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Distribution of trace elements in the tissues of benthic and pelagic fish from the Kerguelen Islands

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Abstract: New information on the concentrations of Cd, Cu, Hg and Zn in the liver, kidney and muscles of 8 marine benthic and pelagic sub-Antarctic fish species are presented to determine the importance of these metals in the marine systems of the Kerguelen Islands. Compared to the reported metal concentrations in other Antarctic fish species, the present results are globally within the same range of concentrations, although Cd displayed a very high interspecific variability in liver and kidney. Indeed, the highest Cd concentrations in liver, ranging from 10.0 to 52.1 $\mu\text{g}\cdot\text{g}^{-1}$ dry wt but also the lowest Cd concentrations in muscles ($< 0.030 \mu\text{g}\cdot\text{g}^{-1}$ dry wt) have been displayed by the pelagic Myctophidae *Gymnoscopelus piabilis*. Metal concentrations differences might be related to diet and feeding habits of benthic and pelagic fish species. However, Cd and Hg concentrations in the edible muscle are lower than the French limit values ($\leq 0.155 \mu\text{g Cd g}^{-1}$ dry wt and $\leq 1.51 \mu\text{g Hg g}^{-1}$ dry wt) for these toxic metals as well as for edible and non-commercially interesting fish species. Results for Cd in fish tissues are consistent with the hypothesis of Cd-enrichment in the polar food webs typically explained by essential elements depletion. In fact, Zn concentrations in fish from the Kerguelen Islands are comparable to those of other areas but low Cu concentrations

in fish livers, ranging from 0.9 to 24.7 $\mu\text{g}\cdot\text{g}^{-1}$ dry wt, might indicate low availability of this essential element in these sub-Antarctic waters.

Keywords : Antarctic ; Heavy metals ; Cd-enrichment ; Myctophidae

INTRODUCTION

Heavy metal concentrations in the organisms from the Austral Ocean have been globally poorly investigated. However, all the studies report high toxic metal concentrations, especially for Cd and Hg, in comparison with those from northern temperate waters. Such metals enrichment [have been reported for polar areas \(AMAP 1998, Petri and Zauke 1993, Sanchez-Hernandez 2000\)](#) but in the Antarctic and sub-Antarctic food webs, it remains unclear even if the extreme environmental conditions (e.g. temperature, seasonal alternation, essential elements availability) might play a key role on the processes of uptake, storage and elimination of the metals by organisms. Moreover, local environmental factors such as volcanism or upwellings could increase metal concentrations in the marine environment.

Specifically, in the Kerguelen Islands environment, high concentrations of Cd have been found to occur in 2 species of benthic octopuses (Bustamante et al. 1998). Generally, cephalopods are known to strongly accumulate Cd in their digestive gland but the concentrations reported for the Kerguelen octopuses were particularly high. Such high Cd levels have also been reported for some Antarctic zooplankton species as a likely result of very low essential elements availability to these organisms (Rainbow 1989, Petri & Zauke 1993). The Cd enrichment in the zooplankton cannot be fully considered as the direct result of anthropogenic contamination owing to the very low levels reported for other zooplankton species living in the same waters (see the data compiled by Sanchez-Hernandez 2000).

Several studies report metal concentrations in the tissues of 13 fish species of various Antarctic areas (Honda et al. 1983, Lenihan et al. 1990, Capelli et al. 1991, Szefer et al. 1993, Minganti et al. 1994, 1995, Bargagli et al. 1996, 1998ab, de Moreno et al 1997, Marquez et al. 1998). However, no data for the fish from the Kerguelen Islands are available to date. This sub-Antarctic Archipelago is of a great ecological interest since millions of seabirds and numerous marine mammals breed there every year (Guinet et al. 1996). Furthermore, as in many sub-Antarctic areas, commercial fisheries developed at the end of the 60s, targeting fish of the families Nototheniidae and Channichthyidae (Duhamel & Hureau 1981). Thus,

baseline information on heavy metal concentrations in the tissue of fish are needed in order to evaluate the fish quality for human consumption as well as for piscivorous predators.

For these reasons, selected heavy metals have been analysed in the liver, kidney and muscles of several fish species, including benthic (neritic) and pelagic (oceanic) ones. Thus, 35 specimens representing 8 different species from the Kerguelen Island waters have been individually analysed for each Cd, Cu, Hg and Zn contents. Metal concentrations and tissue distribution, are compared between species from different Antarctic areas, and influence of the diet of benthic and pelagic species on the accumulation are discussed.

MATERIALS AND METHODS

Sampling and sample preparation

Pelagic fish were collected on cruises of the RV “La Curieuse” during austral summer. Myctophidae and Gempylidae were caught in the eastern part of the peri-insular shelf in February 1998, using a IYGPT trawl (International Young Gadoid Pelagic Trawl, opening 12 X 7 m) with 10 mm mesh size in the cone.

Benthic fish were captured either by net fishing overnight in the Morbihan Bay (*Notothenia rossii* and *Paranotothenia magellanica*) or by commercial trawling on the Kerguelen shelf (*Channichthys rhinoceratus*, *Chamscephalus gunnari* and *Lepidonotothen squamifrons*) (Table 1).

The fish were separated by species and stored on board at -20°C in plastic bags prior to analysis. Subsequently, the length, weight and sex of fish were determined. Moreover, otoliths of Myctophidae species were taken out to ensure identification of the species. Then, specimens were dissected and liver, kidney and muscle tissues were treated separately. The remainders of each dissected individual were also analysed individually in view of determining the percentage distribution of the metals. Characteristics of the samples (i.e. family, species, length, weight and sex) are shown in Table 1.

Analytical procedure

Separated tissue samples were dried to a constant weight for several days at 60°C and then homogenised. Whenever possible, 2 aliquots of approx. 300 mg of each homogenised dry

sample were digested with 5 ml of 65% HNO₃ and 0.3 ml of 70% HClO₄ at 80°C for 24 h. The residues obtained after evaporation of the acids were dissolved in 0.3 N nitric acid. Cd, Cu and Zn were assayed using flame and graphite furnace atomic absorption spectrophotometer Varian 250 Plus with deuterium background correction.

For Hg, aliquots ranging from 10 to 50 mg of dried material have been analysed directly in a Advanced Mercury Analyser spectrophotometer, Altec AMA 254. Hg determination involved evaporation of Hg by progressive heating until 800°C under oxygen atmosphere for 3 min and subsequent amalgamation on a Au-net. Afterwards, the net was heated to liberate the collected mercury and subsequently measured by UV atomic absorption spectrophotometry. However, Hg analysis were not performed in the liver and kidney of the pelagic fish nor for *Lepidonotothen squamifrons*.

