydroxycinnamic acid were from Sigma-Aldrich. All other chemicals and reagents were of the highest commercially available grade.

Plant Materials and Treatments—*Arabidopsis thaliana* ecotype Columbia (Col-0), plants were cultivated in hydroponic conditions as described previously (31). Briefly plants were cultivated in a growth chamber at 20 °C with an 8-h light (150 microeinstein $m^{-2} s^{-1}$)/16-h dark cycle at 70% relative humidity. Plants were mounted on 35 × 35 × 0.6-cm polystyrene rafts floating in a basin filled with 8 liters of nutrient medium (1.25 mM KNO_3 , 0.75 mM $MgSO_4$, 1.5 mM $Ca(NO_3)_2$, 0.5 mM KH_2PO_4 , 0.1 mM Na_2SiO_3 , 50 μM FeEDTA, 50 μM H_3BO_3 , 12 μM $MnSO_4$, 1 μM $ZnSO_4$, 0.7 μM $CuSO_4$, 0.24 μM MoO_4Na_2) and cultivated for up to 7 weeks. The effects of NaCl and H_2O_2 were then studied by complementing the nutrient solution with 100 mM NaCl for 2 and 4 h or 2 mM H_2O_2 for 15 min prior to root excision. Transgenic seedlings were cultivated for 8 days on half-strength Murashige and Skoog medium (32) without any antibiotic selection. The plantlets were then transferred for 2 or 4 h into a nutrient solution as described above complemented or not with 100 mM NaCl.

Purification of PIPs—A microsomal fraction was obtained from roots (31). Plasma membrane vesicles were purified by aqueous two-phase partitioning of the microsomal fraction in a mixture of polyethylene glycol 3350/dextran T-500, 6.4% (w/w) each in the presence of 5 mM KCl, as described previously (31). Protein concentration was measured using a modified Bradford procedure (31). The mean yield of PM extraction was 20 μg of protein/g of fresh weight. Extrinsic membrane proteins were stripped with a urea and NaOH treatment according to a previously described procedure (31). The abundance of AtPIP2 isoforms in PM samples was evaluated by an ELISA using an antibody raised against the last 17 amino acids of the AtPIP2;1 sequence as described previously (33). The mean yield of AtPIP2 isoform was 5.3 pmol of PIP2/ μg of PM proteins. Proteins were separated by SDS-PAGE on 12% acrylamide gels (31).

Protein Digestion and Phosphopeptide Purification—The migrating band at 28 kDa was excised from SDS-PAGE and prepared for proteolytic digestion as described previously (31). Gel pieces containing 350 pmol of AtPIP2 aquaporins were reswollen in the presence of

² Y. Boursiac, J. Boudet, O. Postaire, D.-T. Luu, C. Tournaire-Roux, and C. Maurel, submitted manuscript.

Lys-C at an enzyme:aquaporin ratio of 1:25 at 37 °C for 16 h. The supernatant of the digest was collected, and the remaining peptides were extracted in 0.1% TFA, 60% acetonitrile by sonication for 15 min. Supernatants were pooled, and the final volume was reduced to 10 μ l using a centrifuge evaporator. To build up a TiO₂ microcolumn, a small piece was stamped out of an Empore C₈ disk by using a 200- μ l pipette tip and placed at the constricted end of the GELoader tip, and TiO₂ beads in suspension in acetonitrile were packed (34). The protein digest was then diluted in a loading buffer containing 80% acetonitrile and 0.1% TFA and loaded on the column, and the column was washed with 30 μ l of loading buffer. Phosphopeptides were eluted with 3 μ l of NH₄OH at pH 12. 0.8 μ l of eluted peptides was mixed with 0.8 μ l of 20 mg/ml 2,5-hydroxybenzoic acid dissolved in acetonitrile, water, and phosphoric acid (50:44:6, v/v/v) and spotted onto the MALDI target for crystallization. The quantification of At-PIP2;1 C-terminal phosphorylation was performed by adding the synthetic labeled peptides corresponding to the C terminus of At-PIP2;1 (unmodified:singly phosphorylated:diphosphorylated, 1:1:3) to the protein digest prior to loading onto the TiO₂ column. The abundance of the unmodified form was quantified from the flow-through of the TiO₂ column. The flow-through was desalted using ZipTip μ C₁₈ columns (Millipore, Bedford, MA). The desalted sample (0.8 μ l) was mixed with 0.8 μ l of matrix solution (α -cyano-4-hydroxycinnamic acid at half-saturation in 1:1 (v/v) H₂O/acetonitrile, 0.1% TFA) and spotted onto the MALDI target.

Mass Spectrometric Analysis—MALDI-TOF MS and MS/MS analyses were performed, in positive reflector mode, using an UltraFlex II mass spectrometer (Bruker Daltonics, Bremen, Germany) equipped with a smartbeam™ laser. MS/MS spectra were obtained by PSD-LIFT™ without adding a collision gas. MS data were analyzed using the FlexAnalysis software (Bruker Daltonics). All MS and MS/MS spectra shown were externally calibrated and are raw data spectra, *i.e.* without recalibrating, smoothing, or base-line subtracting. MS and MS/MS spectra annotation was performed manually. *De novo* sequencing was performed and was facilitated by the knowledge of all aquaporin sequences. All peptides proposed as phosphorylated were first checked for the presence of the major fragment ion [MH - H₃PO₄]⁺ = MH - 98 Da corresponding to the loss of the phosphate moiety. In addition, all MS/MS spectra were carefully checked manually for assignment of phosphorylation sites.

Gene Constructs and Expression in Transgenic Plants—Mutagenesis of AtPIP2;1 C-terminal phosphorylation sites was carried out by PCR on a cDNA of AtPIP2;1 fused by its N terminus to the green fluorescent protein (GFP) (GFP-PIP2;1). For this, we used a sense primer containing an XhoI restriction site: 5'-TTTCTCGAGATGGT-GAGCAAGGGCGAGG-3'. The antisense primer allows the introduction of an XbaI restriction site as well as the desired mutation. The mutagenic primers used to generate the following mutations (bold characters) were: S280A, 5'-TTC TAG ATT AGA CGT TGG CAG CAC TTC TGA **ATG CTC** C-3'; S283A, 5'-TTC TAG ATT AGA CGT TGG **CAG CAG CTC TG**-3'; and S283D, 5'-TTT CTA GAT TAG ACG TTG GCA GCA **TCT CTG AA**-3'. The fragments amplified by PCR were digested by XhoI and XbaI and cloned in a pBluescript vector. The presence of the mutations was checked by DNA sequencing (Genoscreen, Lille, France). The GFP-PIP2;1 sequences were placed under the control of a cauliflower mosaic virus 35S and *RbcS* terminator by cloning into the EcoRI and ClaI sites of a pGREEN vector (35). The constructs were then transferred into *Agrobacterium tumefaciens* strain GV3101 by electroporation with a selection for tetracycline, rifampicin, and kanamycin resistance. The bacterial strains were used for transformation of *Arabidopsis* Col-0 by the floral dip method (36). To select for transformed plants, seeds were surface-sterilized and germinated in a medium containing a half-strength Murashige and Skoog medium (32) complemented with 7 g/liter agar and 0.04 g/liter

