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Variation of heavy metal concentrations (Ag, Cd, Co, Cu, Fe, Pb, V, and Zn) during the life cycle of the common cuttlefish *Sepia officinalis*

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ABSTRACT

The evolution of the concentration of 8 essential and non essential heavy metals (Ag, Cd, Cu, Co, Fe, Pb, V, Zn) in the tissues (digestive gland, cuttlebone and whole animal) of the common cuttlefish *Sepia officinalis* collected in the Bay of Seine has been tracked since the end of the embryogenesis until the reproduction period. Compared to the embryos, the juveniles after hatching display much higher concentrations of Ag, Cu, Fe and Zn suggesting an efficient incorporation from seawater. Conversely, the amounts of Cd, Pb and V in hatchlings remain suggesting that these metals are barely bioavailable for the juveniles. Once the juveniles start to feed, the digestive gland appears to play a major role in the storage of all metals. After only one month of benthic life, the digestive gland already contains up to 90% of the total metal body burden, indicating that digestive gland plays a major role in the storage and presumed detoxification of the selected metals. Metal concentrations in the digestive gland increase in a logarithmic fashion with age during the entire life of cuttlefish, except for Ag which decreases as soon as cuttlefish migrate to open sea. This strongly suggests that (1) Ag is depurated from the digestive gland in relation to supposedly lower exposure in less contaminated environments compare to coastal waters and (2) the digestive gland of cephalopod could be a very good indicator of Ag contamination in the marine environment.

Keywords: Trace element; bioaccumulation; detoxification; cephalopods; sexual maturity; embryogenesis

INTRODUCTION

As other cephalopods, the cuttlefish *Sepia officinalis* has a very short life cycle during which migrations related to growth and reproduction take place. In the English Channel and the Atlantic areas, the reproduction period after which these organisms die, occurs between April and August of their second year of life, when they are aged between 14 and 18 months (Boucaud-Camou et al., 1991; Legoff and Daguzan, 1991). Only some male individuals not attaining their sexual maturity during this period can survive for another year. Therefore, the life of a cuttlefish never exceeds 2 years in those areas (Richard, 1971). During this period, the growth rate of cuttlefish is very high. Their weight can even be multiplied by a factor of 2,000, cuttlefish weighing 0.25 g after hatching and more than 600 g at the sexual maturity

(Richard, 1971; Pascual, 1978; Forsythe et al., 1994). This exceptional growth rate can be explained in terms of their active metabolism owing to their carnivore diet (Mangold, 1989). Regardless of such a short life cycle, the strong capability of cuttlefish to concentrate a large number of metals in their tissues has been previously shown, as well as the major role of the digestive gland in the bioaccumulation processes (e.g. Declair et al., 1978; Miramand and Bentley, 1992; Bustamante et al., 1998; 2002a). This capability seems to be shared by many other species of cephalopods, octopodidae, teuthoidae or nautilidae (Rocca, 1969; Nardi et al., 1971; Schipp and Hevert, 1978; Miramand and Guary, 1980; Smith et al., 1984; Finger and Smith, 1987; Bustamante, 1998; Bustamante et al., 1998; 2000). More recently, radiotracer experimental studies have shown the relative importance of the transfer pathways (water, food, sediments) in the bioaccumulation of 4 heavy metals (Ag, Cd, Co, Zn) in juvenile and adult cuttlefish (Bustamante et al., 2002b; 2004), confirming the storing nature of the digestive gland regardless of the uptake pathway.

However, little is known about the evolution of metal concentrations and burdens during the embryogenesis and growing of *Sepia officinalis*. The aim of this study was therefore to investigate the variation of the concentrations of 8 essential and non essential (Ag, Cd, Cu, Co, Fe, V, Pb, Zn) heavy metals during the growth of the cuttlefish from the embryogenesis to the reproduction period. Eggs, hatched juveniles, juveniles during their 2 first months of benthic life, immature individuals and mature adults returning to the coast to mate were analysed. Thus, both the physiological changes and the migrations have been considered. The former occur during the life cycle of cuttlefish and may modify the bioaccumulation processes of metals, whereas during the migrations the animals move from coastal areas (with metallic inputs of anthropogenic origin) to the open sea (supposed to be less polluted than the coast) hence potentially modifying the metal concentrations in their tissues. In this work, special attention has been paid to the digestive gland owing to its major role in the metabolism of metals. To this end, the zone of study chosen is particularly interesting since direct arrivals of metallic contaminants, especially Ag (RNO, 2001) and Cd (Chiffoleau et al., 1994; 1996; Miramand et al., 2001) are poured by the Seine River. Furthermore, the bay of Seine constitutes a laying and nursery area for the common cuttlefish that is subjected to fishery activities of important economical impact in the region (Boucaud-Camou and Boismery, 1991).

MATERIALS AND METHODS

Biological material

1) Animals from the field

Between July 1989 and July 1991, 153 cuttlefish were collected in 6 times in the bay of Seine off the harbour of Ouistreham (Normandy, France). The age of animals was estimated according to the previous works of Medhioub (1986) and Boucaud-Camou et al. (1991) using the model of seasonal growth established by Pauly and Gaschütz (1979). According to their mantle length (ML), the animals were pooled in estimated age classes for heavy metal analysis as follows:

- about 1-week old: 3 pools of 25 individuals (mean ML = 10 ± 1 mm);
- about 2-weeks old: 4 pools of 8 individuals (mean ML = 18 ± 3 mm);
- about 1-month old: 3 pools of 4 individuals (mean ML = 33 ± 4 mm);
- about 2-months old: 4 pools of 4 individuals (mean ML = 59 ± 6 mm);
- about 12-months old (immature cuttlefish): 3 pools of 5 individuals (mean ML = 133 ± 19 mm);
- about 18-months old (mature cuttlefish): 3 pools of 1 individual (mean ML = 215 ± 5 mm).

The cuttlefish specimens were immediately frozen on board and stored at -20° C. After a short frozen period (< 1 month), each individual was dissected in the laboratory: digestive gland and cuttlebone were removed from the remaining tissues and pooled to allow metal analyses in minute organs and tissues. Our sampling of cuttlefish is supposed to represent a single troop, at least for the individuals of the first year.

2) Eggs reared in the laboratory

Eggs collected in the same area and the same period as the cuttlefish were incubated at the laboratory in aquaria with an open circuit system. 4 pools of 25 eggs were dissected and embryos, yolk and eggshells were separated for analysis. The embryos collected from the eggs were about to hatching and were completely formed but still attached to the yolk sack, which was not completely resorbed. The rest of the eggs were maintained in aquaria until hatching, and the former 25 juvenile hatched (ML = 10 ± 1 mm) were dissected as previously described.

Sample preparation and analytical procedure

All the samples were dried at 60° C for several days to constant weight and then reduced to powder using porcelain mortar and pestle. Aliquots ranging from 10 to 300 mg of the homogenised samples were digested with 4 ml of 14N ultrapur HNO_3 and 1 ml of 22N ultrapur HClO_4 at 100° C on a hot plate during 3 days. After evaporation of the acids, the

residues were taken up in 5 ml 0.3 N HNO₃. Ag, Cd, Co, Cu, Pb and V were analyzed by Zeeman graphite furnace atomic absorption spectrophotometry and Fe and Zn by flame atomic absorption spectrophotometry. Heavy metal concentrations in whole individuals were calculated for reconstructed individuals from analysis of remaining tissues, cuttlebone and digestive gland. However, the high Ca content in the cuttlebone hinder Co measures in this organ, not allowing calculations for the whole juveniles and adults.