Quality assurance was assessed using dogfish liver DOLT-2 (NRCC) and dogfish muscle DORM-2 (NRCC) as reference materials. These standards were treated and analysed under the same conditions as the fish samples, and recoveries of the metals ranged from 92 to 105%. Detection limits, calculated as 3 SD of the mean of eight blanks, were 0.004 for Cd, 0.5 for Cu, 3 for Zn and 0.005 for Hg ($\mu\text{g}\cdot\text{g}^{-1}$ dry wt). All metal concentrations in fish tissues are also reported in $\mu\text{g}\cdot\text{g}^{-1}$ dry wt. Water contents allowing recalculations of the metal concentrations from dry wt to wet wt are given in Table 1.

Statistical procedures

Statistically analysis of results used commercially available packages. As concentration of some elements did not follow a normal distribution, non parametric analysis using Kruskal-Wallis test for multiple comparisons and Mann-Whitney U-test provided by Minitab 13.1 for Windows. A statistically significant difference was considered to exist whenever the probability is lower than $P \leq 0.05$.

RESULTS

Results on heavy metal concentrations in the tissues of fish from the Kerguelen Islands are compiled in Tables 2, 3 and 4 for liver, kidney and muscle, respectively.

In liver, metal concentrations exhibited a large variability among species. For Hg, concentrations differ in 2 orders of magnitude, i.e. between 0.042 and 1.51 $\mu\text{g}\cdot\text{g}^{-1}$ dry wt. The Myctophidae *Gymnoscopelus piabilis* and the Notothenidae *Lepidonotothen squamifrons*

showed significantly higher Cd and Zn concentrations in liver. For Cu and Zn, *Paranotothenia magellanica* also exhibited the highest concentrations. Among the 3 organs considered, liver displays the highest Cd and Cu concentrations. Compared to muscle, hepatic Cd concentrations are 2 to 4 orders of magnitude higher whereas Cu concentrations are only 2 to 10 times higher (Table 2).

Despite the heterogeneity of metal concentrations in liver, no clear segregation between pelagic and benthic fish could be done. This was also the case for kidney. Compared to muscle, kidney shows elevated concentrations of Cd and Cu. Among the 3 tissues, kidney had the highest Zn but the lowest Hg concentrations. As in liver, the Myctophidae *Gymnoscopelus piabilis* and the Notothenidae *Lepidonotothen squamifrons* exhibited significantly higher renal Cd concentrations compared to the other fish species. Renal Cu concentrations were significantly elevated only for *G. piabilis* (Table 3).

Compared to the results for liver and kidney, Cd, Cu and Zn concentrations appeared to be low in fish muscles although the important variability among species is remarkable (Table 4). Muscular Cd and Zn concentrations were generally lower in pelagic fish than in benthic ones. Similarly, mean Hg concentrations varied on 3 orders of magnitude in the muscle (from 0.044 to 1.19 $\mu\text{g}\cdot\text{g}^{-1}$ dry wt) in both benthic and pelagic fish. Nevertheless, the Channichthyidae *Channichthys rhinoceratus* exhibited clearly the highest Hg concentrations (Table 4).

Complete dissection of fish allowed to calculate the distribution of metals between liver, kidney (representing less than 5% and than 1% of the whole body mass) and the remaining tissue including muscle. The percentage of each metal contained in these compartments is shown in Table 5.

Cd and Cu concentrations displayed the same distribution pattern in the tissues of benthic and pelagic fish: liver > kidney > muscle. Despite several orders of magnitude between Cd concentrations in muscle and liver, the latter generally contained less than 50% of the total body burden of Cd. Thus, most of this Cd was muscular except for the Channichthyidae *Channichthys rhinoceratus* (87% of Cd located in liver; Table 5). Owing to the small differences between muscle and liver Cu concentrations and [to the respective proportions of these tissues in the fresh weight of fish](#), less than 20% of the metal was stored in the hepatic compartment (Table 5).

Zn concentrations showed the following sequence : kidney > liver > muscle but the two first organs contained less than 15% of the total body burden of Zn.

Although no clear differences of concentrations between the three tissues appears, the muscular parts of fish contained up to 90% of the total body burden of Hg due to its significance (Table 5).

DISCUSSION

Levels of trace metals

Despite the past and present human fishing pressure on some of the Austral Ocean areas and their major role in the ecosystems (Guinet et al. 1996, Cherel et al. 2000), data on metal levels in the fish still lacks. Such assessment is particularly true for the sub-antarctic zone while Antarctic Ocean is more documented. Thus, this study represent the first investigation concerning the distribution of trace elements in the tissue of 8 fish species from the sub-Antarctic Kerguelen Islands. A recent review of trace element contamination around Antarctica (Sanchez-Hernandez 2000) allows to compare the present results with those of several fish species from the Antarctic Ocean.

Overall, the present results about metal concentrations in the tissues of benthic and pelagic fish from the Kerguelen Islands waters fall within the range for the other fish species from the Antarctic Ocean (data compiled in Table 6-8). Reported data for fish liver (Table 6) range from 0.86 to 21.6 $\mu\text{g Cd g}^{-1}$, from 3.3 to 92 $\mu\text{g Cu g}^{-1}$, from 0.02 to 0.82 $\mu\text{g Hg g}^{-1}$ and from 87.7 to 106 $\mu\text{g Zn g}^{-1}$ (values are expressed in dry wt after conversion from wet wt for some data using a 4.0 correction factor). Generally, the heavy metal levels in fish liver from the Kerguelen Islands tended to be slightly higher than those reported in the current literature (Table 6), except for Hg which is closely in the same concentration range.

Concerning kidney, data for Antarctic fish are rarely reported (Table 7). They range from 0.17 to 10.1 $\mu\text{g Cd g}^{-1}$, and from 0.09 to 2.60 $\mu\text{g Hg g}^{-1}$ (dry wt) while the only values reported for Cu and Zn concern two benthic species, i.e. *Notothenia coriiceps* (Marquez et al. 1998) and *Trematomus bernacchii* (Bargagli et al. 1998b). Despite such few data, it also appears that several fish species from the Kerguelen Islands show higher heavy metal concentrations in the kidney compared to other Antarctic fish species. More fish muscle values from the Antarctic Ocean are available (Table 7). Thus, our results are closely within the same range of the reported data: from 0.01 to 1.0 $\mu\text{g Cd g}^{-1}$, from 0.2 to 2.5 $\mu\text{g Cu g}^{-1}$, from 0.01 to 1.79 $\mu\text{g Hg g}^{-1}$, and from 2.0 to 125 $\mu\text{g Zn g}^{-1}$ (values are expressed in dry wt after conversion from wet wt for some data using a 5.0 correction factor). Considering the French limit value for human consumption of toxic metals, i.e. 0.5 $\mu\text{g.g}^{-1}$ for Cd and 2.5 $\mu\text{g.g}^{-1}$ for Hg (CSHPPF, 1995),

metal concentrations in fish muscles appear to be low, as well for the commercially fished species like *Channichthys rhinoceratus*, *Champsocephalus gunnari* and *Notothenia rossii*, than for the other species presented here.