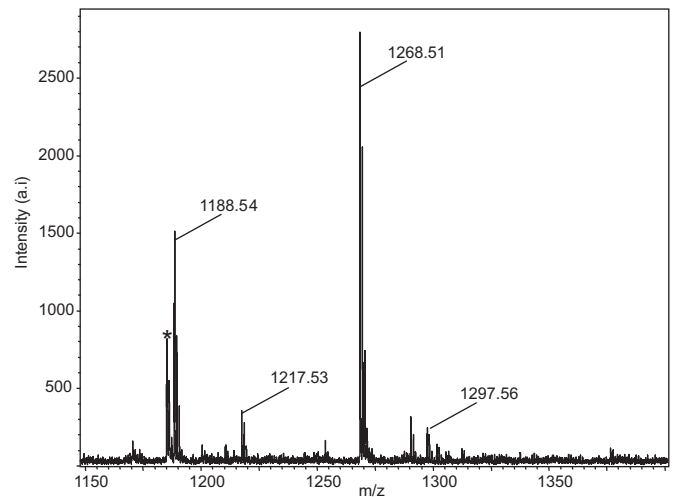


FIG. 1. MALDI-TOF MS spectrum of phosphopeptides from plant PM aquaporins. The 28-kDa band from root PM enriched in hydrophobic proteins was digested by Lys-C. Phosphopeptides were enriched using a TiO₂ column. All C-terminal phosphopeptides expected from root AtPIP aquaporins cannot be simultaneously recorded onto the same MS spectrum. The present spectrum illustrates the presence of the singly (*m/z* 1188.54) and diphosphorylated (*m/z* 1268.51) peptides of AtPIP2;1 and/or AtPIP2;2 (see text) and of the singly (*m/z* 1217.53) and diphosphorylated (*m/z* 1297.56) peptides of AtPIP2;7. *, metastable decomposition of peptide with *m/z* 1268.51. *a.i.*, absolute intensity.

hygromycin as described previously (12). Two, three, two, and two independent lines were obtained for the GFP-PIP2;1, GFP-PIP2;1-S280A, GFP-PIP2;1-S283A, and GFP-PIP2;1-S283D genotypes, respectively.

Microscopic Observations of Transgenic Plants—The roots of transgenic lines expressing GFP-PIP2;1 fusions were observed under a confocal microscope (LSM 510 AX70, Zeiss, Göttingen, Germany) with two to three independent lines characterized for each construct. The argon laser wavelength was 488 nm; GFP emission was detected with the filter set for fluorescein isothiocyanate (bandwidth from 500 to 530 nm). The acquisition software used was LSM 510 version 3.0, and the image processing software was Zeiss LSM Image Browser. Cells were individually examined through a z series of images.

RESULTS

A Phosphoproteomics Analysis Reveals Novel Phosphorylation Sites in the C-terminal Tail of AtPIP Aquaporins—A PM fraction was purified from *Arabidopsis* roots by aqueous two-phase partitioning and enriched in hydrophobic proteins with a urea and NaOH treatment (31). This extract was used to make a systematic inventory of the C-terminal phosphorylations of AtPIP2 isoforms. For this, the extract was first treated with the endoproteinase Lys-C, which is predicted to release the C-terminal tail of all AtPIP2 aquaporins. Phosphorylated C-terminal peptides were then enriched using TiO₂ microcolumns (19). A typical MALDI MS spectrum is shown in Fig. 1. The candidate phosphopeptides were initially assigned by MALDI-TOF MS from 79.96-Da mass increments per phosphate moiety relative to the unmodified peptides. During MALDI-TOF MS, phosphopeptides also lose phosphoric acid

TABLE I
Phosphorylation sites in the C-terminal tail of PIP aquaporins

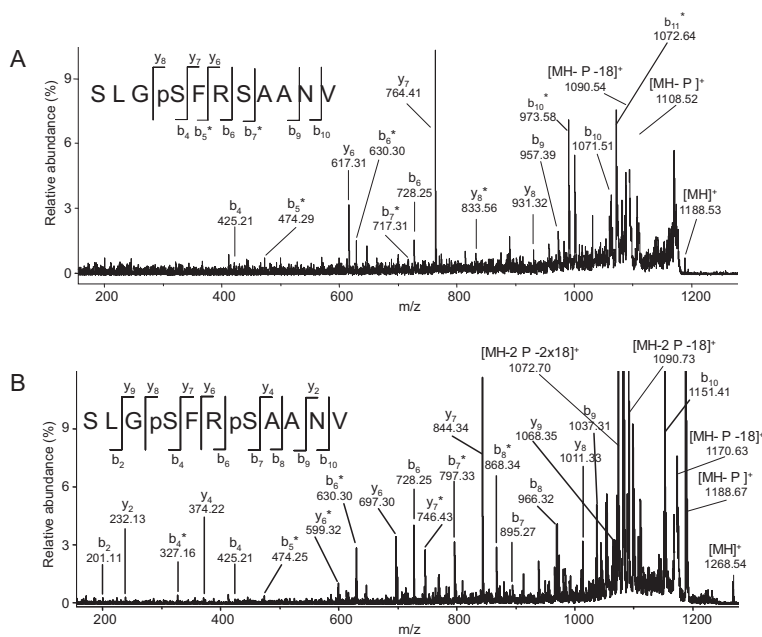
The first and second columns describe the name of the aquaporin and the peptide sequence, respectively. The third column (n_ph) refers to the number of phosphorylation sites present in the peptide. pS, phosphorylated serine; pT, phosphorylated threonine.

Protein	Sequence	n_ph	Ref.
AtPIP2;1/2;2/2;3	²⁷⁷ SLGpSFR ²⁸²	1	25, 31 ^a
AtPIP2;1/2;2/2;3	²⁷⁷ SLGpSFRSAANV ²⁸⁷	1	Present work
AtPIP2;1/2;2/2;3	²⁷⁷ SLGpSFRpSAANV ²⁸⁷	2	25, 26, and present work
AtPIP2;4	²⁷⁷ ALGpSFGSFGSFRSFA ²⁹¹	1	Present work
AtPIP2;4	²⁷⁷ ALGpSFGpSFGSFRSFA ²⁹¹	2	Present work
AtPIP2;4	²⁷⁷ ALGpSFGpSFGpSFRsFA ²⁹¹	3	Present work
AtPIP2;4	²⁷⁷ ALGpSFGpSFGSFRpSFA ²⁹¹	3	Present work
AtPIP2;6	²⁸² pSQLHELHA ²⁸⁹	1	25
AtPIP2;7	²⁷⁰ ALGpSFRSNATN ²⁸⁰	1	Present work
AtPIP2;7	²⁷⁰ ALGpSFRpSNATN ²⁸⁰	2	26, 34, ^b and present work
AtPIP2;7	²⁷⁰ ALGpSFRpSNApTN ²⁸⁰	3	Present work
SoPIP2;1	²⁷¹ ALGpSFRSNPTN ²⁸¹	1	22
SoPIP2;1	²⁷¹ ALGpSFRpSNPTN ²⁸¹	2	22

^a One phosphorylation site was detected but not identified.

^b Two phosphorylation sites were detected, but only Ser²⁷³ was identified as phosphorylated.

FIG. 2. Phosphopeptide sequencing by MALDI-TOF/TOF of the C-terminal tail of AtPIP2;1. A, MS/MS spectrum of singly phosphorylated ²⁷⁷SLGpSFRSAANV²⁸⁷ (*m/z* 1188.53). *y*₇ and *y*₈ ions allowed identification of Ser²⁸⁰ as the phosphorylated residue. B, MS/MS spectrum of the corresponding diphosphorylated peptide (*m/z* 1268.54). *y*₄, *y*₅, *y*₇, and *y*₉ ions allowed identification of the two phosphorylated residues as Ser²⁸⁰ and Ser²⁸³. *, fragment ions arising from loss of phosphoric acid (−98 Da). pS, phosphorylated serine; [MH]⁺, precursor ion; [MH − P]⁺, precursor ion with a loss of one metaphosphoric acid (−80 Da); [MH − P − 18]⁺, precursor ion with a loss of one phosphoric acid (−98 Da); [MH − 2P − 18]⁺, precursor ion with a loss of one metaphosphoric acid (−80 Da) and one phosphoric acid (−98 Da); [MH − 2P − 2 × 18]⁺, precursor ion with a loss of two phosphoric acids (−196 Da).



