

Compound-specific isotope analysis. Application to archaelogy, biomedical sciences, biosynthesis, environment, extraterrestrial chemistry, food science, forensic science, humic substances, microbiology, organic geochemistry, soil science and sport.

Eric Lichtfouse

▶ To cite this version:

Eric Lichtfouse. Compound-specific isotope analysis. Application to archaelogy, biomedical sciences, biosynthesis, environment, extraterrestrial chemistry, food science, forensic science, humic substances, microbiology, organic geochemistry, soil science and sport.. Rapid Communications in Mass Spectrometry, 2000, 14 (15), pp.1337 - 1344. 10.1002/1097-0231(20000815)14:153.0.CO;2-B . hal-00174900

HAL Id: hal-00174900 https://hal.science/hal-00174900

Submitted on 25 Sep 2007

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. *Rapid Communications in Mass Spectrometry* Revised version. May 4nd, 2000.

Compound-specific isotope analysis (CSIA). Application to archaeology, biomedical sciences, biosynthesis, environment, extraterrestrial chemistry, food science, forensic science, humic substances, microbiology, organic geochemistry, soil science and sport.

Eric LICHTFOUSE

Soil and Environment Laboratories, INRA-ENSAIA/INPL, BP 172, 54505 Vandoeuvre-les-Nancy, France. <Eric.Lichtfouse@ensaia.inpl-nancy.fr>

ABSTRACT

The isotopic composition, e.g. ¹⁴C/¹²C, ¹³C/¹²C, ²H/¹H, ¹⁵N/¹⁴N and ¹⁸O/¹⁶O, of matter elements is heterogeneous. It is ruled by physical, chemical and biological mechanisms. Isotopes thus be enable to follow the fate of mineral and organic compounds during biogeochemical transformations. The determination of the isotopic composition of organic substances occurring at trace level into very complex mixtures such as sediments, soils and blood, has been made possible during the last 20 years due to the rapid development of molecular level isotopic techniques. After a brief glance at pioneer studies revealing isotopic breakthroughs the molecular and intramolecular levels, this paper reviews selected applications of compound-specific isotope analysis in various scientific fields.

INTRODUCTION

Most stable isotopic research relies on two concepts. First, natural and artificial chemical reactions fractionate isotopes, thus leading to the occurrence of various organic and inorganic materials having different isotopic compositions. Isotopes can thus record biogeochemical changes. Second, isotopes can be used as natural or artificial tracers to follow the behaviour of organic molecules in complex media such as living organisms and ecosystems. The determination of ¹³C/¹²C ratios¹ of natural organic matter has been restricted for a long time to the analysis of bulk samples². Then, the development of gas chromatography coupled to a combustion furnace then to an isotope ratio mass spectrometer³⁻⁵ (GC-C-IRMS, Figure) has allowed the analysis of individual substances occurring at trace levels in very complex mixtures. It thus opened new research fields in various scientific areas⁶⁻⁸ such as organic geochemistry^{9,10}, food science¹¹, medecine¹², nutrition¹³, pharmacy¹⁴, sport¹⁵, phytochemistry^{16,17}, archaeology¹⁸, soil science^{19,20}, environment²¹⁻²⁵, humic substances²⁶⁻²⁸, extraterrestrial science²⁹ and forensic science^{30,31}. ¹⁵N/¹⁴N and ²H/¹H ratios of individual molecules have also been measured using a GC coupled to an IRMS³²⁻³⁵. Besides, since GC is limited to the study of volatile substances, another hyphenated technique has been developed, coupling liquid chromatography to an isotopic ratio mass spectrometer via a combustion furnace (LC-C-IRMS), thus enabling high-molecular, polar, or thermally sensible molecules to be analysed^{36,37}. Further, determination of ²H/¹H and ¹³C/¹²C ratios of each atomic site of a pure substance can be performed by site-specific natural isotope fractionation nuclear magnetic resonance (SNIF-NMR)³⁸⁻⁴⁰. ¹⁴C dating of individual substances occurring in complex mixtures has been recently achieved using preparative-GC to isolate pure lipids⁴¹. Hereafter, in the Early Days section, several examples will show how some pioneers have been able to determine ¹³C/¹²C ratios of individual substances, and even of individual atomic sites, before GC-C-IRMS has been available. Then, selected applications of GC-C-IRMS in various scientific fields will be given.