Heavy metals investigations in fish mainly consider concentrations in muscle and/or in liver, in detriment of gills or kidney as well. Furthermore, little information concerning metal distribution has been reported especially for the Antarctic fish. For this reason, comparison with fish outside the Austral Ocean was necessary and revealed that metal distribution in the tissues of Austral Ocean fish is very similar to that of fish from the temperate waters (Miramand et al. 1991).

Interferences on routes of metal uptake

The very low Cd concentrations in the muscle of most of the fish species investigated (including Myctophidae) suppose effective processes of Cd sequestration in liver and kidney and demonstrate that dietary uptake of Cd is likely to be the most important route of assumption in Kerguelen Islands fish. Enrichment of cadmium in liver and kidney with respect to muscles is particularly evident for zooplankton-eating fish, such as Myctophidae (Hulley 1990). Thus, these species from the Kerguelen Islands displayed the highest Cd concentrations in liver and kidney, i.e. from 10.0 to 52.1 $\mu\text{g.g}^{-1}$ dwt in *Gymnoscopelus piabilis* compared to 0.86 to 21.6 $\mu\text{g.g}^{-1}$ dwt in the liver of the species from other areas (Table 6). The diet of *Gymnoscopelus* consists of crustaceans plankton including euphausiids and euphausiid larvae, hyperiids, and mysids (Hulley 1990). Some of these planktonic species from the Austral Ocean (e.g. euphausiids) exhibit relatively high levels of Cd for crustaceans, with values ranging from 0.15 to 3.4 $\mu\text{g.g}^{-1}$ dwt in the krill *Euphausia superba* (Rainbow 1989, Petri & Zauke 1993). However, some other planktonic species could reach extremely high levels of Cd like the hyperiid amphipod *Themisto gaudichaudii*, which concentrate the metal from 8.0 to 118 $\mu\text{g.g}^{-1}$ dwt (Hennig et al. 1985, Rainbow 1989). Similarly to those reported results, *T. gaudichaudii* show very high Cd concentrations ranging from 21.2 to 81.7 $\mu\text{g.g}^{-1}$ dwt in the Kerguelen Island waters (Bustamante, unpublished data). Thus, the high Cd levels recorded in the kidney and liver of *G. piabilis* would be a direct result of a Cd-rich diet. On the contrary, fish species having the lowest Cd concentrations in their tissues, e.g. *Notothenia rossii* or *Paranotothenia magellanica*, are benthic feeders, and include algae, polychaetes, crustaceans, gastropods and fish in their diet (Gon & Heemstra 1990). However, the

Trematomus fish which are typical benthic feeders of the Antarctic Ocean, exhibit relatively high values reaching 21.6 and 10.1 $\mu\text{g Cd g}^{-1}$ in their liver and kidney, respectively (Tables 6 & 7). This difference can be due to a substantial heterogeneity of age. Thus, these benthic species, as *N. rossii* and *P. magellanica* used in our study were juveniles (less than 4 years old) while *Trematomus* were supposed to be adults after length measurements as suggested by Gon & Heemstra (1990).

Interference on the origin of the metals

Elevated Cd concentrations in the biota from the Austral Ocean were surprisingly high for this area remote from human activities. However, except in some localised spots, anthropogenic Cd contamination does not occur in the Austral Ocean, suggesting that the high concentrations found in the examined marine species are essentially due to natural conditions. Thus, the elevated concentrations of this toxic element in both benthic and pelagic invertebrates seem to correspond to Cd-abnormalities in polar waters as inferred for crustaceans and molluscs (Petri & Zauke 1993, Bargagli et al. 1996, de Moreno et al. 1997, Ritterhoff & Zauke 1997, Bustamante et al. 1998, Sanchez-Hernandez 2000). Indeed, the low Cd concentrations in fish muscle indicate low artificial contamination but the elevated Cd concentrations in both liver and kidney highlight exposure of the fish to the metal. Thus, the high Cd levels in liver and kidney of several Antarctic fish species is a matter of concern.

As it occurs for the Arctic fish (MacDonald & Sprague 1988, Hellou et al. 1992, Zauke et al. 1999), bioaccumulation of Cd in the fish from the Austral Ocean is difficult to explain since the Cd concentrations in the sea water from polar regions are low. Nevertheless, the occurrence of upwelling of deep waters in the Kerguelen region (Plancke 1977) should bring an enrichment of Cd in the surface waters. The increase of Cd bioavailability in the Kerguelen Island waters by upwelled deep waters might be at the origin of an increase of Cd concentrations in the marine biota on a local scale, as discussed for benthic octopus from this area (Bustamante et al. 1998). On the other hand, Bucciarelli et al. (2001) found an iron enrichment of the coastal waters of the Kerguelen Islands and explained it in terms of direct inputs of terrestrial material, as a consequence of riverine discharge, soil leaching by rainwater and aeolian input by strong winds, but also by inputs from the sediments due to effluxes from the sediment at the water interface and resuspension from the sediment. However, our results for Cd but also for Cu, Hg and Zn in the tissue of Kerguelen Islands fish do not show an enrichment following the same processes as Fe. Indeed, Cd concentrations in

both liver and kidney of fish caught in close coastal waters (i.e. *Notothenia rossii* and *Paranotothenia magellanica*) are the lowest among the 8 studied species. Similarly to fish, metal concentrations in crustaceans also distinguish the coastal samples from the continental-shelf ones. For example, the hyperiid amphipod *Themisto gaudichaudii* display Cd concentrations from 21.2 to 27.3 $\mu\text{g}\cdot\text{g}^{-1}$ dwt in the coastal waters (Morbihan Gulf) while 68.2 to 80.7 $\mu\text{g}\cdot\text{g}^{-1}$ dwt were found in the individuals from the shelf (unpublished data). These new findings point out the necessity to investigate carefully the metal concentrations, particularly for Cd, in the waters surrounding Kerguelen Islands, to determine the sources to biota in this area.