Quality control was assayed by heavy metal analyses in blanks and reference materials. Thus, Orchard–Leaves (National Bureau of Standards) and MA-A-1, MA-A-2, respectively copepods and fish flesh standard (IAEA) were treated and analysed in the same way as the samples. Our results for the standard reference materials were in good agreement with the certified values (Table 1). The detection limits were ($\mu\text{g}\cdot\text{g}^{-1}$ dry weight): 0.004 (Cd), 0.02 (Ag), 0.1 (Co, Pb), 0.5 (Cu, V, and Zn) and 2.5 (Fe). Results are also expressed in micrograms per gram of the dry tissue weight ($\mu\text{g}\cdot\text{g}^{-1}$ dwt).

Statistical analysis

Comparison of the metal concentrations between eggshell and embryos/yolk were assessed using *t*-test comparison of means. Differences between metal concentrations at the different ages were tested using the non-parametric Kruskal-Wallis test. Regressions between the concentrations measured in the cuttlefish digestive gland and mantle length were tested using regression procedures for linear and non linear fitted models. Accumulation of metals in the cuttlefish was described by linear and logarithmic fitted models, with the exception of Ag in the digestive gland which is described by a two-order linear component model. Statistical analyses were performed using XLStat Pro 7.0. The level of significance was always set at $\alpha=0.05$.

RESULTS

Levels of concentration and distribution among compartments

The heavy metal concentrations in eggs, embryos, juveniles of different sizes, immature and mature *Sepia officinalis* are given in Table 2.

At the end of the embryo development, the eggs were dissected between eggshell, yolk and embryo. Heavy metal concentrations were similar between yolk and embryo, except for Cd and Cu. However, when compared metal concentrations of those internal compartments with the external one (i.e. the eggshell), results appear contrasted. For Co, Fe, Pb, and V, the

concentrations in the eggshell were significantly ($P_{t-test} < 0.0001$ for the 4 metals) higher than in yolk and embryo whereas they were significantly lower for Ag ($P_{t-test} = 0.015$) and Cu ($P_{t-test} < 0.0001$).

Overall, metal concentrations in the embryos extracted from the eggs were generally lower (Ag, Cu, Fe, and Zn) or in the same range (Cd, Pb and V) than in hatchlings. These concentrations were between 2 and 4 times lower than those measured in mature individuals. During the whole-life cycle of cuttlefish, metal concentrations showed significant variations (H-value ranges from 15.32 to 17.34, and $P_{Kruskal-Wallis}$ -value ranges from 0.004 to 0.009). Overall, the highest metal concentrations correspond to those of the essential metals Cu, Fe and Zn (between 30 and 200 $\mu\text{g}\cdot\text{g}^{-1}$ dwt) and the lowest to non essential metals (between 0.13 and 2.4 $\mu\text{g}\cdot\text{g}^{-1}$ dwt).

The dissection of cuttlefish from 7 days old to 2 years old showed that the digestive gland exhibited about one order of magnitude higher metal concentrations compared to the whole animal (Table 3). Consequently, this organ generally contained a great proportion of the whole-body burden of the metals (Table 4). In contrast, the cuttlebone displayed very low non-essential metal concentrations (Table 5). In this organ, Pb and V concentrations remained close to the detection limit during the whole-life cycle and Ag and Cd concentrations were one order of magnitude lower than those measured in the whole animal. Therefore, the cuttlebone only contained a small fraction of the total non essential metal load (Table 6). Finally, the concentrations of the essential metals measured in the cuttlebone are similar (Fe and Zn) or lower (Cu) than those found in the whole organisms (Table 5).

Variation of metal concentrations and burdens with age

Immediately after hatching, the concentrations of Cu, Fe, and Zn have strongly increased in cuttlefish (Table 2). Indeed, the very young cuttlefish displayed concentrations values of 1.5 to 2 (Cu and Zn) and 6 (Fe) times higher than those found in the embryos taken from the eggs immediately before hatching. However, considering the whole-body concentrations, Cu and Zn increased significantly with age following a logarithmic fashion ($R=0.408$, $ddl=18$, $P<0.05$ and $R=0.865$, $ddl=18$, $P<0.0001$, respectively) whereas Fe tended to decrease significantly ($R=0.657$, $ddl=18$, $P<0.001$). Among toxic elements, Ag was the only metal showing a significant tendency to decrease with age ($R=0.558$ $ddl=18$, $P<0.01$) whereas Cd, Pb and V globally increase (Table 2). However, such variations in the whole-body animals are likely to be due to the variations in the storage organs such as the digestive gland. These tissue-specific variations are actually faked by natural dilution by non-storage organs. Therefore, the tissues

presently dissected at all stages of the life cycle, i.e. the digestive gland and the cuttlebone, were considered to investigate metal variations with age (Figure 1 & 2, respectively).

Regardless of the essential or non essential role of the metals, their concentrations in the digestive gland showed significant variations during the whole-life of cuttlefish (Table 3). The amounts of metals contained in the digestive gland relatively to the whole-body burden also show remarkable changes during the life cycle (Table 4). With the exception of Ag, all metal concentrations and burdens in the digestive gland increase significantly with age following a logarithmic fashion (Figures 1 & 3). Ag concentrations increase linearly ($R=0.812$, $ddl=13$, $P<0.001$) until the age of 2 months (ML = 59 mm), then decrease linearly ($R=0.855$, $ddl=9$, $P<0.001$) when cuttlefish migrate to open ocean waters.

In the cuttlebone, the concentrations of Ag, Cu and Fe decrease with age following a logarithmic fashion ($R=0.861$, $ddl=18$, $P<0.001$, $R=0.910$, $ddl=18$, $P<0.001$, $R=0.660$, $ddl=18$, $P<0.005$, respectively) whereas Zn display a linear accumulation ($R=0.989$, $ddl=18$, $P<0.001$) (Figure 2). The other metals either varied randomly (i.e. Cd) or were below the detection limit (i.e. Pb and V).

Figure 3 shows the variation of metal burdens in relation to the mantle length (age) of the cuttlefish. Both essential and non essential element burdens clearly increase during the whole life cycle of the cuttlefish, with the exception of Ag whose burdens have decreased in mature individuals.

DISCUSSION

Metal bioaccumulation in eggs and hatchlings

At the end of the embryonic development, the embryos showed very low metal concentrations compared to older cuttlefish (Table 2). Such concentrations are close or even identical to those measured in the vitellus of the early spawned eggs, suggesting that the vitellus contains a sufficient amount of essential metals (Cu, Fe, and Zn) necessary for the development of the embryo. Previous studies with radio-labelled metals have demonstrated that ^{65}Zn , ^{109}Cd and ^{57}Co were mainly retained on the eggshell (96% after 11d of exposure), acting as a protective barrier limiting/hindering the incorporation of waterborne metals. However, the eggshell thickness of the eggshell changes during embryonic development. When spawned, its thickness is in the mm-region, i.e. ± 1.5 mm (Lemaire, 1971) and is composed of albumin and other proteins, hardening when put into contact with seawater. Then, it becomes thinner during embryonic development, being almost transparent at the moment of eclosion (Wolf et

al., 1985). Very weak concentrations of Pb and V in the embryos also suggest a specific limitation of their incorporation into the eggs, but this issue should be addressed specifically in the future.