FIGURE



EARLY DAYS

Before the advent of hyphenated techniques allowing the on-line purification of complex mixtures, off-line isotopic analysis relied on the availability of a pure substance in reasonable amounts, from about 10µg to 10mg, for quartz tube CuO combustion followed by CO₂ transfer into the mass spectrometer. Therefore the analysis of individual compounds occurring in complex media such as living organisms and sediments required careful analytical fractionation to yield pure substances. Here, a milestone has been set by Abelson and Hoering⁴² who isolated individual amino acids from algal and bacterial cultures by several analytical steps including ion-exchange chromatography. They found that amino acids and lipids were respectively ¹³C-enriched and ¹³C-depleted relative to the total organic carbon. Moreover, after ninhydrin decarboxylation of amino acids, they showed that carboxyl groups are strongly enriched in carbon 13 relative to the whole molecule, thus making ¹³C investigations a promising tool to study biosynthesis. This non-statistical isotopic composition at each atomic site has also been shown by NMR⁴³ for algal amino acids grown on ¹³C-CO₂.

Parker measured ¹³C/¹²C ratios of individual fatty acids isolated from algae and grasses by preparative GC^{44,45}. The results show that lipids, e.g. fatty acids, were ¹³C-depleted up to 15‰ relative to the bulk organic carbon. Using the Schmidt decarboxylation⁴⁶, Vogler and Hayes showed that the carboxyl group of fatty acids from corn, soybean and algae is enriched in carbon 1347. These authors have also set up a method to synthesise fatty acids with known isotope value of the carboxyl carbon⁴⁸. Monson and Hayes cleaved the double bonds of unsaturated fatty acids with ozone in order to determine intramolecular isotopic compositions⁴⁹⁻⁵⁰. Meinschein et al. found that the methyl group of acetic acid from cider vinegar is ¹³C-depleted relative to the whole molecule, while commercial acetic acid from inorganic synthesis showed the reverse⁵¹. In close agreement, the carboxyl carbon of acetic acid excreted by the bacterium Acetobacter suboxydans grown on ethanol is ¹³C-enriched by 3.8% relative to the methyl carbon⁵². Rinaldi et al. determined the isotopic composition of individual atomic sites of 3-hydroxy-2-butanone from apple cider vinegar and found that ¹³C/¹²C ratios increase with increasing oxidation level : ketone, alcohol, then methyl⁵³. Remarkably, they write in 1974 that « because the distribution of isotopes within biological compounds are controlled by kinetic and equilibrium effects, intramolecular isotopic analyses may provide a means of defining the equilibria between metabolic pathways and of recognising the molecular remnants of pre-existent organisms in geological samples ».

At a time when the biological origin of petroleum was still a matter of debate, Welte, using preparative GC, measured heterogeneous ¹³C/¹²C values of individual *n*-alkanes from an Eocene crude petroleum, a possible clue to their biological origin^{54,55}. Galimov and Shirinsky reported ¹³C/¹²C values of various natural

organic substances isolated from marine algae, krill zooplancton and lupin plant by thin layer chromatography⁵⁶. These authors proposed a model to calculate intramolecular isotope values. Galimov et al. reported the intramolecular isotope heterogeneity of benzaldehydes, e.g. vanillin, isolated from an Asiatic reed by distillation⁵⁷.

ARCHAEOLOGY

Isotope analysis of individual substances could be a powerful way to bring some light on the origin and on the history of ancient debris, especially when morphological details are absent. For instance, Evershed et al. showed that ¹³C/¹²C ratios of lipids preserved in archaeological potsherds are consistent with those of modern cabbage, thus revealing ancient dietary practices¹⁸. Dudd and Evershed further assessed the use of milk in archaeological economies, using ¹³C/¹²C ratios of fatty acids extracted from pottery vessels of late Saxon and Iron age⁵⁸. Analysing bone cholesterol from fossil whales, Stott et al. recorded ¹³C/¹²C ratios within the range expected for the fat of marine mamals⁵⁹. They also showed a similar depletion between cholesterol and collagen for both modern and ancient whale bones, indicating the reliable preservation of the isotope signal. Further, the recent set up of ¹⁴C dating at the molecular level by Eglinton et al. now enables to date and source individual organic substances occurring in archaeological debris^{60,61}.