Cd enrichment in marine animals from the Austral Ocean was also proposed to occur for organisms suffering essential elements deficiency, which have evolved very efficient mechanisms of elemental uptake (Petri & Zauke 1993). However, these mechanisms are probably non specific to the essential metal and so, Cd might be absorbed by the same pathways than elements like Cu or Zn. Zn deficiency is not evident in the Kerguelen Island waters as its concentrations in fish tissues from this area are of the same order than concentrations reported for fish from various areas (Hellou et al. 1992, Roméo et al. 1999, Zauke et al. 1999). Similarly to fish, Zn concentrations found in the digestive gland of octopus from the Kerguelen Islands are of the same order compared to other cephalopod species (Bustamante et al. 1998). As for Cd, Zn could also be carried by upwelled waters, becoming more available in this area than in open Ocean where upwellings do not occur. Contrarily to Zn, low concentrations of Cu in the liver of fish from the Kerguelen Islands might be due to a low availability of this element. Indeed, Cu concentrations appear to be under close physiological regulation in most species (Thompson 1990). In the same way, low Cu concentrations were also found in octopus from Kerguelen Islands compared to data reported for other cephalopods.

CONCLUSION

The present work provides new information on the distribution of heavy metals in fish from the Kerguelen Islands. Considering the muscle, concentrations of Cd and Hg in both benthic and pelagic species are below the values fixed as a limit by the CSHPF (1995). On the other hand, liver and kidney display very high Cd consequently to high exposure through diet, but very low hepatic Cu concentrations. This is in accordance with the hypothesis of Cd enrichment related to Cu deficiency. In this context, studies on the detoxification and storage

processes of Cd in the liver and kidney of Kerguelen Islands fish should be thoroughly carried out.

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REFERENCES

AMAP. AMAP Assessment Report: Arctic Pollution Issues. Chapter 7: Heavy metals. Arctic Monitoring and Assessment Programme (AMAP), Oslo, Norway, 1998.

Bargagli R, Nelli L, Ancora S, Focardi S. Elevated cadmium accumulation in marine organisms from Terra Nova Bay (Antarctica). *Polar Biol* 1996; 16: 513-520.

Bargagli R, Monaci F, Sanchez-Hernandez JC, Cateni D. Biomagnification of mercury in an Antarctic marine food web. *Mar Ecol Prog Ser* 1998a; 169: 65-76.

Bargagli R, Carsolini S, Fossi MC, Sanchez-Hernandez JC, Focardi S. Antarctic fish *Trematomus bernacchii* as biomonitor of environmental contaminants at Terra Nova Bay Station (Ross Sea). *Mem Natl Insdt PolarRes, Spec Issue* 1998b; 52: 220-229.

Bucciarelli E, Blain S, Tréguer P. Iron and manganese in the wake of the Kerguelen Islands (Southern Ocean). *Mar Chem* 2001; 73: 21-36.

Bustamante P, Cherel Y, Caurant F, Miramand P. Cadmium, copper and zinc in octopuses from Kerguelen Islands, Southern Indian Ocean. *Polar Biol* 1998; 19: 264-271.

Capelli R, Miganti V, Fiorentino F de Pellegrini R. Mercury and selenium in *Adamussium colbecki* and *Pagothenia bernacchii* from the Ross Sea (Antarctica) collected during Italian expeditions 1988-89. *Ann Chim* 1991; 81: 357-369.

Cherel Y, Weimerskirch H, Trouvé C. Food and feeding ecology of the neritic-slope forager black-browed albatross and its relationships with commercial fisheries in Kerguelen waters. *Mar Ecol Prog Ser* 2000; 207: 183-199.

CSHPF (Conseil Supérieur d'Hygiène Publique de France). Plomb, cadmium et mercure dans l'alimentation : évaluation et gestion du risque. Ministère du Travail et des Affaires Sociales (ed). Lavoisier Tec et Doc, Paris, 1995.

Duhamel G, Hureau JC. La situation de la pêche aux Iles Kerguelen en 1981. *Pêche Marit* 1981; 1238: 272-279.

Gon O, Heemstra PC. Fishes of the Southern Ocean. JLB Smith Institute of Ichthyology, Grahamstown, 1990, 462 pp. 12pls.

Guinet C, Cherel Y, Ridoux V, Jouventin P. Consumption of marine resources by seabirds and seals in Crozet and Kerguelen waters: changes in relation to consumer biomass 1962-85. *Antarct Sci* 1996; 8: 23-30.

Hennig HFKO, Eagle GA, McQuaid CD, Rickett LH. Metal concentrations in Antarctic zooplankton species. In: Siefried WR, Condy PR and Laws RM (eds) *Antarctic Nutrient Cycles and Food Webs*. Springer-Verlag Berlin, Heidelberg, 1985, p 656-661.

Hellou J, Warren WG, Payne JF, Belkhode S, Lobel P. Heavy metals and other elements in three tissues of cod *Gadus morhua* from the Northwest Atlantic. *Mar Pollut Bull* 1992; 24: 452-458.

Honda K, Sahrul M, Hikada H, Tatsukawa R. Organ and tissue distribution of heavy metals, and their growth-related changes in Antarctic fish, *Pagothenia borchgrevinki*. *Agric Biol Chem* 1983; 47: 2521-2532.

Hulley PA. Family Myctophidae. In: Gon O & Heemstra PC (eds) *The Fishes of the Southern Ocean*. Grahamstown: JLB Smith Institute of Ichthyology, 1990, p146-178.

Lenihan HS, Oliver JS, Oakden JM, Stephenson MD. Intense and localised benthic marine pollution around McMurdo Station, Antarctica. *Mar Pollut Bull* 1990; 21: 422-430.

Macdonald CR, Sprague JB. Cadmium in marine invertebrates and arctic cod in the Canadian Arctic. Distribution and ecological implications. *Mar Ecol Prog Ser* 1988; 47: 17-30.

Marquez M, Vodopivec C, Casaux R, Curtosi A. Metal (Fe, Zn, Mn and Cu) levels in the Antarctic fish *Notothenia coriiceps*. *Polar Biol* 1998; 20: 404-408.

Miganti V, Fiorentino F, de Pellegrini R, Capelli R. Bioaccumulation of mercury in the Antarctic bony fish *Pagothenia bernacchii*. *Int J Environ Anal Chem* 1994; 55: 197-202.

Miganti V, Capelli R, Fiorentino F, de Pellegrini R, Vacchi M. Variations of mercury and selenium concentrations in *Adamussium colbecki* and *Pagothenia bernacchii* from Terra Nova Bay (Antarctica) during a five years period. *Int J Environ Anal Chem* 1995; 61: 239-248.

Miramand P, Lafaurie M, Fowler SW, Lemaire P, Guary JC, Bentley D. Reproductive cycle and heavy metals in the organs of red mullet, *Mullus barbatus* (L.), from the northwestern Mediterranean. *Sci Tot Environ* 1991; 103: 47-56.

de Moreno JEA, Gerpe MS, Moreno VJ, Vodopivec C. Heavy metals in Antarctic organisms. *Polar Biol* 1997; 17: 131-140.