Immediately after hatching, the rapid increase of Cu, Fe and Zn concentrations in cuttlefish tissues (Table 2) suggests that the hatchlings are highly dependent on the essential metals to fulfil their metabolic demands. It therefore follows that the metals are rapidly uptaken once they are in contact with seawater. At this stage, the accumulation only occurs through the dissolved pathways by branchial absorption and tegumental adsorption, the hatchlings still living only on their vitellian reserves for a week (Mangold and Bidder, 1989). High ^{65}Zn accumulation rate has been experimentally shown for juveniles exposed to radiolabelled seawater (Bustamante et al., 2002b). Similarly to essential elements, Ag concentrations are 10 times higher in hatchlings than in the embryos (Table 2), showing a very rapid bioaccumulation of the metal. Similarly to ^{65}Zn , exposure of hatchlings to $^{110\text{m}}\text{Ag}$ via seawater leads to elevated concentration factors (i.e. 320) after a period of only 36h (Bustamante et al., 2004). Therefore, the dissolved pathway appears as a very important route for essential elements and Ag, whereas its role appears rather limited for Cd (Bustamante et al., 2002b), or Pb and V (Miramand et al., 1980, 1981) whose concentrations remained very low. Further investigations on the uptake of V and Pb are specifically needed to assess this hypothesis.

Metal bioaccumulation during growth

After a week of benthic life, the hatchlings use up their vitellian reserves and therefore need to eat. Once they start to feed, cuttlefish rapidly grown (data not shown) but at the same time, are exposed to a new source of metals. Despite dietary intakes, the concentrations of most of the metals in the whole organisms did not vary significantly and only a few metals show a significant increase of the concentrations in the whole cuttlefish (i.e. Cd, Fe, Table 2). This could be due either to 1) control of essential element concentrations through homeostatic regulation, or to 2) dilution of non essential accumulated metals related to very fast somatic growth. With the exception of Fe, the essential metals seem to be well regulated and Cu and Zn showed remarkably homogenous concentrations during the whole life (Table 2). Among non essential elements, Ag concentration is also slightly influenced by dietary supplies. Concentrations of Pb and V in the whole cuttlefish remain very low and will only be over the detection limit only after 1 and 2 months of benthic life, respectively. Therefore, in what concerns the whole animal, cuttlefish bioaccumulate poorly these 2 metals.

Because of the metal dilution with the somatic growth, it was necessary to consider the tissues separately and particularly the digestive gland as a storage organ of metals. With the first feedings, the immature digestive gland of the juveniles starts to evolve to be fully functional at the age of one month (Yim and Boucaud-Camou, 1980). The food intake provokes the beginning of the digestive processes, i.e. secretion of enzymes, absorption of nutrients, and excretion of digestive residues. In a few weeks, the digestive gland takes a dark colour due to the development of digestive cells called “Boules” cells (e.g. Boucaud-Camou, 1973; Boucaud-Camou et al., 1983; Boucher-Rodoni et al., 1987). The development of the digestive gland in the retention process of metals is very important and is reflected by the increased fraction of the whole body burdens of all metals in this organ (Table 4). This process is particularly significant in the case of Fe, for which the metal content stored in the digestive gland raises respectively, from 6 to about 50% before and after the digestive gland development. After only two months of benthic life, the digestive gland contains between 30 (Pb) and 90 % (Ag) of the total amount of metals (Table 4). These contents are identical to those previously measured in adult cuttlefish collected in the same area (Miramand and Bentley, 1992).

Thus, the digestive gland plays a major role in the metabolism of all metals in cuttlefish, and especially Cd (Figure 1). Indeed, during the period of development of the digestive gland (i.e. the first month of benthic life), Cd concentrations are increased 9 times compared to a factor 2 for the remaining metals. During this period, the digestive gland weight increased 30 times and the Cd amounts in this organ were multiplied by 230, i.e. 3 to 5 times higher than for the other metals (Figure 3). The digestive gland has a primarily role in the accumulation of Cd in cuttlefish, as also reported after both field and experimental studies (Miramand and Bentley, 1992; Bustamante, 1998; Bustamante et al., 2002b). Such a role of the digestive gland can be related both to the very efficient detoxification processes of the metal occurring in this organ (Bustamante et al., 2002a) and to the very high Cd assimilation efficiency (*viz.* 62 % for juvenile cuttlefish). Both processes lead to a strong retention of the metal in the digestive gland, which biological half-life ($T_{b1/2}$) exceeds 8 months in juveniles (Bustamante et al., 2002b).

Influence of age

The process of bioaccumulation of all metals except Ag appears as age dependent in the digestive gland (Figure 1). Indeed, the digestive gland weight is increased by 3,500 between hatching and sexual maturity whereas its metal amounts is increased between 6,000 and 9,000

for the essential metals (Cu, Fe, Zn), more than 20,000 (Cd and V) and about 70,000 (Co) (Figure 3). Thus, the digestive gland contains after 2 years of life between 15 and 90 µg of Ag, Co, Pb and V, 165 µg of Cd and between 3 and 7 mg of Cu, Fe and Zn (Figure 3). Therefore, the digestive gland bioaccumulate large concentrations of the metals even those not mostly incorporated through the food pathway, such as for Ag. It is therefore noteworthy that different uptake pathway for metals, i.e. via seawater for Ag and via food for Cd, lead to large concentrations of both toxic metals in the digestive gland. Since the beginning of cuttlefish benthic life, the digestive gland shows 5 to 6 times higher Ag and Cd concentrations compared to those measured in the whole organisms (Tables 2 & 3) and already accounts for 50 to 90% of the total amount of these two metals (Table 4).

Consequently to its storage capacities, all the metals concentrations increased following a logarithmic shape in the digestive gland. Ag is the only exception to this pattern (Figure 1) and its concentrations seem strongly influenced by the migration behaviour of the cephalopods. Indeed, after a few months in coastal areas, juveniles migrate to deeper ocean waters and return to coastal ground to mate only the following year (Richard, 1971). The migration to less polluted area will provoke a decrease in metal concentrations when they have a relatively rapid turn over. In cuttlefish, Ag has short biological half-life either following exposure from seawater or from food (less than 2 wks), as a consequence of its high depuration rates (Bustamante et al., 2004). Our results are likely due to the equilibrium established between contamination (when the animals live by the coast) and decontamination (when the animals live in the open sea) episodes. Therefore, both the elevated bioaccumulation of Ag and its weak retention in the digestive gland of cephalopods will allow the use of these molluscs to monitor Ag concentrations in the marine environment.

The decontamination is actually much slower in the case of other metals as Cd or Co for which the biological $\frac{1}{2}$ life measured in adults after incorporation of Cd or Co by the food pathway is largely higher than the lifetime of cuttlefish (Bustamante et al., 2002b; 2004). Owing to this, it is very likely that since the beginning of the migration to the open sea, the digestive gland of adults is strongly decontaminated in Ag and very little in Co and Cd. Since the main incorporation mechanism of V to the marine organisms seems to be the food pathway (Miramand et al., 1981, Miramand and Fowler, 1998), the behaviour of V is similar to Cd and Co and is strongly retained by cuttlefish. Thus, it would be very interesting to confirm experimentally such hypothesis.