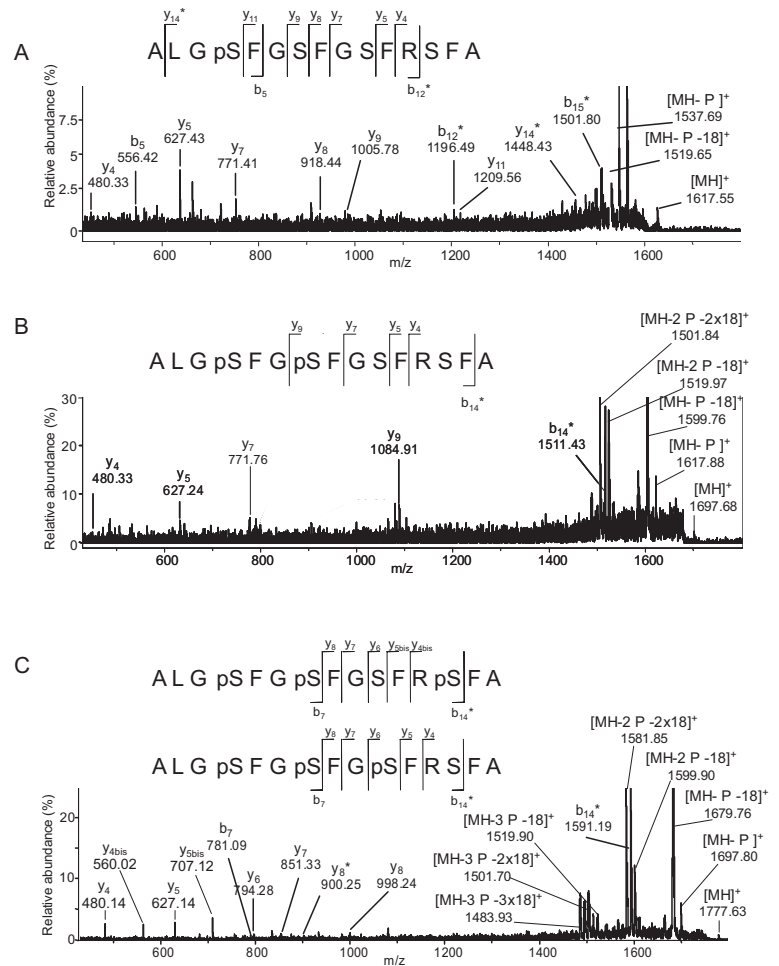
as H₃PO₄ (98 Da) with the concomitant production of metastable ions with an apparent mass loss of 83 Da. Their presence was utilized as reliable indicators for phosphopeptides. A computational analysis of the mass spectra and comparison with the known aquaporin sequences allowed prediction of the presence of putative singly and diphosphorylated peptides of AtPIP2;1, AtPIP2;2, AtPIP2;3, AtPIP2;4, and AtPIP2;7 (Table I). In addition, triphosphorylated forms could be assigned to AtPIP2;4 and AtPIP2;7 isoforms (Table I). The putative phosphopeptides assigned to AtPIP isoforms were then sequenced by MALDI-TOF/TOF for confirmation and for identification of the phosphorylated residues. The positioning of the phosphorylated residue(s) was more specifically based on the

identification of dehydroalanine residue-containing ions in the MS/MS spectrum.

We note that AtPIP2;1 and AtPIP2;2 are among the most abundant aquaporins in roots (12, 31, 37). The phosphopeptides derived from these isoforms were systematically detected in MS spectra. By contrast, phosphopeptides attributed to the less abundant AtPIP2 isoforms such as AtPIP2;4 and AtPIP2;7 were only occasionally detected as shown in Fig. 1.

Because they share identical C-terminal sequences, the AtPIP2;1, AtPIP2;2, and AtPIP2;3 that were predicted to be singly and diphosphorylated could not be distinguished in our study. By contrast to AtPIP2;1 and AtPIP2;2, AtPIP2;3 is barely expressed in roots (31), and for the sake of simplifica-

FIG. 3. Phosphopeptide sequencing by MALDI-TOF/TOF of the C-terminal tail of AtPIP2;4. A, MS/MS spectrum of singly phosphorylated ²⁷⁷ALGSFGSFGSFRSFA²⁹¹ (*m/z* 1617.55). *b*₅ and *y*₁₁ ions allowed identification of Ser²⁸⁰ as the phosphorylated residue. B, MS/MS spectrum of the corresponding diphosphorylated peptide (*m/z* 1697.68). *y*₄, *y*₅, *y*₇, and *y*₉ ions allowed identification of phosphorylated Ser²⁸⁰ and Ser²⁸³. C, MS/MS spectrum of the corresponding triphosphorylated peptide (*m/z* 1777.63) that is a mixture of two forms: *b*₇ ion allowed determination of the phosphorylation of the two residues Ser²⁸⁰ and Ser²⁸³, *y*₄, *y*₅, and *y*₆ ions indicated that Ser²⁸⁶ can be phosphorylated; *y*_{4bis}, *y*_{5bis}, and *y*₆ showed that alternatively Ser²⁸⁹ can carry the third phosphorylation. *, *pS*, [MH]⁺, [MH - P]⁺, [MH - P - 18]⁺, [MH - 2P - 18]⁺, and [MH - 2P - 2 × 18]⁺ are as explained in the legend of Fig. 2.