Petri G, Zauke GP. Trace metal in the crustaceans in the Antarctic Ocean. *Ambio* 1993; 22: 529-536.

Plancke. Phytoplankton biomass and productivity in the Subtropical Convergence area and shelves of the western Indian subantarctic Islands. In : Llano GA (ed) Adaptations within Antarctic ecosystems. Proceedings of the third SCAR Symposium on Antarctic Biology, Washington DC, Aug 26-30, 1974. Smithsonian Institution, Washington, DC, 1977; pp 51-73.

Rainbow PS. Copper, cadmium and zinc concentrations in oceanic amphipod and euphausiid crustaceans, as a source of heavy metals to pelagic seabirds. *Mar Biol* 1989; 103: 513-518.

Ritterhoff J, Zauke GP. Trace metals in field samples of zooplankton from the Fram Strait and the Greenland sea. *Sci Tot Environ* 1997; 199: 255-270.

Roméo M, Siau Y, Sidoumou Z, Gnassia-Barelli M. Heavy metal distribution in different fish species from the Mauritanian coast. *Sci Tot Environ* 1999; 232: 169-175.

Sanchez-Hernandez JC. Trace element contamination in Antarctic ecosystems. *Rev Environ Contam Toxicol* 2000; 166: 83-127.

Santovito G, Irato P, Piccini E, Albergoni V. Relationship between metallothionein and metal contents in red-blooded and white-blooded Antarctic teleosts. *Polar Biol* 2000; 23: 383-391.

Szefer P, Czarnowski W, Pempkowiak J, Holm E. Mercury and major essential elements in seals, penguins, and other representative fauna of the Antarctic. *Arch Environ Contam Toxicol* 1993; 25: 422-427.

Thompson DR. Metal levels in marine invertebrates. In: Furness RW and Rainbow PS (eds). *Heavy metals in the marine environment*. CRC Press, Boca Raton, 1990, p143-182.

Zauke GP, Savinov VM, Ritterhoff J, Savinova T. Heavy metals in fish from the Barents Sea (summer 1994). *Sci Tot Environ* 1999; 227: 161-173.

Table 1. Characteristics of the fish samples, together with the water content in the tissues and organs allowing conversion of dry wt to wet wt metal concentrations.

Family Species	Localisation	Sample size	Length (mm)	Fresh weight (g)	Sex	Water content (%)		
						Liver	Kidney	Muscle
Pelagic fish								
Gempylidae								
<i>Paradiplospinus gracilis</i>	Oceanic zone	1	370	67	♂	55	78	71
Myctophidae								
<i>Gymnoscopelus nicholsi</i>	Oceanic zone	4	144 ± 15	31 ± 8	4 ♀	42 ± 6	45 ± 11	62 ± 2
<i>Gymnoscopelus piabilis</i>	Oceanic zone	5	151 ± 11	37 ± 6	5 ♀	56 ± 8	48 ± 15	76 ± 3
Benthic fish								
Channichthyidae								
<i>Champscephalus gunnari</i>	Kerguelen shelf	5	314 ± 13	202 ± 34	1 ♂, 4 ♀	73 ± 2	73 ± 5	76 ± 2
<i>Channichthys rhinoceratus</i>	Kerguelen shelf	5	344 ± 52	435 ± 150	2 ♂, 3 ♀	75 ± 4	83 ± 2	81 ± 3
Nototheniidae								
<i>Paranotothenia magellanica</i>	Morbihan bay	5	157 ± 8	92 ± 17	2 ♂, 3 ♀	77 ± 2	81 ± 2	78 ± 0
<i>Notothenia rossii</i>	Morbihan bay	5	237 ± 41	300 ± 135	2 ♂, 3 ♀	73 ± 4	81 ± 1	80 ± 1
<i>Lepidonotothen squamifrons</i>	Kerguelen shelf	5	284 ± 29	279 ± 90	3 ♂, 2 ♀	71 ± 8	78 ± 4	76 ± 2

Table 2. Mean \pm SD and range of the metal concentrations ($\mu\text{g}\cdot\text{g}^{-1}$ dry wt.) in the liver of benthic and pelagic fish from the Kerguelen Island waters.

Metal	Species	N	Mean \pm SD	Range	Group			
					1	2	3	
Cd	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	0.94			
		<i>Gymnoscopelus nicholsi</i>	4	4.23 \pm 0.34	3.90-4.66			
		<i>Gymnoscopelus piabilis</i>	5	28.5 \pm 16.9	10.0-52.1			
	Benthic	<i>Notothenia rossii</i>	5	2.82 \pm 1.60	0.82-4.26			
		<i>Paranotothenia magellanica</i>	5	4.15 \pm 2.19	2.03-7.35			
		<i>Channichthys rhinoceratus</i>	5	4.37 \pm 2.00	2.73-6.78			
		<i>Champscephalus gunnari</i>	5	5.52 \pm 4.48	1.04-10.6			
<i>Lepidonotothen squamifrons</i>	5	10.8 \pm 4.59	5.42-15.4					
Cu	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	14.1			
		<i>Gymnoscopelus nicholsi</i>	4	5.8 \pm 1.3	5.1-7.5			
		<i>Gymnoscopelus piabilis</i>	5	10.2 \pm 3.5	6.3-14.8			
	Benthic	<i>Notothenia rossii</i>	5	4.8 \pm 1.6	3.2-7.3			
		<i>Paranotothenia magellanica</i>	5	16.8 \pm 4.5	13.7-24.7			
		<i>Channichthys rhinoceratus</i>	5	4.0 \pm 1.2	3.2-6.1			
		<i>Champscephalus gunnari</i>	5	3.1 \pm 2.5	0.9-7.1			
<i>Lepidonotothen squamifrons</i>	5	3.8 \pm 1.1	2.5-5.6					
Hg	Pelagic	<i>Paradiplospinus gracilis</i>	1	NA	-			
		<i>Gymnoscopelus nicholsi</i>	4	NA	-			
		<i>Gymnoscopelus piabilis</i>	5	NA	-			
	Benthic	<i>Notothenia rossii</i>	5	0.513 \pm 0.208	0.219-0.743			
		<i>Paranotothenia magellanica</i>	5	0.370 \pm 0.114	0.245-0.524			
		<i>Channichthys rhinoceratus</i>	5	0.686 \pm 0.510	0.160-1.51			
		<i>Champscephalus gunnari</i>	5	0.047 \pm 0.006	0.042-0.055			
<i>Lepidonotothen squamifrons</i>	5	0.078 \pm 0.032	0.052-0.133					
Zn	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	112			
		<i>Gymnoscopelus nicholsi</i>	4	92.9 \pm 18.2	70.8-113			
		<i>Gymnoscopelus piabilis</i>	5	142 \pm 30.6	108-184			
	Benthic	<i>Notothenia rossii</i>	5	99.0 \pm 18.9	75.6-119			
		<i>Paranotothenia magellanica</i>	5	143 \pm 16.2	123-165			
		<i>Channichthys rhinoceratus</i>	5	70.4 \pm 13.8	61.4-93.0			
		<i>Champscephalus gunnari</i>	5	62.7 \pm 32.0	28.8-100			
<i>Lepidonotothen squamifrons</i>	5	110 \pm 32.3	63.5-149					

N, independent samples. NA : not analysed. Bars (|) indicates groups identified by non parametric tests.