In contrast to the digestive gland, the concentrations measured in the cuttlebone of juveniles are very low, with most of the metals showing concentrations close to those observed in

juvenile just after hatching (Table 2 & 5). The shell in cuttlefish is internal, therefore not exposed to metal adsorption phenomena by direct contact with seawater, unlike other molluscs, especially mussels (Van Weers, 1973; Fowler & Benayoun, 1976; Ünlü & Fowler, 1976; Miramand et al. 1980). As discussed above, albeit the significant penetration of the metals into the tissues, internal transfer of metals to the cuttlebone appear to be very slow and somatic growth of this organ lead to overall dilution of most metals. The only exception was Zn which concentrations significantly increase with age whereas the other analysed elements tended to decrease (Figure 2). Consequently, the metal amounts contained in the cuttlebone correspond in fact either to their weight importance (Cu, Pb, Fe) or are clearly lower (Ag, Cd, V and Zn) (Table 6). This observation is in good agreement with the experimental radiotracer results obtained for Ag, Cd and Zn (Bustamante et al. 2002b, 2004).

Influence of sexual maturation

In two years old individuals (sexually mature), Ag, Cd, Cu and Pb concentrations are lower than those in one-year old animals (immature) against those of Fe, V, and Zn (Table 2). This would be due to physiological changes related to the sexual maturation as well as to the growth of the gonads. These organs exhibit globally low metal levels in cephalopods (Miramand and Bentley, 1992) and contribute to dilute the non-essential metal (Ag, Cd and Pb) concentrations in the whole animals. This is clearly exemplified by the fact that, with the exception of Zn, metal concentrations in the digestive gland do not significantly differ between mature and immature individuals (Table 3). High levels of Zn in the genital tract of the males and females of *Sepia officinalis* (190 ± 22 and $123 \pm 3 \mu\text{g.g}^{-1}$ dwt, respectively) have been reported (Miramand and Bentley, 1992), which would explain that Zn concentrations still increased after sexual maturation. This 1.5-fold increase observed in the whole animals is in all likelihood due to a Zn accumulation in the digestive gland. This value represents a very significant concentration in the mature individuals and is 3 times and even 2 times higher than that measured in the juveniles and in the immature individuals, respectively (Table 3). During this period, the cuttlebone also showed a concentration of Zn higher than that measured in the juveniles and in the one-year old immature individuals. This observation could be translated by an increased flux of Zn in the haemolymph of cuttlefish during this period of sexual maturity. This metal could therefore be partially accumulated by the cuttlebone. In this way, in the animals collected at the end of their sexual maturity the amounts of Zn associated to the cuttlebone (expressed as total Zn % contained in the organisms) are almost 2 times higher compared to the contents calculated in the juveniles and

in the one-year old immatures. Interestingly enough, this phenomenon seems to be limited to Zn. The concentrations of the remaining 7 metals measured in the cuttlebone and the percentages they represent do not vary during this period (Tables 5 & 6).

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REFERENCES

- Boucaud-Camou E. Etude de l'appareil digestif de *Sepia officinalis* L. (Mollusque Céphalopode). Essai d'analyse expérimentale des phénomènes digestifs. PhD Thesis, University of Caen, France. 1973.
- Boucaud-Camou E. Exploitation de la seiche (*Sepia officinalis* L.) en Basse-Normandie. Bulletin de la Société Zoologique de France 1988;113(3):305-310.
- Boucaud-Camou E, Boucher-Rodoni R. Feeding and digestion in cephalopods. In: The Mollusca, Vol 5, Part 2. Saleudin ASM, Wilbur KM editors. New York, London: Academic Press, 1983. p.149-87.
- Boucaud-Camou E, Boismery J. The migration of the cuttlefish *Sepia officinalis* L. in the English Channel. In: "The Cuttlefish" Act. Proceedings of the first international Symposium on the cuttlefish *Sepia*. Boucaud-Camou E editor. Caen: Centre de Publications de l'Université de Caen; 1991. p. 179-89.
- Boucaud-Camou E, Kouéta N, Boismery J, Medhioub A. The sexual cycle of *Sepia officinalis* L. from the bay of Seine. In: "The Cuttlefish" Act. Proceedings of the first international Symposium on the cuttlefish *Sepia*. Boucaud-Camou E editor. Caen: Centre de Publications de l'Université de Caen; 1991. p. 141-51.
- Boucher-Rodoni R, Boucaud-Camou E, Mangold K. Feeding and digestion. In: Cephalopod Life Cycles vol II. Comparative reviews. Boyle PR editor. New York, London: Academic press; 1987. p. 85-108.
- Bustamante P. Etude des processus de bioaccumulation et de détoxification d'éléments traces (métaux lourds et terres rares) chez les mollusques céphalopodes et bivalves pectinidés. Implication de leur biodisponibilité pour le transfert vers les prédateurs. PhD thesis, University of La Rochelle, France; 1998.

- Bustamante P, Chérel Y, Caurant F, Miramand P. Cadmium, copper and zinc in octopuses from Kerguelen Islands, Southern Indian Ocean. *Polar Biology* 1998;19:264-271.
- Bustamante P, Grigioni S, Boucher-Rodoni R, Caurant F, Miramand P. Bioaccumulation of 12 trace elements in the tissues of the nautilus *Nautilus macromphalus* from New Caledonia. *Marine Pollution Bulletin* 2000;40:688-696.
- Bustamante P, Cosson RP, Gallien I, Caurant F, Miramand P. Cadmium detoxification processes in the digestive gland of cephalopods in relation to accumulated cadmium concentrations. *Marine Environmental Research* 2002a;53:227-241.
- Bustamante P, Teyssié JL, Fowler SW, Cotret O, Danis B, Miramand P, Warnau M. Biokinetics of zinc and cadmium accumulation and depuration at different stages in the life cycle of the cuttlefish *Sepia officinalis*. *Marine Ecology Progress Series*, 2002b;231:167-177.
- Bustamante P, Teyssié JL, Danis B, Fowler SW, Miramand P, Cotret O, Warnau M. Uptake, transfer and distribution of silver and cobalt in tissues of the common cuttlefish *Sepia officinalis* at different stages of its life cycle. *Marine Ecology Progress Series* 2004;269:185-195.
- Calabrese A, Collier RS, Nelson DA, MacInnes JR. The toxicity of heavy metals to embryos of the American oyster *Crassostrea virginica*. *Marine Biology* 1973;18:162-166.
- Castillo LV, Maita Y. Isolation and partial characterisation of cadmium binding proteins from the oceanic squid, *Ommastrephes bartrami*. *Bulletin Faculty Fisheries Hokkaido University* 1991;42:26-34.
- Chiffolleau JF, Cossa D, Auger D, Truquet I. Trace metal distribution, partition and fluxes in the Seine estuary (France) in low discharge regime. *Marine Chemistry* 1994;47:145-158.
- Chiffolleau JF, Michel P, Cossa D, Auger D, Averty B, Chartier E, SanJuan J, Truquet I. Distribution des contaminants métalliques dans l'estuaire de Seine en période de crue (Février 1995). *Rapport 1995/fin-3. Programme Scientifique Seine Aval Thème Chimie des Contaminants*, Rouen; 1996.
- Declair W, Vlaeminck A, Geladi P, Van Grieken R. Determination of protein-bound copper and zinc in some organs of the cuttlefish *Sepia officinalis* L.. *Comparative Biochemistry and Physiology* 1978.60B:347-350.
- Denis V, Robin JP. Present status of the French Atlantic fishery for cuttlefish (*Sepia officinalis*). *Fisheries Research* 2001;52:11-22.