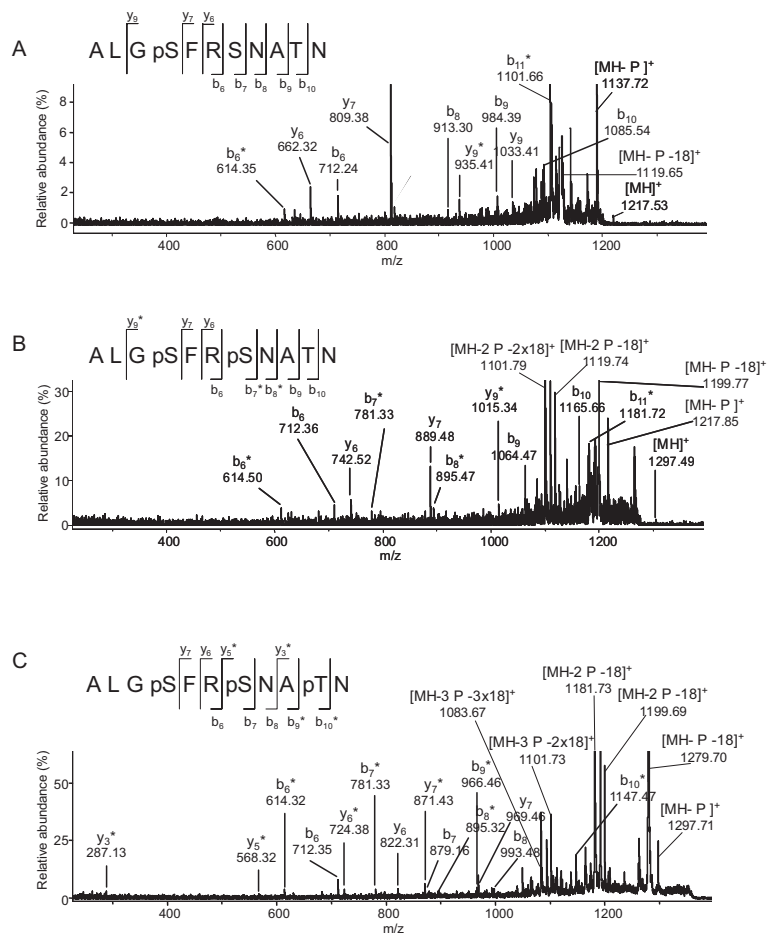
tion, these peptides were attributed to AtPIP2;1 here. Using this approach, the sequencing of the peptides at *m/z* 1188.54 and *m/z* 1268.51 revealed single and diphosphorylation of AtPIP2;1 on Ser²⁸⁰ and Ser²⁸⁰ and Ser²⁸³, respectively (Fig. 2). The fragmentation of the C-terminal peptides of AtPIP2;4 revealed a single phosphorylation on Ser²⁸⁰ (peptide at *m/z* 1617.55) and a diphosphorylation on Ser²⁸⁰ and Ser²⁸³ (peptide at *m/z* 1697.68). The MS/MS analysis of a putative low abundance triphosphorylated peptide of AtPIP2;4 (peptide at *m/z* 1777.63) revealed that it actually corresponded to a mixture of two isobaric forms of the AtPIP2;4 C-terminal tail (Fig. 3C). Sequencing showed a conserved phosphorylation at residues Ser²⁸⁰ and Ser²⁸³ and an additional phosphorylation of either Ser²⁸⁶ or Ser²⁸⁹. The C-terminal tail of AtPIP2;7 was found to be singly phosphorylated on Ser²⁷³ (peptide at *m/z* 1217.53), diphosphorylated on Ser²⁷³ and Ser²⁷⁶ (peptide at *m/z* 1297.49), or triphosphorylated on Thr²⁷⁹ in addition to the two Ser residues (peptide at *m/z* 1377.51) (Fig. 4). Table I summarizes all phosphorylation sites identified in this work. Table I shows that six new phosphorylation sites were identified in the C-terminal tail of aquaporins of *Arabidopsis* in addition to four previously known phosphorylation sites. This

work also allowed the discovery that not only Ser residues but also a Thr residue can be phosphorylated in plant aquaporins.

In theory, by considering all peptide forms, two, three, and four phosphorylation sites should result in peptides with four, eight, and 16 phosphorylation states, respectively. However, a lower number of peptides was observed in AtPIP2;1, AtPIP2;4, and AtPIP2;7, suggesting that phosphorylation events in these proteins might be interdependent. It appeared that phosphorylation of the most distal residues in a C-terminal sequence was only observed in association to phosphorylation of upstream neighboring Ser residue(s). In AtPIP2;1 for instance, Ser²⁸³ was never found to be singly phosphorylated, and its phosphorylation was always associated to that of Ser²⁸⁰. Similarly in AtPIP2;7 phosphorylation of Thr²⁷⁹ was linked to that of Ser²⁷⁶, which was itself linked to that of Ser²⁷³. In AtPIP2;4, the phosphorylation of the distal residues Ser²⁸⁶ and Ser²⁸⁹ appeared to be linked to phosphorylation of both Ser²⁸⁰ and Ser²⁸³.

C-terminal Phosphorylation of AtPIP2;1 Is Quantitatively Modified following Treatments of Plants with NaCl or H₂O₂—AtPIP2;1 is one of most abundant aquaporins in *Arabidopsis* root and therefore must significantly contribute to L_p, and to

FIG. 4. Phosphopeptide sequencing by MALDI-TOF/TOF of the C-terminal tail of AtPIP2;7. A, MS/MS spectrum of singly phosphorylated 270 ALGSFRSNA-TN 280 (m/z 1217.53). b_6 , y_7 , and y_9 ions allowed identification of the phosphorylated residue as Ser 273 . B, MS/MS spectrum of the corresponding diphosphorylated peptide (m/z 1297.49). b_6 and b_7 ions allowed identification of Ser 273 and Ser 276 as the phosphorylated residues. C, MS/MS spectrum of the corresponding triphosphorylated peptide (m/z 1377.51). b_6 , b_7 , b_{10} , y_3 , and y_5 ions allowed identification of Ser 273 , Ser 276 , and Thr 279 as the phosphorylated residues. *, pS , $[MH]^+$, $[MH - P]^+$, $[MH - P - 18]^+$, $[MH - 2P - 18]^+$, and $[MH - 2P - 2 \times 18]^+$ are as explained in the legend of Fig. 2. pT , phosphorylated threonine.



its regulation. In addition, AtPIP2;1 displays a less complex phosphorylation pattern than other AtPIP2 isoforms. For these reasons, AtPIP2;1 was chosen as a model root aquaporin, and qualitative and/or quantitative changes in its C-terminal phosphorylation status in response to NaCl and H $_2$ O $_2$ treatments were investigated. MS/MS sequencing of the singly and diphosphorylated forms (m/z 1188.53 and m/z 1268.54) of AtPIP2;1 in plants exposed to a 2- or a 4-h treatment with 100 mM NaCl revealed that, as in control conditions, the singly phosphorylated residue was Ser 280 (supplemental Fig. 1) and that diphosphorylation had occurred on Ser 280 and Ser 283 (supplemental Fig. 2). Thus, the NaCl treatment did not qualitatively change the phosphorylation pattern of AtPIP2;1. Similar conclusions were obtained in plants treated by 2 mM H $_2$ O $_2$ for 15 min (data not shown).

The unmodified C-terminal AtPIP2;1 peptide and its singly and diphosphorylated forms were quantified using a strategy adapted from the absolute quantification method (38). For this, we used three reference synthetic peptides that, with respect to endogenous peptides, had incorporated stable isotopes. This labeling induced a mass increment of 10, 10, and 6 Da with respect to the unmodified, singly phosphorylated, and diphosphorylated endogenous peptides, respec-

tively (Fig. 5). The reference peptides were introduced into the peptide digest prior to the purification of phosphopeptides with TiO $_2$ microcolumns. The phosphorylated and unmodified peptides were quantified in the MALDI MS spectra arising from the elution of the microcolumns and from their flow-through, respectively. More specifically, native peptides were quantified from the ratio of the monoisotopic peak area of the native and of the corresponding reference peptide (Fig. 5). Four independent biological experiments were performed to quantitatively study the phosphorylation status of AtPIP2;1 in plants that had been exposed to a 2- or 4-h treatment with 100 mM NaCl or to a 15-min treatment with 2 mM H $_2$ O $_2$. Fig. 6A shows that the 2- or 4-h NaCl treatment induced a statistically significant 30% decrease in the abundance of the diphosphorylated form (Mann and Whitney, $p < 0.05$). A tendency toward an increase in abundance of the unmodified and singly phosphorylated forms was also observed in these experiments (Fig. 6A). By contrast, an H $_2$ O $_2$ treatment induced a statistically significant 2-fold decrease in abundance of the unmodified form (Mann and Whitney, $p < 0.05$) (Fig. 6B). This decrease was accompanied by a slight relative (20%) increase in the abundance of the diphosphorylated form (Fig. 6B).