BIOMEDICAL

The analysis of stable isotopes at the molecular level for biomedical issues is particularly promising because substances labelled with stable isotopes could ultimately replace the use of radioactive substances. Recent applications of stable isotopes in biomedical fields such as human metabolism¹², nutrition^{12,13}, and pharmacy¹⁴ have been reviewed. Tissot et al. monitored the metabolic fate of ¹³C-glucose in rats and humans by CG-C-IRMS⁶². In a similar way, Khalfallah et al. designed a method to measure plasma lactate after derivatization⁶³. Koziet et al. followed the incorporation of ¹³C-enriched ethanol into human plasmatic triglycerides⁶⁴. Preston and Slater discussed the respective advantages of IRMS, continuous flow (CF)-IRMS, GC-MS and GC-C-IRMS for the use of amino acid as tracers⁶⁵. Yarasheski et al. monitored by GC-C-IRMS the incorporation of ¹³C-leucine into muscle protein of rats⁶⁶. To test the effect of the anti-inflammatory agent isoprofen on acute-phase protein response in patients with colonic cancer, Preston et al. measured the ¹⁵N enrichment of plasma and tissue glycine after administration of ¹⁵N-glycine⁶⁷. Following oral loads of [¹³C-carboxyl]-isoleucine, Meier-Augenstein et al. followed the ¹³C-enrichment of 2-keto-3-methylpentanoic acid in human plasma⁶⁸.

Abnormalities in cholesterol metabolism are implicated in the development of cardiovascular disease in humans. Here, Guo et al. studied by GC-C-IRMS the turnover of human plasma cholesterol following oral administration of [3,4-¹³C]-cholesterol⁶⁹. Deruaz et al. reported a comparison of GC-MS, GC-C-IRMS and GC-atomic emission detector (AED) for the analysis of [3,4-¹³C]-progesterone⁷⁰. The turnover of fatty acid

can also be studied by GC-C-IRMS. For instance, Goodman and Brenna followed the ¹³C-enrichment with time of stearic acid (C_{18}) in human plasma following administration of ¹³C-stearic acid⁷¹. Brossard et al. studied the incorporation of polyunsaturated fatty acids into rat lipoproteins and red cells after ingestion of ¹³C-labelled triglycerides⁷². Similarly, the incorporation of fatty acids in rat brain has been described by Brookes et al.⁷³. Since fatty acids are usually analysed as their methyl ester derivatives, methods to take into account the isotopic value of the added methyl group have been described^{74,75}. In order to measure triacylglycerol synthesis rate in humans, Scrimgeour et al. used GC-Py-IRMS to measure the incorporation of D₂O into plasma fatty acids⁷⁶.

BIOSYNTHESIS

Since ¹³C/¹²C ratios of biological substances depend upon isotope effects associated with metabolism and biosynthesis, as outlined by Hayes⁷⁷, geochemists have studied the extend of isotope fractionation in order to bring some light on the biological sources of sedimentary lipids. Fang et al. reported ¹³C/¹²C ratios of fatty acids from hydrocarbon seep mytilids and vestimentiferan⁷⁸. The results suggest isotope fractionation during fatty acid desaturation and elongation. Investigations of plants using C₃, C₄ and CAM metabolisms⁷⁹ showed differences of ¹³C/¹²C ratios between *n*-alkanes homologues from the same plant^{16,17,80}. Lockheart et al. reported seasonal variations of ¹³C/¹²C ratios of *n*-alkanes from tree leaves⁸¹. Lichtfouse inferred a similar biosynthetic source for long-chain linear alkanes, alcohols and acids in soils based on isotopic signals⁸². Summons et al. found that *n*-fatty acids from the methanotrophic bacterium Methylococcus capsulatus were unexpectedly ¹³C-enriched compared to polyisoprenoids⁸³. Cultures of antartic methanogens revealed isotope effects up to 80% between the substrate and phytanyl ethers⁸⁴. Van der Meer reported ¹³C-enriched values for lipids from the green sulphur bacterium Chlorobium limicola, an organism using the reversed tricarboxylic acid cycle⁸⁵. ²H/¹H isotope variations as large as 150‰ have been observed among isoprenoid lipids from a single organism³⁵. By incubating a sediment with ¹³C-labelled microlalgae, Sun followed fatty acid metabolites by gas chromatography coupled to mass spectrometry (GC-MS)⁸⁶.