Finger JM, Smith JD. Molecular association of Cu, Zn, Cd, and ^{210}Po in the digestive gland of the squid *Nototodarus gouldi*. *Marine Biology* 1987;95:87-91.

Fowler SW, Benayoun G. Accumulation and distribution of selenium in mussel and shrimp tissues. *Bulletin of Environmental Contamination and Toxicology* 1976;16:339-346.

Forsythe JW, DeRusha RH, Hanlon RT. Growth, reproduction and life of *sepia officinalis* (Cephalopoda: Mollusca) cultured through seven consecutive generations. *Journal of Zoology, London* 1994;223:175-192.

Legoff R, Daguzan J. Etude des déplacements de la seiche commune *Sepia officinalis* L. dans le Golfe du Morbihan au cours de la période de reproduction : premiers résultats. In: "The Cuttlefish" Act. Proceedings of the first international Symposium on the cuttlefish *Sepia*. Boucaud-Camou E editor. Caen: Centre de Publications de l'Université de Caen; 1991. p. 167-77.

Lemaire J. Etude du développement embryonnaire de *Sepia officinalis* L. PhD Thesis, University of Lille, France; 1971.

Mangold K. Reproduction, croissance et durée de vie. In: *Traité de Zoologie, Tome V. Céphalopodes*. Grassé PP editor. Paris: Masson; 1989.

Mangold K, Bidder AM. L'appareil digestif et la digestion. In: *Traité de Zoologie, Tome V. Céphalopodes*. Grassé PP editor. Paris: Masson; 1989.

Martin JH, Flegal AR. High copper concentrations in squid livers in association with elevated levels of silver, cadmium and zinc. *Marine Biology* 1975;30:51-55.

Martin M, Osborn KE, Billig P, Glickstein N. Toxicities of ten metals to *Crassostrea gigas* and *Mytilus edulis* embryos and *Cancer magister* larvae. *Marine Pollution Bulletin* 1981;12:183-188.

Medhioub A. Etude de la croissance et du cycle sexuel de la seiche (*Sepia officinalis*) des côtes normandes. PhD Thesis, University of Caen, France; 1986.

Miramand P, Guary JC. High concentrations of some heavy metals in tissues of the Mediterranean *Octopus*. *Bulletin of Environmental Contamination and Toxicology* 1980;24:738-788.

Miramand P, Guary JC, Fowler SW. Vanadium transfer in the mussel *Mytilus galloprovincialis*. *Marine Biology* 1980;56:281-293.

- Miramand P, Guary JC, Fowler SW. Uptake, assimilation and excretion of vanadium in the shrimp *Lysemata seticaudata* (Risso) and the crab *Carcinus maenas* (L.). *Journal of Experimental Marine Biology and Ecology* 1981;49:267-287.
- Miramand P, Bentley D. Concentration and distribution of heavy metals in tissues of two cephalopods, *Eledone cirrhosa* and *Sepia officinalis*, from the French coast of the English Channel. *Marine Biology* 1992;114:407-414.
- Miramand P, Guyot T, Rybarczyk H, Elkaim B, Mouny P, Dauvin JC, Bessineton C. Contamination of the biological compartment in the Seine estuary by Cd, Cu, Pb and Zn. *Estuaries* 2001;24,6B:1056-1065.
- Miramand P, Fowler SW. Bioaccumulation and transfer of vanadium in marine organisms. In: *Vanadium in the Environment Part 1: Chemistry and Biochemistry*. Nriagu JO editor. John Wiley & Sons New York, Vol. 30 in the Wiley Series in Advances in Environmental Sciences and Technology; 1998. p. 167-97.
- Nardi G, Muzii E, Puca M. Ferritin in the hepatopancreas of *Octopus vulgaris* Lam. *Comparative Biochemistry and Physiology* 1971;48B:199-205.
- Pascual E. Crecimiento y alimentación de tres generaciones de *Sepia officinalis* in cultivo. *Investigaciones Pesqueras* 1978;42:412- 442.
- Pauly D, Gaschütz G. A simple method for fitting oscillating growth data, with a program for pocket calculators. *Conseil International pour l'Exploration de la Mer* 1979;CM/G:24-25.
- Richard A. Contribution à l'étude expérimentale de la croissance et de la maturation sexuelle de *Sepia officinalis* L. (Mollusque céphalopode). PhD Thesis, University of Lille, France; 1971.
- RNO. Surveillance du milieu marin, travaux du RNO-Edition 2001. L'argent, le cobalt, le nickel et le vanadium dans les mollusques du littoral français. Paris, Nantes: IFREMER et Ministère de l'Environnement; 2001.
- Rocca E. Copper distribution in *Octopus vulgaris* Lam. Hepatopancreas. *Comparative Biochemistry and Physiology* 1969;28:67-82.
- Schipp R, Hevert F. Distribution of copper and iron in some central organs of *Sepia officinalis* (Cephalopoda). A comparative study by flameless atomic absorption and electron microscopy. *Marine Biology* 1978;47:391-399.

Smith JD, Plues L, Heyraud M, Cherry RD. Concentrations of the elements Ag, Al, Ca, Cd, Cu, Fe, Mg, Mn, Pb and Zn and the radionuclides ²¹⁰Pb and ²¹⁰Po in the digestive gland of the squid *Nototodarus gouldi*. *Marine Environmental Research* 1984;13:55-68.

Tanaka T, Hayashi Y, Ishizawa M. Subcellular distribution and binding of heavy metals in the untreated liver of the squid; comparison with data from the livers of cadmium and silver exposed rats. *Experientia (Basel)* 1983;39:746-748.

Ünlü MY, Fowler SW. Factors affecting the flux of arsenic through the mussel *Mytilus galloprovincialis*. *Marine Biology* 1976;51:209-219.

Van Weers AW. Uptake and loss of ⁶⁵Zn and ⁶⁰Co by the mussel *Mytilus edulis* L. In: *Radioactive contamination of the marine environment*, Vienna: International Atomic Energy Agency; 1973. p. 385-401.

Wolf G, Verheyen E, Vlaeminck A, Lemaire J, Declair W. Respiration of *Sepia officinalis* during embryonic and early juvenile life. *Marine Biology* 1985;90:35-39.

Yim M, Boucaud-Camou E. Etude cytologique du développement post-embryonnaire de la glande digestive de *Sepia officinalis* L (Mollusque, Céphalopode). *Archives d'Anatomie Microscopique et de Morphologie Expérimentale* 1980;69(2):6-79

Table 1 Comparison of elemental concentrations ($\mu\text{g}\cdot\text{g}^{-1}$ dry weight) of Orchard-leaves standard, SRM 1571 (National Bureau of Standards), copepods homogenate, MA-A-1 and fish flesh homogenate, MA-A-2 (International Agency of Atomic Energy) obtained in present study with certified values.