Table 3. Mean \pm SD and range of the metal concentrations ($\mu\text{g.g}^{-1}$ dry wt.) in the kidney of benthic and pelagic fish from the Kerguelen Island waters.

Metal	Species	N	Mean \pm SD	Range	Group					
					1	2	3	4	5	
Cd	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	1.69					
		<i>Gymnoscopelus nicholsi</i>	4	2.66 \pm 1.03	2.07-4.19					
		<i>Gymnoscopelus piabilis</i>	5	15.7 \pm 8.10	5.92-28.5					
	Benthic	<i>Notothenia rossii</i>	5	0.13 \pm 0.09	0.04-0.27					
		<i>Paranotothenia magellanica</i>	5	0.28 \pm 0.20	0.10-0.59					
		<i>Channichthys rhinoceratus</i>	5	0.31 \pm 0.10	0.20-0.43					
		<i>Champscephalus gunnari</i>	4	2.57 \pm 0.78	1.58-3.31					
<i>Lepidonotothen squamifrons</i>	5	4.28 \pm 1.42	1.99-5.89							
Cu	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	7.2					
		<i>Gymnoscopelus nicholsi</i>	4	4.8 \pm 1.3	3.5-6.4					
		<i>Gymnoscopelus piabilis</i>	5	10.7 \pm 5.4	6.1-19.5					
	Benthic	<i>Notothenia rossii</i>	5	3.7 \pm 0.7	3.0-4.9					
		<i>Paranotothenia magellanica</i>	5	4.4 \pm 0.8	3.7-5.7					
		<i>Channichthys rhinoceratus</i>	5	3.2 \pm 0.7	2.1-3.8					
		<i>Champscephalus gunnari</i>	4	2.2 \pm 0.7	1.2-2.7					
<i>Lepidonotothen squamifrons</i>	5	4.0 \pm 1.0	2.9-4.8							
Hg	Pelagic	<i>Paradiplospinus gracilis</i>	1	NA	-					
		<i>Gymnoscopelus nicholsi</i>	4	NA	-					
		<i>Gymnoscopelus piabilis</i>	5	NA	-					
	Benthic	<i>Lepidonotothen squamifrons</i>	5	NA	-					
		<i>Notothenia rossii</i>	5	0.470 \pm 0.252	0.134-0.721					
		<i>Paranotothenia magellanica</i>	5	0.102 \pm 0.010	0.092-0.119					
		<i>Channichthys rhinoceratus</i>	5	0.533 \pm 0.327	0.172-1.05					
<i>Champscephalus gunnari</i>	4	0.030 \pm 0.006	0.024-0.038							
Zn	Pelagic	<i>Paradiplospinus gracilis</i>	1	-	110					
		<i>Gymnoscopelus nicholsi</i>	4	86.2 \pm 6.8	77.4-93.5					
		<i>Gymnoscopelus piabilis</i>	5	113 \pm 23.1	86.1-146					
	Benthic	<i>Notothenia rossii</i>	5	84.1 \pm 8.8	70.5-92.3					
		<i>Paranotothenia magellanica</i>	5	138 \pm 15.9	122-161					
		<i>Channichthys rhinoceratus</i>	5	143 \pm 38.3	102-202					
		<i>Champscephalus gunnari</i>	4	85.7 \pm 15.2	69.4-106					
<i>Lepidonotothen squamifrons</i>	5	181 \pm 66.4	113-264							

N, independent samples. NA : not analysed. Bars (|) indicates groups identified by non parametric tests.

Table 4. Mean \pm SD and range of the metal concentrations ($\mu\text{g}\cdot\text{g}^{-1}$ dry wt.) in the muscle of benthic and pelagic fish from the Kerguelen Island waters.

Metal	Species	Mean \pm SD	Range	N	Group				
					1	2	3	4	
Cd	Pelagic	<i>Paradiplospinus gracilis</i>	-	0.006	1				
		<i>Gymnoscopelus nicholsi</i>	0.010 \pm 0.008	0.004-0.021	4				
		<i>Gymnoscopelus piabilis</i>	0.016 \pm 0.009	0.006-0.029	5				
	Benthic	<i>Notothenia rossii</i>	0.049 \pm 0.011	0.034-0.064	5				
		<i>Paranotothenia magellanica</i>	0.035 \pm 0.021	0.014-0.063	5				
		<i>Channichthys rhinoceratus</i>	0.051 \pm 0.025	0.024-0.090	5				
		<i>Champscephalus gunnari</i>	0.086 \pm 0.052	0.034-0.155	5				
	<i>Lepidonotothen squamifrons</i>	0.053 \pm 0.026	0.026-0.085	5					
Cu	Pelagic	<i>Paradiplospinus gracilis</i>	-	0.4	1				
		<i>Gymnoscopelus nicholsi</i>	2.5 \pm 0.7	1.9-3.4	4				
		<i>Gymnoscopelus piabilis</i>	1.2 \pm 0.4	0.8-1.7	5				
	Benthic	<i>Notothenia rossii</i>	0.7 \pm 0.1	0.6-0.9	5				
		<i>Paranotothenia magellanica</i>	0.9 \pm 0.0	0.9-1.0	5				
		<i>Channichthys rhinoceratus</i>	0.5 \pm 0.2	0.2-0.8	5				
		<i>Champscephalus gunnari</i>	0.6 \pm 0.1	0.4-0.8	5				
	<i>Lepidonotothen squamifrons</i>	1.0 \pm 0.3	0.7-1.4	5					
Hg	Pelagic	<i>Paradiplospinus gracilis</i>	-	0.251	1				
		<i>Gymnoscopelus nicholsi</i>	0.205 \pm 0.126	0.157-0.297	4				
		<i>Gymnoscopelus piabilis</i>	0.310 \pm 0.126	0.177-0.475	5				
	Benthic	<i>Notothenia rossii</i>	0.255 \pm 0.059	0.192-0.344	5				
		<i>Paranotothenia magellanica</i>	0.140 \pm 0.037	0.097-0.191	5				
		<i>Channichthys rhinoceratus</i>	1.19 \pm 0.367	0.606-1.51	5				
		<i>Champscephalus gunnari</i>	0.044 \pm 0.012	0.034-0.065	5				
	<i>Lepidonotothen squamifrons</i>	0.126 \pm 0.033	0.094-0.180	5					
Zn	Pelagic	<i>Paradiplospinus gracilis</i>	-	10.0	1				
		<i>Gymnoscopelus nicholsi</i>	9.2 \pm 4.0	6.6-15.0	4				
		<i>Gymnoscopelus piabilis</i>	9.9 \pm 1.2	8.4-11.3	5				
	Benthic	<i>Notothenia rossii</i>	19.6 \pm 1.1	18.6-21.0	5				
		<i>Paranotothenia magellanica</i>	22.0 \pm 1.6	20.2-23.7	5				
		<i>Channichthys rhinoceratus</i>	33.2 \pm 7.4	23.6-40.7	5				
		<i>Champscephalus gunnari</i>	29.1 \pm 1.9	27.1-31.1	5				
	<i>Lepidonotothen squamifrons</i>	19.9 \pm 2.6	16.7-22.9	5					