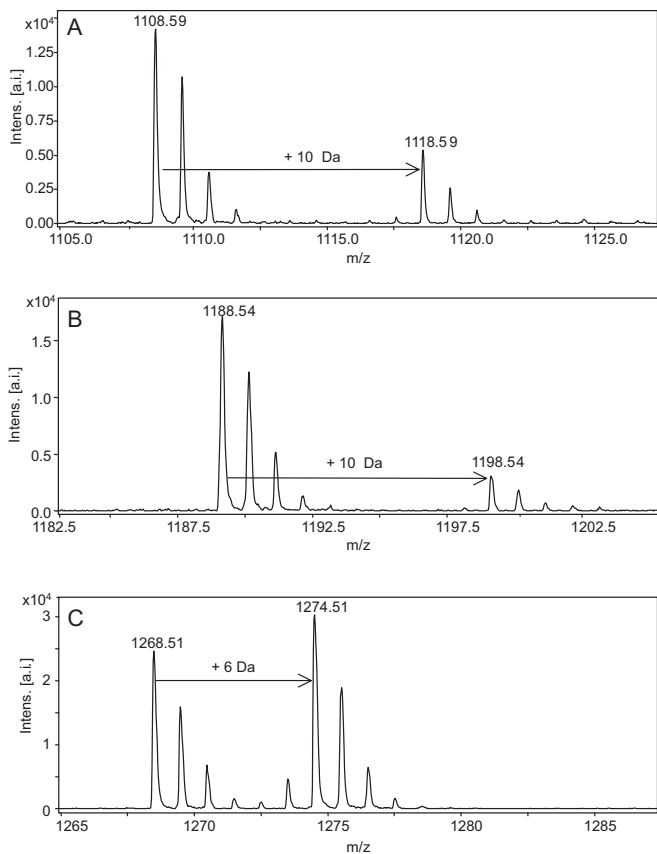


FIG. 5. Principle for quantification of phosphorylation of the C-terminal tail of AtPIP2;1. Synthetic peptides were added to the peptide digest prior to phosphopeptide enrichment on TiO₂ columns (see “Experimental Procedures”). The native and synthetic unmodified peptides (A) were purified from the column flow-through and analyzed by MS. Native and synthetic singly (B) and diphosphorylated (C) peptides were enriched after elution from the columns and analyzed by MS. The synthetic unmodified and singly phosphorylated peptides displayed a 10-Da mass increment when compared with their native counterparts (A and B). A 6-Da mass increment was displayed by the diphosphorylated synthetic peptide (C). The ratio between the monoisotopic peak surface of native and synthetic peptides was used to determine the absolute quantity of native peptides. The proportion of each C-terminal form of AtPIP2;1 in each sample was then calculated. Intens., intensity; a.i., absolute intensity.

The Phosphorylation of Ser²⁸³ Is Involved in the Targeting of AtPIP2;1 to the PM and in Its Intracellular Accumulation upon a NaCl Treatment—The role of specific phosphorylated residues in gating plant aquaporins has been well described (8, 21, 22). By contrast, the role of phosphorylation in the regulation of plant aquaporin trafficking has not yet been investigated. We previously showed that a fusion of AtPIP2;1 with GFP labels the PM of root cells and that an NaCl treatment induces the additional labeling of intracellular structures, suggesting a relocalization mechanism in response to NaCl (12).² The finding that an NaCl treatment decreased the phosphorylation of Ser²⁸³ of AtPIP2;1 prompted us to investigate the role of this modification in the subcellular trafficking of the

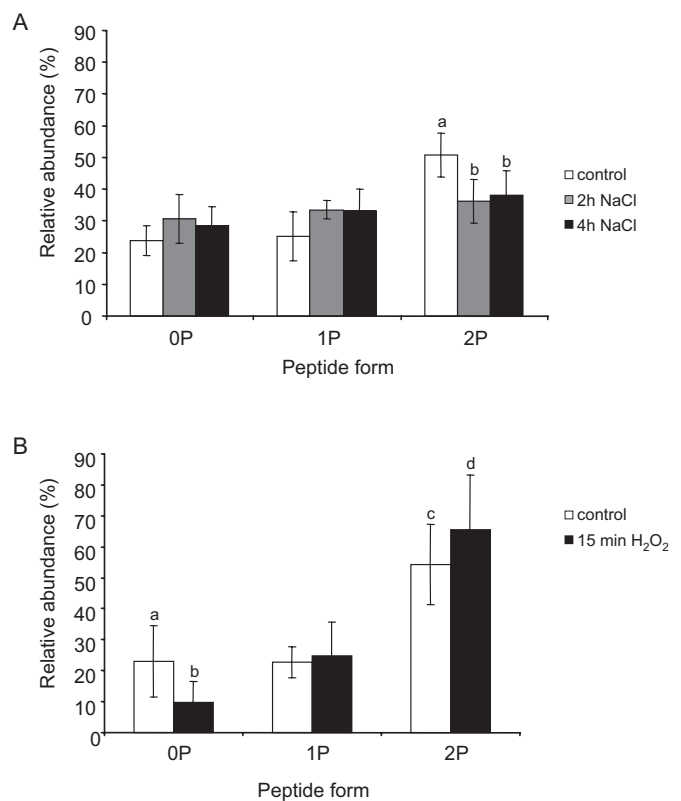


FIG. 6. Quantification of C-terminal phosphorylation of AtPIP2;1 upon NaCl (A) and H₂O₂ (B) treatments. A, plants were either untreated (white bars) or treated with 100 mM NaCl for 2 h (gray bars) or 4 h (black bars). The proportion of the unmodified (0P), singly phosphorylated on Ser²⁸⁰ (1P), and diphosphorylated on Ser²⁸⁰ and Ser²⁸³ (2P) peptides was quantified in plant extracts as exemplified in Fig. 5. Letters a–d indicate statistically different values (Mann and Whitney, $p < 0.05$). B, plants were untreated (white) or treated with 2 mM H₂O₂ for 15 min (black). The same procedures and conventions as in A were used.

protein. For this, GFP was fused to the N-terminal tail of AtPIP2;1, either wild type (WT) or carrying Ser to Ala mutations at positions 280 (S280A) or 283 (S283A) or a Ser to Asp mutation at position 283 (S283D). The fusion proteins were expressed in transgenic *Arabidopsis*, and their expression in epidermal cells at 1 cm from the apex was observed by laser-scanning confocal microscopy. In normal growth conditions, root cells of plants expressing the fusions of GFP with WT-PIP2;1 (GFP-PIP2;1) or the PIP2;1-S280A mutant (GFP-PIP2;1-S280A) showed a labeling pattern consistent with predominant localization of the proteins in the PM (Fig. 7A). By contrast, plants expressing GFP-PIP2;1-S283A showed an intracellular reticulation pattern in 40% of root cells (Fig. 7, A and B). Because of a pronounced localization around the nucleus and its fuzzy aspect throughout the cell, this intracellular staining was partly assigned to endoplasmic reticulum (ER) structures (39). Interestingly plants expressing GFP-PIP2;1-S283D, whereby the introduced mutation is supposed to mimic a constitutive phosphorylation at position 283, displayed a consistent PM staining in root cells