Standard	Ag	Cd	Co	Cu	Fe	Pb	V	Zn
Orchard leaves								
Present study	-	0.10 ± 0.05	0.17 ± 0.04	10 ± 1	272 ± 14	38 ± 2	0.5 ± 0.1	22 ± 6
Certified values	-	0.11 ± 0.02	(0.2)	12 ± 1	300 ± 20	45 ± 3	(0.6)	25 ± 3
MA-A-1								
Present study	0.2 ± 0.1	0.73 ± 0.06	0.12 ± 0.02	6.9 ± 0.4	61 ± 4	2.0 ± 0.5	-	161 ± 1
Certified values	0.33 ± 0.06	0.75 ± 0.03	0.12 ± 0.01	7.6 ± 0.2	60 ± 2	2.1 ± 0.3	-	158 ± 2
MA-A-2								
Present study	0.12 ± 0.01	0.07 ± 0.01	0.09 ± 0.04	3.4 ± 0.7	65 ± 5	0.43 ± 0.14	-	35 ± 4
Certified values	0.10 ± 0.01	0.066 ± 0.004	0.08 ± 0.01	4.0 ± 0.1	54 ± 1	0.58 ± 0.07	-	33 ± 1

() : recommended values

Table 2. Heavy metal concentrations (Mean \pm SD; $\mu\text{g g}^{-1}$ dwt) in whole *Sepia officinalis* sampled in the bay of Seine at different stage of its life cycle. n: number of pool analysed. (): number of individuals in each pool. ML: mantle length; NM: not measured.

Life stage Mantle length (mm)	Estimated age	n	Ag	Cd	Co	Cu	Fe	Pb	V	Zn
Eggs										
Eggshell	-	4 (25)	0.07 ± 0.01	0.59 ± 0.13	1.75 ± 0.26	14 ± 2	398 ± 64	1.38 ± 0.35	3.43 ± 0.59	69 ± 2
Embryos	-	4 (25)	0.16 ± 0.04	0.76 ± 0.17	0.25 ± 0.05	31 ± 1	14 ± 8	< 0.10	< 0.10	68 ± 3
Yolk	-	1 (25)	0.22	0.40	NM	19	11	< 0.10	< 0.10	50
Hatchlings										
ML = 10 mm	0 days	1 (25)	1.80	0.50	NM	60	80	< 0.10	< 0.10	100
Juveniles										
ML = 10 ± 1 mm	7 days	3 (25)	1.82 ± 0.35	0.44 ± 0.14	NM	63 ± 2	107 ± 16	< 0.10	< 0.10	100 ± 3
ML = 18 ± 3 mm	15 days	4 (8)	2.40 ± 0.63	1.32 ± 0.15	NM	77 ± 6	196 ± 51	< 0.10	< 0.10	119 ± 5
ML = 33 ± 4 mm	1 month	3 (4)	1.42 ± 0.20	1.25 ± 0.31	NM	73 ± 3	38 ± 8	< 0.10	< 0.10	113 ± 4
ML = 59 ± 6 mm	2 months	4 (4)	3.04 ± 0.29	0.71 ± 0.09	NM	67 ± 4	23 ± 6	< 0.10	0.6 ± 0.14	113 ± 3
Immature adults										
ML = 133 ± 19 mm	12 months	3 (5)	1.82 ± 0.49	2.03 ± 0.07	NM	104 ± 3	41 ± 9	0.65 ± 0.05	0.47 ± 0.15	145 ± 7
Mature adults										
ML = 215 ± 5 mm	18 months	3 (1)	0.60 ± 0.10	1.10 ± 0.10	NM	70 ± 1	30 ± 5	0.20 ± 0.10	0.50 ± 0.20	156 ± 6

Table 3: Heavy metal concentrations (Mean \pm SD; $\mu\text{g g}^{-1}$ dry wt) in the digestive gland of *Sepia officinalis* sampled in the bay of Seine at different stage of its life cycle. n: number of pool analysed. (): number of individuals in each pool. ML: mantle length.

Mantle length (mm)	Age (estimation)	n	Ag	Cd	Co	Cu	Fe	Pb	V	Zn
Juveniles										
10 \pm 1	7 days	3 (25)	10 \pm 1	2.0 \pm 0.1	0.3 \pm 0.1	175 \pm 5	90 \pm 10	< 0.1	0.3 \pm 0.2	240 \pm 5
18 \pm 3	15 days	4 (8)	18 \pm 7	11 \pm 1	1.3 \pm 0.3	400 \pm 10	160 \pm 10	< 0.1	0.8 \pm 0.1	400 \pm 10
33 \pm 4	1 month	3 (4)	14 \pm 1	15 \pm 3	2.6 \pm 0.5	450 \pm 10	210 \pm 10	0.5 \pm 0.2	1.6 \pm 0.4	500 \pm 10
59 \pm 6	2 months	4 (4)	35 \pm 7	7.5 \pm 0.2	3.5 \pm 0.5	450 \pm 20	110 \pm 10	0.5 \pm 0.2	2.5 \pm 0.7	400 \pm 10
Immature										
133 \pm 19	12 months	3 (5)	19 \pm 7	21.6 \pm 1.5	6.8 \pm 0.9	760 \pm 80	340 \pm 30	2.4 \pm 0.5	2.9 \pm 0.1	770 \pm 40
Mature adults										
215 \pm 5	18 monthss	3 (1)	13 \pm 2	25 \pm 5	10 \pm 2	600 \pm 10	390 \pm 10	2.2 \pm 0.5	3.3 \pm 0.1	1400 \pm 500

Table 4. Percentage distribution (Mean \pm SD; %) of heavy metals in the digestive gland of *Sepia officinalis* sampled in the bay of Seine at different stage of its life cycle. n: number of pool analysed. (): number of individuals in each pool.

Life stage Mantle length (mm)	Estimated age	n	Proportion of whole wt (%)	Ag	Cd	Cu	Fe	Pb	V	Zn
Juveniles										
ML = 10 \pm 1 mm	7 days	3 (25)	10 \pm 2	55 \pm 7	49 \pm 4	29 \pm 6	9 \pm 3	< 10	18 \pm 10	20 \pm 3
ML = 18 \pm 3 mm	15 days	4 (8)	8 \pm 1	61 \pm 8	72 \pm 2	43 \pm 2	7 \pm 2	< 10	17 \pm 5	28 \pm 1
ML = 33 \pm 4 mm	1 month	3 (4)	7 \pm 1	76 \pm 3	88 \pm 5	48 \pm 1	48 \pm 5	< 10	49 \pm 18	34 \pm 2
ML = 59 \pm 6 mm	2 months	4 (4)	8 \pm 1	93 \pm 1	82 \pm 5	57 \pm 4	46 \pm 3	28 \pm 2	38 \pm 19	34 \pm 1
Immature adults										
ML = 133 \pm 19 mm	12 months	3 (5)	8 \pm 1	80 \pm 9	87 \pm 6	59 \pm 6	68 \pm 10	30 \pm 7	54 \pm 20	43 \pm 1
Mature adults										
ML = 215 \pm 5 mm	18 months	3 (1)	3 \pm 1	54 \pm 16	75 \pm 5	42 \pm 8	48 \pm 5	31 \pm 14	20 \pm 15	40 \pm 2

Table 5: Heavy metal concentrations (Mean \pm SD; $\mu\text{g g}^{-1}$ dry wt) in the cuttlebone of *Sepia officinalis* sampled in the bay of Seine at different stage of its life cycle. n: number of pool analysed. (): number of individuals in each pool.