N, independent samples. Bars (|) indicates groups identified by non parametric tests.

Table 5. Distribution of the metals in benthic and pelagic fish from the Kerguelen Islands waters. Mean % \pm SD referred to the fresh weight of organs and tissues.

Metal	Habitat	Species	N	Liver	Kidney	Muscular parts and remainders
Cd	Pelagic	<i>Paradiplospinus gracilis</i>	1	11.7	1.5	86.8
		<i>Gymnoscopelus nicholsi</i>	4	36.1 \pm 13.1	4.0 \pm 1.7	59.9 \pm 13.9
		<i>Gymnoscopelus piabilis</i>	5	47.8 \pm 15.4	5.7 \pm 2.5	46.5 \pm 17.6
	Benthic	<i>Champscephalus gunnari</i>	5	13.8 \pm 12.0	0.6 \pm 0.6	85.6 \pm 12.3
		<i>Channichthys rhinoceratus</i>	5	87.1 \pm 7.5	0.6 \pm 0.4	12.4 \pm 7.2
		<i>Paranotothenia magellanica</i>	5	21.3 \pm 5.2	0.6 \pm 0.3	78.1 \pm 5.7
		<i>Notothenia rossii</i>	5	41.2 \pm 14.2	0.5 \pm 0.3	58.3 \pm 15.0
		<i>Lepidonotothen squamifrons</i>	5	28.7 \pm 11.2	2.1 \pm 1.1	69.2 \pm 12.7
Cu	Pelagic	<i>Paradiplospinus gracilis</i>	1	19.6	0.7	79.7
		<i>Gymnoscopelus nicholsi</i>	4	6.2 \pm 2.5	1.0 \pm 0.6	92.4 \pm 5.7
		<i>Gymnoscopelus piabilis</i>	5	11.3 \pm 3.0	2.3 \pm 0.8	86.5 \pm 3.3
	Benthic	<i>Champscephalus gunnari</i>	5	5.1 \pm 5.0	0.3 \pm 0.3	94.5 \pm 5.3
		<i>Channichthys rhinoceratus</i>	5	7.5 \pm 1.1	0.5 \pm 0.2	92.0 \pm 0.9
		<i>Paranotothenia magellanica</i>	5	10.9 \pm 2.7	1.1 \pm 0.1	87.9 \pm 3.9
		<i>Notothenia rossii</i>	5	6.8 \pm 2.8	1.4 \pm 0.3	91.8 \pm 2.6
		<i>Lepidonotothen squamifrons</i>	5	6.5 \pm 5.2	0.9 \pm 0.1	92.6 \pm 8.8
Hg	Benthic	<i>Champscephalus gunnari</i>	5	2.8 \pm 0.8	0.1 \pm 0.1	97.1 \pm 1.8
		<i>Channichthys rhinoceratus</i>	5	4.3 \pm 1.6	0.4 \pm 0.3	95.4 \pm 1.2
		<i>Paranotothenia magellanica</i>	5	5.6 \pm 3.0	0.6 \pm 0.3	93.8 \pm 4.2
		<i>Notothenia rossii</i>	5	7.1 \pm 2.6	1.7 \pm 0.6	91.2 \pm 2.9
		<i>Lepidonotothen squamifrons</i>	5	1.5 \pm 1.0	ND	98.5 \pm 1.5-
Zn	Pelagic	<i>Paradiplospinus gracilis</i>	1	8.9	0.6	81.5
		<i>Gymnoscopelus nicholsi</i>	4	10.7 \pm 2.0	1.9 \pm 0.9	87.3 \pm 3.9
		<i>Gymnoscopelus piabilis</i>	5	2.3 \pm 5.2	1.9 \pm 0.3	95.9 \pm 5.2
	Benthic	<i>Champscephalus gunnari</i>	5	2.0 \pm 1.1	0.2 \pm 0.2	97.8 \pm 1.4
		<i>Channichthys rhinoceratus</i>	5	3.1 \pm 0.8	0.7 \pm 0.7	96.3 \pm 1.7
		<i>Paranotothenia magellanica</i>	5	3.7 \pm 0.5	1.4 \pm 0.2	95.0 \pm 1.3
		<i>Notothenia rossii</i>	5	6.0 \pm 1.6	1.4 \pm 0.4	92.6 \pm 2.1
		<i>Lepidonotothen squamifrons</i>	5	4.7 \pm 2.7	1.2 \pm 0.2	94.1 \pm 5.0

ND : not determined.

Table 6. Metal concentrations in the liver of fish from the Antarctic Ocean. Values represent the ranges or mean \pm SD, expressed as $\mu\text{g}\cdot\text{g}^{-1}$ dwt or wwt.