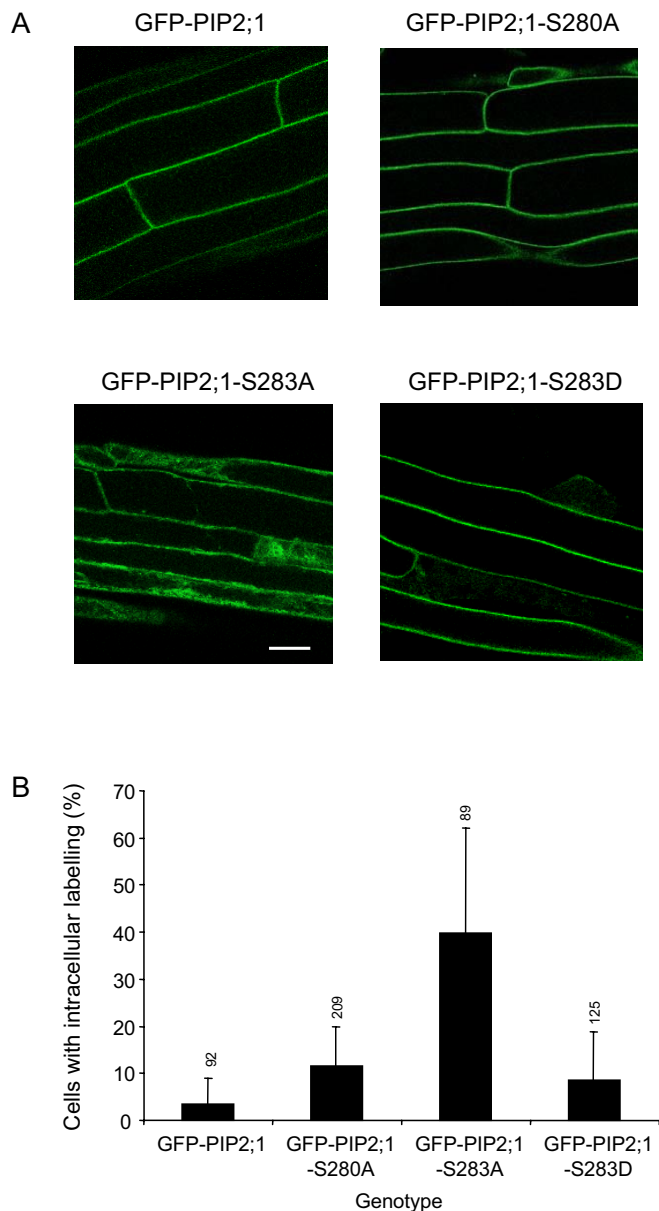


FIG. 7. Role of C-terminal phosphorylation in the subcellular localization of AtPIP2;1. *A*, the figure shows typical laser-scanning confocal micrographs of the fluorescence emitted by root epidermal cells at 1 cm from the apex in transgenic plants that express GFP-PIP2;1, GFP-PIP2;1-S280A, GFP-PIP2;1-S283A, or GFP-PIP2;1-S283D. Scale bar, 20 μm . *B*, the graph represents the proportion of root cells at 1 cm from the apex with an intracellular staining. Data were obtained from transgenic plants that express the following constructs: GFP-PIP2;1 ($n = 13$ plants from two independent transgenic lines), GFP-PIP2;1-S280A ($n = 9$ plants from three independent transgenic lines), GFP-PIP2;1-S283A ($n = 7$ plants from two independent transgenic lines), and GFP-PIP2;1-S283D ($n = 6$ plants from two independent transgenic lines). The numbers on the graph correspond to the total number of observed cells. The error bars represent S.D.

(Fig. 7, *A* and *B*). Altogether these observations indicated that residue Ser²⁸³, and very likely its phosphorylation, is necessary for a proper targeting of AtPIP2;1 to the PM.

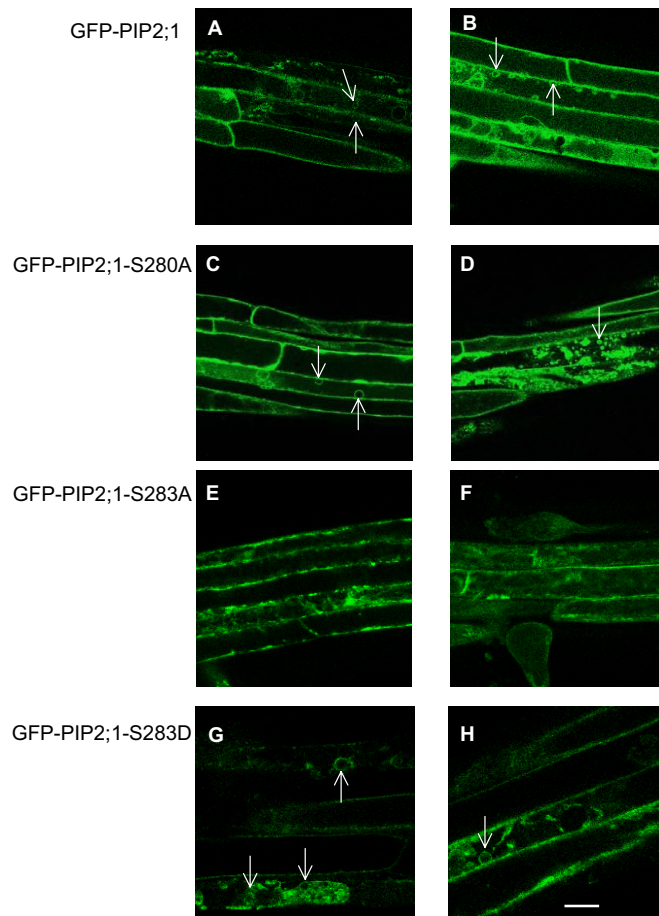
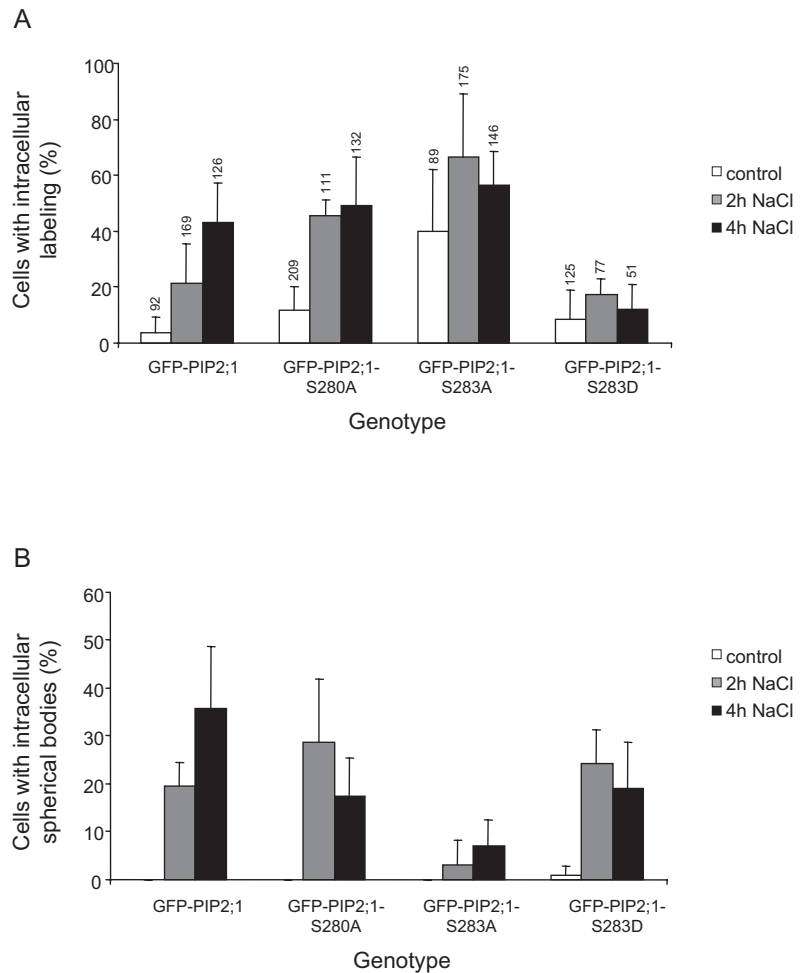


FIG. 8. Role of C-terminal phosphorylation in the subcellular localization of AtPIP2;1 in response to salinity. The figure shows typical laser-scanning confocal micrographs of the fluorescence emitted by transgenic root cells located at 1 cm from the apex and expressing GFP-PIP2;1 (*A* and *B*), GFP-PIP2;1-S280A (*C* and *D*), GFP-PIP2;1-S283A (*E* and *F*), or GFP-PIP2;1-S283D (*G* and *H*). Plants were treated during 2 h (*A*, *C*, *E*, and *G*) or 4 h (*B*, *D*, *F*, and *H*) with 100 mM NaCl prior to microscopic observations. Scale bar, 20 μm .