Mantle length (mm)	Age (estimation)	n	Ag	Cd	Cu	Fe	Pb	V	Zn
Hatching		1 (25)	0.03 \pm 0.10	0.08 \pm 0.02	38 \pm 6	21 \pm 2	0.6 \pm 0.3	< 0.5	30 \pm 2
Juveniles									
10 \pm 1	7 days	3 (25)	0.7 \pm 0.2	0.1 \pm 0.1	25 \pm 6	28 \pm 3	< 0.1	< 0.5	75 \pm 5
18 \pm 3	15 days	4 (8)	0.7 \pm 0.2	0.13 \pm 0.02	14 \pm 4	25 \pm 1	< 0.1	< 0.5	75 \pm 5
33 \pm 4	1 month	3 (4)	0.10 \pm 0.02	0.06 \pm 0.01	12 \pm 8	28 \pm 1	< 0.1	< 0.5	75 \pm 5
59 \pm 6	2 months	4 (4)	0.10 \pm 0.02	0.08 \pm 0.01	9 \pm 1	23 \pm 3	< 0.1	< 0.5	80 \pm 5
Immature									
133 \pm 19	12 months	3 (5)	0.08 \pm 0.02	0.22 \pm 0.09	8 \pm 2	15 \pm 3	0.9 \pm 0.1	< 0.5	116 \pm 7
Mature adults									
215 \pm 5	18 monthss	3 (1)	0.08 \pm 0.02	0.03 \pm 0.01	5 \pm 1	25 \pm 2	0.3 \pm 0.2	< 0.5	150 \pm 20

Table 6. Percentage distribution (Mean \pm SD; %) of heavy metals in the cuttlebone of *Sepia officinalis* sampled in the bay of Seine at different stage of its life cycle. n: number of pool analysed. (): number of individuals in each pool.

Mantle length (mm)	Estimated age	n	Proportion of whole wt (%)	Ag	Cd	Cu	Fe	Pb	V	Zn
Hatching		1 (25)	13 \pm 8	4.0 \pm 0.1	1.3 \pm 0.4	14 \pm 7	24 \pm 16	14 \pm 3	< 10	8 \pm 4
Juveniles										
10 \pm 1	7 days	3 (25)	8 \pm 1	3.1 \pm 0.6	1.9 \pm 0.4	3.2 \pm 0.9	1.6 \pm 1.2	< 10	< 10	5.7 \pm 0.6
18 \pm 3	15 days	4 (8)	8 \pm 1	2.8 \pm 1.6	0.8 \pm 0.1	1.7 \pm 0.3	1.2 \pm 0.3	< 10	< 10	5.5 \pm 0.6
33 \pm 4	1 month	3 (4)	8 \pm 1	0.9 \pm 0.6	0.5 \pm 0.3	1.8 \pm 1.2	6.7 \pm 1.5	< 10	< 10	7.3 \pm 1.2
59 \pm 6	2 months	4 (4)	8 \pm 1	0.4 \pm 0.3	1.6 \pm 0.2	1.5 \pm 0.6	13 \pm 3.8	< 10	< 10	10 \pm 1.4
Immature										
133 \pm 19	12 months	3 (5)	14 \pm 1	0.7 \pm 0.4	1.5 \pm 0.6	1.0 \pm 0.3	5.4 \pm 1.6	<10	< 10	11 \pm 1.0
Mature adults										
215 \pm 5	18 months	3 (1)	20 \pm 2	2.2 \pm 0.1	0.7 \pm 0.1	1.3 \pm 1.0	16 \pm 4.0	<10	< 10	19 \pm 4.0

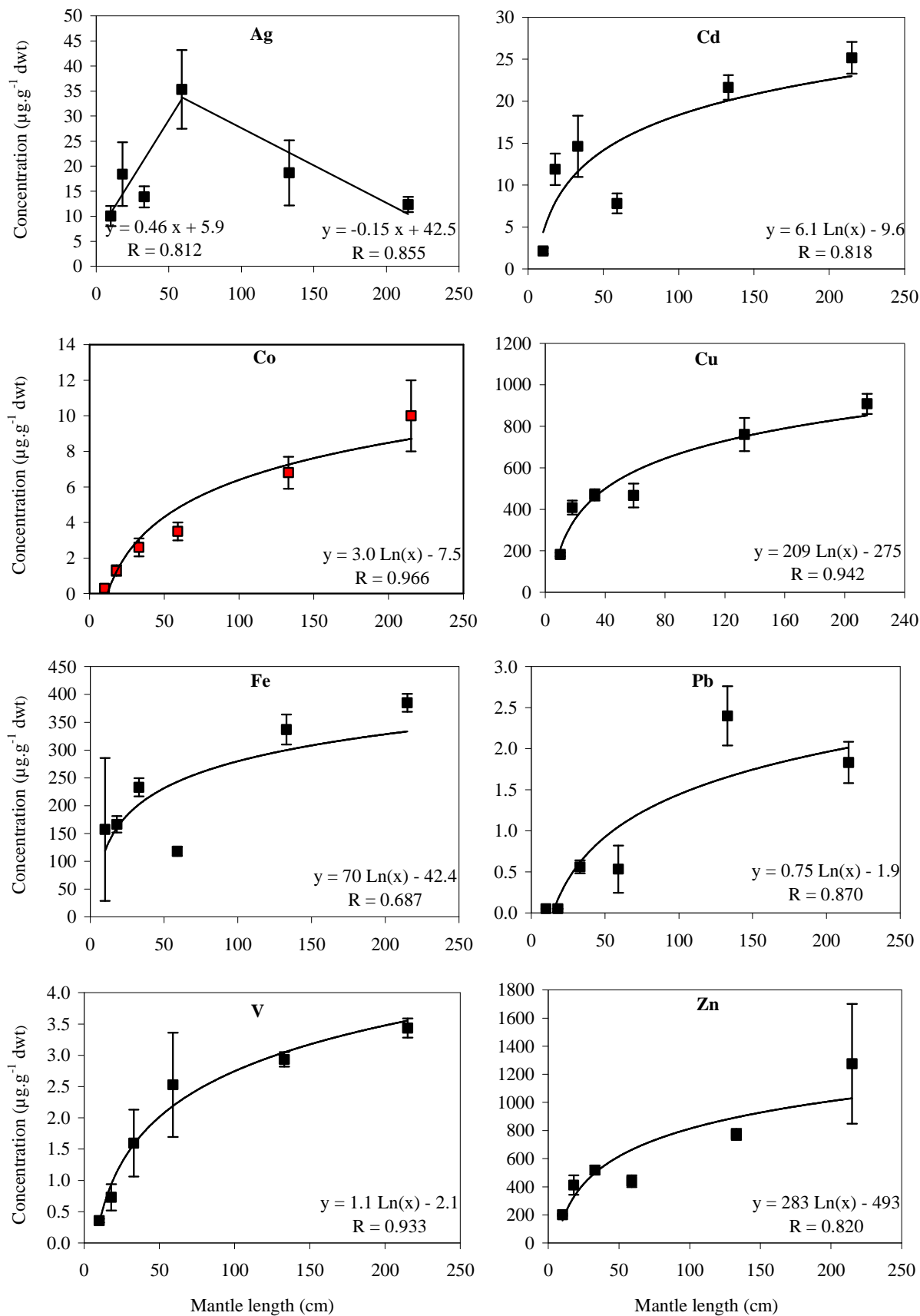


Figure 1. Variations of metal concentrations ($\mu\text{g g}^{-1}$ dry wt) in the digestive gland of *Sepia officinalis* in relation with the mantle length.