Species	Sample area	N	Cd	Cu	Hg	Zn	Basis	Reference
<i>Chaenocephalus aceratus</i>	Antarctic peninsula	4	1.01-1.25	3.3-5.4	-	87.7-106.0	dwt	Szefer et al. (1993)
<i>Chionodraco hamatus</i>	Terra Nova Bay	20	1.02-14.53	-	-	-	dwt	Bargagli et al. (1996)
"	"	18	-	-	0.02-0.44	-	dwt	Bargagli et al. (1998)
"	"	16	0.99 \pm 0.97	0.85 \pm 0.70	-	18.70 \pm 5.13	dwt	Santovito et al. (2000)
<i>Cryodraco antarcticus</i>	"	6	0.86-4.25	-	-	-	dwt	Bargagli et al. (1996)
<i>Notothenia coriiceps</i> (male)	South Shetland Islands	9	-	2.46 \pm 0.84	-	29.55 \pm 8.64	wwt	Marquez et al (1998)
" (female)	"	10	-	1.58 \pm 0.38	-	23.62 \pm 2.20	wwt	Marquez et al (1998)
<i>Pagothenia borchgrevinski</i>	Syowa Station	18	0.30-2.46	0.92-5.88	0.005-0.026	22.4-34.2	wwt	Honda et al. (1983)
<i>Trematomus bernacchii</i>	Terra Nova Bay	18	3.36-21.60	-	-	-	dwt	Bargagli et al. (1996)
"	"	11	-	-	0.10-0.82	-	dwt	Bargagli et al. (1998)
"	"	15	7.30 \pm 5.61	5.19 \pm 3.03	-	44.35 \pm 24.72	dwt	Santovito et al. (2000)
<i>Trematomus hansonii</i>	"	18	5.09-16.42	-	-	-	dwt	Bargagli et al. (1996)
"	"	18	-	-	0.08-0.69	-	dwt	Bargagli et al. (1998)
<i>Trematomus newnesi</i>	"	8	0.98-5.75	-	-	-	dwt	Bargagli et al. (1996)
"	"	7	-	-	0.09-0.28	-	dwt	Bargagli et al. (1998)
<i>Trematomus sp.</i>	Winter Quarters Bay	-	5.0-21.0	5.0-23	-	110-140	wwt	Lenihan et al. (1990)
"	Cinder cones	-	12 \pm 4	14 \pm 2	-	127 \pm 25	wwt	"

Table 7. Metal concentrations in the kidney of fish from the Antarctic Ocean. Values represent the ranges or mean expressed as $\mu\text{g}\cdot\text{g}^{-1}$ dwt or wwt.

Species	Sample area	N	Cd	Cu	Hg	Zn	Basis	Reference
<i>Chionodraco hamatus</i>	Terra Nova Bay	20	0.26-0.87	-	-	-	dwt	Bargagli et al. (1996)
"	"	20	-	-	0.10-0.85	-	dwt	Bargagli et al. (1998a)
<i>Cryodraco antarcticus</i>	"	6	0.18-0.62	-	-	-	dwt	Bargagli et al. (1996)
<i>Notothenia coriiceps</i> (male)	South Shetland Islands	10	-	1.56 ± 0.43	-	23.30 ± 6.63	wwt	Marquez et al (1998)
" (female)	"	10	-	1.72 ± 0.56	-	23.69 ± 10.16	wwt	Marquez et al (1998)
<i>Pagothenia bernacchii</i>	Terra Nova Bay	16	-	-	0.227-2.50	-	dwt	Capelli et al. (1991)
<i>Trematomus bernacchii</i>	Terra Nova Bay	18	2.05-10.11	-	-	-	dwt	Bargagli et al. (1996)
"	"	10	-	-	0.32-2.60	-	dwt	Bargagli et al. (1998a)
"	"	27	1.8-3.0	5.1-5.7	0.47-0.83	110-116	dwt	Bargagli et al. (1998b)
<i>Trematomus hansonii</i>	"	18	1.60-3.29	-	-	-	dwt	Bargagli et al. (1996)
"	"	13	-	-	0.10-1.75	-	dwt	Bargagli et al. (1998a)
<i>Trematomus newnesi</i>	"	8	0.17-3.91	-	-	-	dwt	Bargagli et al. (1996)
"	"	8	-	-	0.09-0.82	-	dwt	Bargagli et al. (1998a)

Table 8. Metal concentrations in the muscle of fish from the Antarctic Ocean. Values represent the ranges or mean expressed as $\mu\text{g}\cdot\text{g}^{-1}$ dwt or wwt.

Species	Sample area	N	Cd	Cu	Hg	Zn	Basis	Reference
<i>Chionodraco hamatus</i>	Terra Nova Bay	20	0.01-0.03	-	-	-	dwt	Bargagli et al. (1996)
"	"	7	-	-	0.01-0.92	-	dwt	Bargagli et al. (1998a)
"	"	10	0.97 ± 0.47	0.70 ± 0.67	-	6.27 ± 3.73	dwt	Santovito et al. (2000)
<i>Chaenocephalus aceratus</i>	Antarctic peninsula	3	0.05-0.13	1.0-2.0	-	28.6-35.5	dwt	Szefer et al. (1993)
<i>Cryodraco antarcticus</i>	Terra Nova Bay	6	0.01-0.06	-	-	-	dwt	Bargagli et al. (1996)
<i>Notothenia coriiceps</i> (male)	South Shetland Islands	10	-	1.00 ± 0.34	-	4.77 ± 1.08	wwt	Marquez et al (1998)
"	South Orkney Islands	11	< 0.05	0.04-0.50	0.01-0.10	1.00-6.70	wwt	de Moreno et al (1997)
" (female)	South Shetland Islands	10	-	4.79 ± 0.81	-	1.01 ± 0.62	wwt	Marquez et al (1998)
"	South Orkney Islands	17	< 0.05	0.05-0.40	0.01-0.09	2.00-5.40	wwt	de Moreno et al (1997)
<i>Notothenia gibberifrons</i>	Antarctica	3	0.02-0.04	0.71-0.98	-	20.7-24.2	dwt	Szefer et al. (1993)
<i>Pagothenia borchgrevinski</i>	Syowa Station	22	0.01-0.04	0.17-1.39	0.002-0.009	4.27-8.15	wwt	Honda et al. (1983)
<i>Pagothenia bernacchii</i>	Terra Nova Bay	18	-	-	0.230-0.990	-	dwt	Minganti et al (1994)
<i>Trematomus bernacchii</i>	Terra Nova Bay	18	0.01-0.08	-	-	-	dwt	Bargagli et al. (1996)
"	"	12	-	-	0.17-1.79	-	dwt	Bargagli et al. (1998a)
"	"	27	0.03-0.05	1.9-2.5	0.49-0.74	22.8-23.6	dwt	Bargagli et al. (1998b)
"	"	10	0.74 ± 0.48	0.32 ± 0.23	-	2.88 ± 1.89	dwt	Santovito et al. (2000)
<i>Trematomus hansonii</i>	"	18	0.01-0.05	-	-	-	dwt	Bargagli et al. (1996)
"	"	15	-	-	0.11-1.08	-	dwt	Bargagli et al. (1998a)
<i>Trematomus newnesi</i>	"	8	0.02-0.05	-	-	-	dwt	Bargagli et al. (1996)
"	"	7	-	-	0.09-0.82	-	dwt	Bargagli et al. (1998a)
<i>Trematomus sp.</i>	Winter Quarters Bay	-	0.1-0.2	0.1-0.3	-	0.4-25	wwt	Lenihan et al. (1990)
"	Cinder cones	-	0.2	0.9 ± 0.8	-	41 ± 13	wwt	"