To investigate the role of C-terminal phosphorylation of AtPIP2;1 in its salt-induced subcellular relocalization, we used the same set of transgenic plants as above. Microscopic observations of root epidermal cells were performed specifically at 1 cm from the apex (Fig. 8). Treatment with 100 mM NaCl during 2 or 4 h induced an intracellular diffuse staining (hereafter referred to as fuzzy staining) in up to 60% of root cells of plants expressing GFP-WT-PIP2;1, GFP-PIP2;1-S280A, and GFP-PIP2;1-Ser283A proteins (Figs. 8, *A–F*, and 9A). A similar staining, but much less abundant, was observed in salt-treated plants expressing PIP2;1-S283D (Figs. 8, *G* and *H*, and 9A). These observations suggested that the accumulation of AtPIP2;1 in fuzzy intracellular compartments observed in response to an NaCl treatment requires a non-phosphorylated form of Ser²⁸³.

A 100 mM NaCl treatment also induced the labeling of small intracellular spherical bodies in up to 35% of root

FIG. 9. Quantification of intracellular fuzzy staining (A) and of spherical bodies (B) in root cells upon an NaCl treatment. The figure represents the proportion of root epidermal cells 1 cm far from the apex that show intracellular fuzzy staining (A) and spherical bodies (B) in control conditions (*white*) and after treatment with 100 mM NaCl during 2 h (*gray*) or 4 h (*black*). Data were obtained from transgenic plants that express the following constructs: GFP-PIP2;1 (two independent transgenic lines; $n \geq 7$ plants per treatment), GFP-PIP2;1-S280A (three independent transgenic lines; $n = 9$ per treatment), GFP-PIP2;1-S283A (two independent transgenic lines; $n \geq 7$ per treatment), and GFP-PIP2;1-S283D (two independent transgenic lines; $n \geq 6$, per treatment). The *numbers* on the graph correspond to the total number of observed cells and are identical in the two figures. The *error bars* represent S.D.



cells of plants expressing GFP-PIP2;1 (Figs. 8, A and B, and 9B). Although these bodies might be related to the endosome/prevacuolar compartment (40),² their precise nature remains uncertain. Salt-treated plants expressing GFP-PIP2;1-S280A and GFP-PIP2;1-S283D showed the same proportion of cells with these spherical structures (Figs. 8, C, D, G, and H, and 9B). By contrast, this type of labeling was almost not visible in root cells expressing GFP-PIP2;1-S283A (Figs. 8, E and F and 9B). These data suggested that residue Ser²⁸³, and very likely its phosphorylated form, is required during the salt-induced relocalization of AtPIP2;1 in intracellular spherical bodies.

DISCUSSION

The present work reports an original proteomics strategy to investigate the phosphorylation of AtPIP aquaporins in the *Arabidopsis* root. Three major steps were involved: (i) an enrichment in aquaporins from purified root PM, (ii) a subsequent enrichment in corresponding phosphopeptides by affinity purification on TiO₂ columns, and (iii) the identification of phosphoresidues by MALDI/TOF-TOF. The present work was focused on the C-terminal tail of AtPIPs, and

overall nine sites were identified in AtPIP2;1 (and/or AtPIP2;2 and AtPIP2;3), AtPIP2;4, and AtPIP2;7. In support for the enhanced resolution of our successive enrichment procedure, we note that six of these sites had not been described previously. AtPIP2;6 recently has been reported to be phosphorylated (25) but is not expressed in roots (12, 31). The present work also shows that bioinformatics predictions of phosphorylation sites cannot be substituted by an experimental identification of sites by MS. For instance, three Ser residues (Ser²⁷³, Ser²⁷⁷, and Ser²⁸⁰) were predicted using NetPhos software to be phosphorylated in the C-terminal tail of AtPIP2;1, whereas the experimental data established the phosphorylation of Ser²⁸⁰ and that of an unpredicted site (Ser²⁸³). Similar discrepancies were observed for AtPIP2;4 and AtPIP2;7 (not shown). Our analysis also identified the phosphorylation of a Thr residue in PIP2;7, a modification that had never been formally described in aquaporins. A remarkable feature of AtPIPs is the presence of multiple, up to three, adjacent phosphorylations on their C-terminal tail. The amino acid sequence alignment of SoPIP2;1, AtPIP2;1, AtPIP2;4, AtPIP2;6, and AtPIP2;7 revealed a correspondence between the first two adjacent

FIG. 10. Sequence alignment of C-terminal phosphorylated plant aquaporins. Arrows indicate residues that can be phosphorylated and that are common between the isoforms.

		↓ ↓	
At PIP2;1	KSKPWDDHWIFWVGPF	IGAAIAAFYHQFVLRASGSKSLG	SFRSAANV---- 287
At PIP2;4	NEKAWDDQWIFWVGPM	IGAAAAAFYHQFILRAAAIKALG	SFGSFGSFRSFA 291
At PIP2;7	NEKAWDDQWIFWVGPF	IGALAAAAYHQYILRASA	IKALGSRSNATN---- 280
So PIP2;1	SNKVWDDQWIFWVGPF	IGAAVAAAAYHQYVLRAAA	IKALGSRSNPTN---- 281
At PIP2;6	NQKAWDDQWIFWVGPF	VGAAIAAFYHQFVLRAGAMKAYG	SVRSQQLHELHA- 289

phosphorylated Ser residues of these proteins (Fig. 10). Whereas independent phosphorylation at n sites yields in theory 2^n peptide forms, we observed a reduced number of phosphorylated forms in AtPIP2;1, AtPIP2;4, and AtPIP2;7. In all three isoforms, phosphorylation of a site was apparently linked to phosphorylation of the closest site, upstream in the peptide sequence, or exceptionally to the second closest site in the case of Ser²⁸⁹ of AtPIP2;4. Interdependent phosphorylation events could result from either a processive or a distributive functioning of the protein kinase along the C-terminal tail (41–43). In the case of PIP2;1, we observed that an S280A mutation did not alter the cellular expression of a GFP-PIP2;1 fusion, whereas an S283A mutation did (Fig. 7). This suggests that phosphorylation of Ser²⁸³ can occur in the GFP-PIP2;1-S280A form in the absence of phosphorylation at position 280. Therefore, we favor a distributivity mechanism whereby a similar protein kinase would act on the two sites with a greater efficiency on Ser²⁸⁰. Similar mechanisms may also be present in animal aquaporins because up to four phosphorylated serine residues have been identified in the C-terminal tail of mammalian aquaporin-2, and a putative interdependency was observed between phosphorylation of two of these sites (Ser²⁵⁶ and Ser²⁶¹) (44).