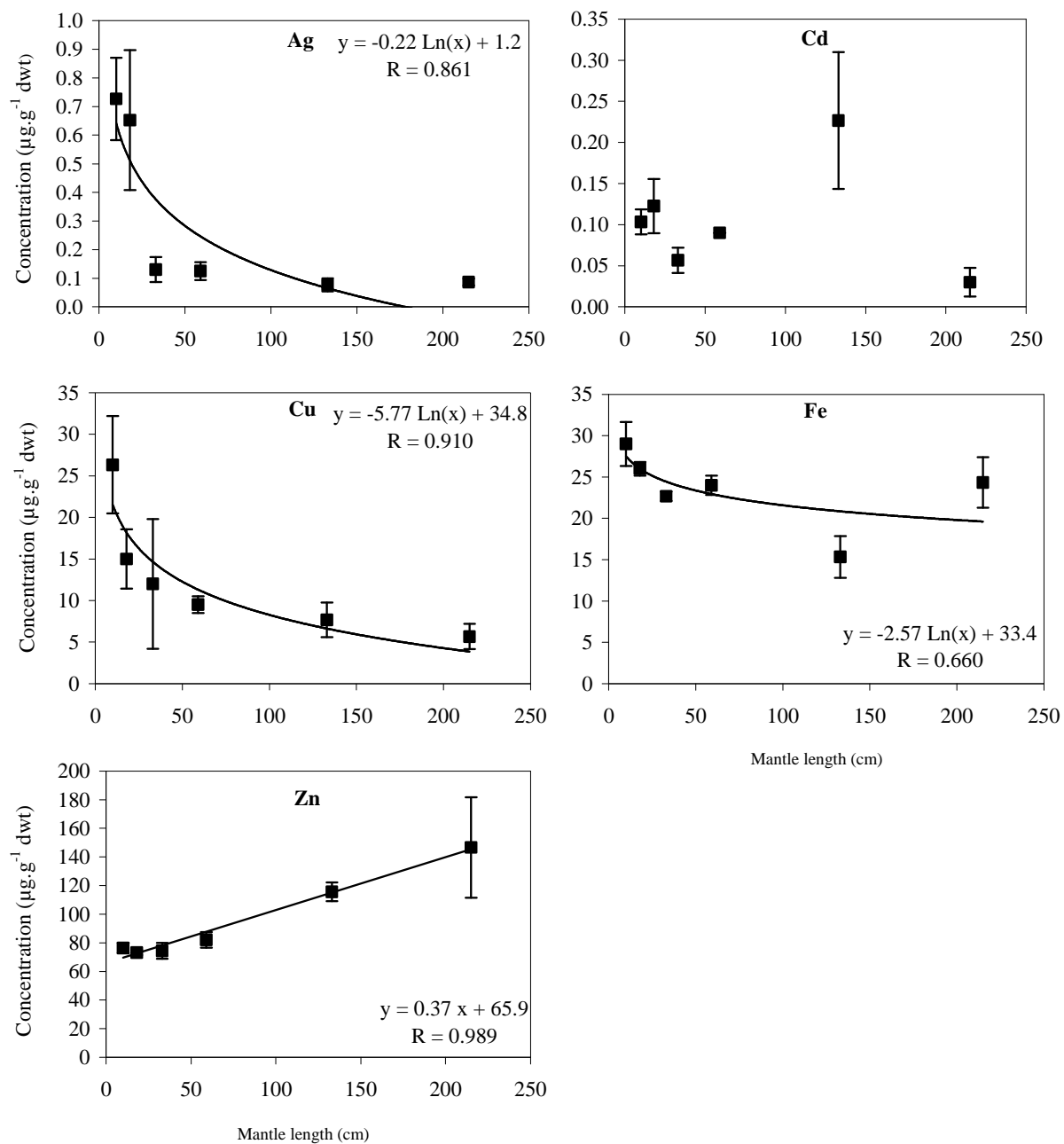


Figure 2. Variations of metal concentrations ($\mu\text{g g}^{-1}$ dry wt) in the cuttlebone of *Sepia officinalis* in relation with the mantle length.

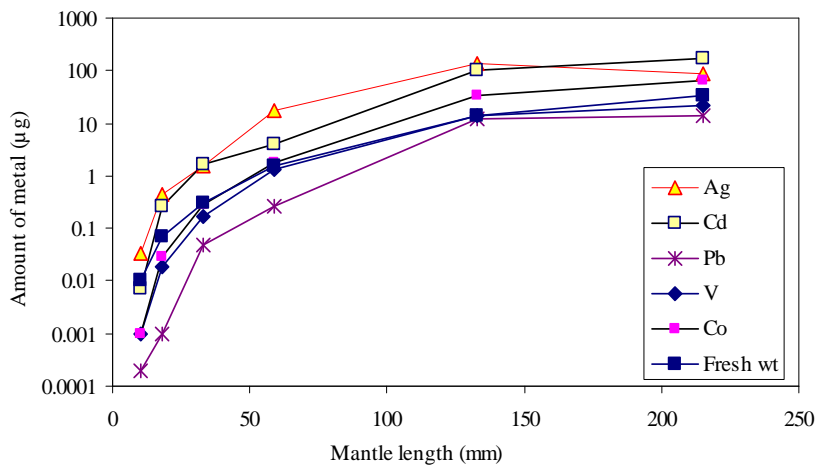
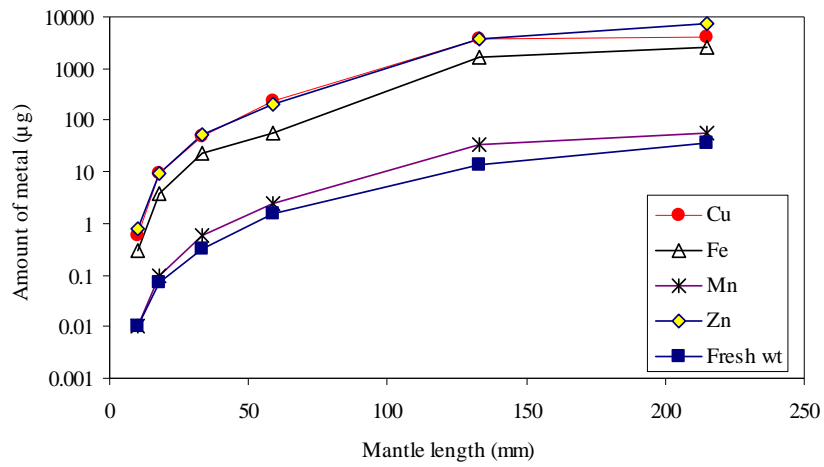


Figure 3. Amounts of metal (μg) contained in the digestive gland of *Sepia officinalis* in relation with the mantle length during the life cycle in the Bay of Seine