Although the present study represents a comprehensive analysis of C-terminal phosphorylation of AtPIP2 aquaporins expressed in roots, other aquaporin phosphorylation sites surely remain to be uncovered. AtPIP1 isoforms represent in addition to AtPIP2 abundant proteins in the 28-kDa SDS-PAGE band (31, 33). Although the C-terminal tail of AtPIP1 is not predicted to be phosphorylated, we attempted to detect phosphopeptides derived from this domain. However, the digestion of AtPIP1 isoforms with several different proteolytic enzymes that should release C-terminal AtPIP1 did not allow detection of these domains (data not shown). A phosphorylation site conserved in loop B of all plant PIP isoforms has been postulated based on immunodetection with an anti-phosphopeptide antibody (45) or functional characterization of site-directed mutants in *Xenopus* oocytes (22). Here again, the corresponding native or phosphorylated peptide could not be detected by MS possibly because of low efficiency of the enzyme digestion, the confinement of these peptides into acrylamide after digestion, or also a low ionization efficiency of these peptides. With recent developments in Fourier transform mass spectrometry and the introduction of dissociation modes other than CID, top-down proteomics may be useful to uncover additional phosphorylation sites (46, 47).

An absolute quantification procedure was developed to quantify the relative abundance of the unmodified, singly, and diphosphorylated forms of AtPIP2;1. Plants growing in normal conditions showed a 1:1:2 relative abundance ratio, indicating that AtPIP2;1 is mainly diphosphorylated in the root PM. We also investigated the effects on AtPIP2;1 phosphorylation of NaCl and H₂O₂ treatments, two stimuli known to typically induce a rapid inhibition of L_p in *Arabidopsis* (12).² In these experiments it was of importance to systematically sequence all singly and diphosphorylated forms of AtPIP2;1. Because no phosphorylation of Ser²⁷⁷ was observed in any of the treatments, our quantitative data can truly be interpreted as reversible changes of phosphorylation at Ser²⁸⁰ and Ser²⁸³. NaCl induced a 30% decrease in the level of Ser²⁸³ phosphorylation together with a tendency for an increased relative abundance of the singly and unphosphorylated forms (Fig. 6A). By contrast, an H₂O₂ treatment increased by 20% the relative abundance of the diphosphorylated form and decreased the abundance of the unmodified form (Fig. 6B). Thus, the AtPIP2;1 phosphorylation status appears to be highly sensitive to environmental stimuli acting on root water transport. However, the changes in AtPIP2;1 phosphorylation were not unequivocally associated to changes in L_p . Because they were of modest amplitude, neither one of these changes may be sufficient to account for the strong decrease in L_p induced by the two stimuli. Thus, additional mechanisms including altered phosphorylation of other root aquaporins (AtPIP2;4, AtPIP2;7, or others) or other as yet unidentified regulatory mechanisms may contribute to the decrease in L_p . Phosphorylation at Ser²⁷⁴ of SoPIP2;1, the spinach homologue of AtPIP2;7, was shown to be decreased in leaves under reduced water potential (hyperosmotic treatment) (22). By contrast, phosphorylation of the corresponding residue in AtPIP2;1 (Ser²⁸⁰) was insensitive to NaCl treatment, whereas phosphorylation of Ser²⁸³ was decreased. In nitrogen-fixing nodules of soybean roots, phosphorylation of the aquaporin Nodulin-26 on a C-terminal serine residue (Ser²⁶²) was enhanced upon a water stress (21). These different observations may be explained by differences in the aquaporin isoform and the tissue considered. Recent results on mammalian aquaporin-2 have also revealed reciprocal changes in phosphorylation of two C-terminal serine residues (Ser²⁵⁶ and Ser²⁶¹) in response to vasopressin exposure, suggesting that these residues may serve distinct roles in aquaporin-2 regulation (44, 48). Overall these different studies point to a critical role for aquaporin phosphorylation in response to various physiological contexts and emphasize the need for a global view of aquaporin

phosphorylation dynamics. In these respects, the present study justifies the development of novel, more comprehensive MS-based strategies based on multiple reaction monitoring and/or stable isotope labeling (14).

Functional and structural analyses in spinach SoPIP2;1 have indicated a role for Ser²⁷⁴ in gating the aquaporin (8, 22). By contrast, a possible role for this or the equivalent site in AtPIP2;1 (Ser²⁸⁰) in controlling aquaporin trafficking has remained unexplored. In addition, the functional significance of the adjacent phosphorylation site (Ser²⁸³ in AtPIP2;1) has remained totally unknown. Here we found that this phosphosite was specifically involved in the response of *Arabidopsis* roots to NaCl. Therefore, we focused on the role of the two sites (Ser²⁸⁰ and Ser²⁸³) in AtPIP2;1 trafficking under normal or NaCl stress conditions. For this, we expressed in transgenic *Arabidopsis* GFP-PIP2;1 fusions carrying Ser to Ala mutations to abolish phosphorylation or Ser to Asp mutations to possibly mimic a constitutive phosphorylation. A S280A mutation did not affect the localization profile of the fusion protein when compared with that of wild type GFP-PIP2;1 (Fig. 7). By contrast, an S283A but not an S283D mutation prevented a proper transfer of the protein at the PM (Fig. 7). These results allow us to exclude a detrimental effect of Ser²⁸³ removal and rather indicate that phosphorylation of this residue, but not of Ser²⁸⁰, is necessary for the subcellular trafficking of AtPIP2;1. The perinuclear staining displayed by GFP-PIP2;1-S283A suggests an accumulation in the ER. Therefore, phosphorylation of AtPIP2;1 at Ser²⁸³ seems to favor export of the protein from the ER. A similar model was proposed for mammalian aquaporin-2 whereby phosphorylation of Ser²⁵⁶ by two distinct protein kinases, with different subcellular localizations, mediates the exit of the aquaporin from the Golgi complex and subsequently its translocation from vesicular compartments to the PM (28).

One of the marked effects of an NaCl treatment was to exacerbate the staining by PIP2;1-GFP of diffuse intracellular structures. These intracellular structures were similar to those stained by GFP-PIP2;1-S283A in resting conditions. Mutant analysis showed in addition that the NaCl effects were less pronounced specifically in GFP-PIP2;1-S283D and therefore may be counteracted by phosphorylation of Ser²⁸³ (Fig. 9A). These results suggest that NaCl acts on AtPIP2;1 with unphosphorylated Ser²⁸³ to favor its intracellular accumulation. However, we cannot distinguish at present between (i) an intracellular retention or a misrouting of neosynthesized proteins on their route to the PM and (ii) a relocalization of proteins from the PM into intracellular compartments. NaCl also induced the labeling of intracellular spherical bodies in transgenic plants expressing GFP-PIP2;1. The precise nature of these structures remains to be elucidated, but due to their size (1–5 μ m) and spherical form, they may be related to the prevacuolar/multivesicular body compartment² and devoted to protein degradation. GFP-PIP2;1-S280A and GFP-PIP2;1-S283D, but not GFP-PIP2;1-S283A, labeled the

spherical bodies in NaCl-treated roots (Fig. 9B) suggesting that phosphorylation at position 283 was required for relocalization of AtPIP2;1 in this compartment. In summary, an NaCl treatment induced an intracellular accumulation of AtPIP2;1 by exerting specific actions onto AtPIP2;1 forms differing in their phosphorylation at Ser²⁸³ to induce their accumulation in distinct intracellular structures. It is noteworthy that NaCl also induced dephosphorylation of Ser²⁸³ as was observed on a purified PM fraction. This may reflect a compensatory mechanism to prevent the relocalization of the phosphorylated AtPIP2;1 in spherical bodies and thus to slow down its degradation. Therefore, a fine and reversible tuning of aquaporin density at the cell surface may be achieved.

In conclusion, aquaporin phosphorylation appears to be a significant target in plants under stress. The present study documents salt-induced quantitative changes in aquaporin phosphorylation and establishes, for the first time, a link with aquaporin subcellular localization. Similar links will have to be investigated in contexts such as stress or nutrient responses where information on aquaporin phosphorylation or trafficking has recently emerged (25).²

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