ENVIRONMENT

Sherwood Lollar and Abrajano have recently edited a special issue of the journal Organic Geochemistry devoted to the use of compound-specific analysis for environmental issues²¹. Pollution of sediments^{22,87,88} and soils²⁴ by petroleum *n*-alkanes has been inferred using ¹³C/¹²C ratios. In soils, the ancient origin of petroleum alkanes was confirmed by ¹⁴C-dating²⁴. Lichtfouse and Eglinton reconstructed the fingerprint of the petroleum contamination using a two end-member mixing model²⁴. Lichtfouse et al. further showed that short-chain *n*-alkanes from crop soils represent a fossil fuel contamination because these compounds did not show any ¹³C/¹²C variation during a 23-year experiment of labelling of soil carbon by maize cultivation⁸⁹.

The occurrence of polycyclic aromatic hydrocarbons (PAHs) in the modern environment^{90,91} has focused interest because these compounds could exhibit toxicological activity. O'Malley et al. reported ¹³C/¹²C distinct values for PAHs from various environmental sources such as carsoot, fireplace, sewage and harbour sediments²³. The production of PAHs by biomass burning did not show significant isotopic fractionation, thus enabling the use of isotope analysis to source sedimentary PAHs derived from fires⁹². Ballentine et al. reported ¹³C/¹²C values of PAHs and fatty acids from aerosols collected during a field burn of sugar cane⁹³. McRae et al. inferred the fossil fuel sources of PAHs from soils and vegetation sampled close to a carbonisation plant⁹⁴. Lichtfouse et al. assessed the fossil fuel origin of PAHs in crop soils using ¹³C-labelling with maize, ¹⁴C-dating and biomarker evidence⁹⁵. Richnow et al. monitored the incorporation of [9-¹³C]-anthracene and its metabolites into soil humus⁹⁶. In an isotopic survey of commercial benzene, toluene, ethylbenzene and xylenes, Harrigton et al. observed a wide range of ¹³C/¹²C values, suggesting applications to qualify and to quantify subsurface processes affecting contaminant concentrations⁹⁷.

Zeng et al. applied GC-C-IRMS to measure ¹³C/¹²C ratios of greenhouse gases from biomass burning and exhausts⁹⁹. Biodegradation of petroleum hydrocarbons has been monitored using ¹³C, ¹⁴C and ²D contents of CH₄ and CO₂^{100,101}. Rowe and Muehlenbachs proposed an isotopic means to assess the contamination of shallow aquifers by petroleum gases (C₁-C₄) migrating from deeper leaking wells¹⁰². Dayan et al. studied the isotopic fractionation of chlorinated ethenes during reductive dehalogenation by metallic iron, a recently developed remediation technology¹⁰³. As shown by Heraty et al., ¹³C/¹²C investigations of chorinated hydrocarbons, e.g. dichloromethane, could lead to applications in the characterisation and remediation of contaminated sites¹⁰⁴. For instance, Sherwood Lollar et al. recorded a ¹³C increase from -30% to -16% during the anaerobic biodegradation of trichloroethylene¹⁰⁵. A similar trend was observed by Stehmeier et al. during benzene degradation¹⁰⁶. Bergamaschi et al. found that trihalomethanes formed by reaction of chlorine with plant leachates are ¹³C-depleted of 12% relative to the whole plant material¹⁰⁷. Further environmental applications will benefit from the set up by Dias and Freeman of GC-C-IRMS analysis of aqueous volatile organic compounds using solid-phase microextraction (SPME)¹⁰⁸.

EXTRATERRESTRIAL CHEMISTRY

A major issue in the analysis of extraterrestrial material such as meteorites is the contamination of samples by terrestrial matter. Here, isotopic analysis is particularly relevant because extraterrestrial compounds have usually sharply different isotopic signatures compared to earth substances. For instance, Epstein et al. found unusually high ²H/¹H and ¹⁵N/¹⁴N ratios in amino acid and monocarboxylic acid fractions from the Murchison meteorite, thus confirming the extraterrestrial origin of both classes of compounds¹⁰⁹. Engel et al.²⁹ further measured by GC-C-IRMS the ¹³C/¹²C ratios of individual amino acids¹¹⁰ from the same meteorite. Alanine being not racemic in meteorites, the ¹³C-enrichment of its D- and L-enantiomers^{110,111} implies that the excess of the L-enantiomer is indigenous rather than from terrestrial

contamination. These results suggest that optically active materials were present in the early Solar System before life began. Such findings bring some light on the question of why living systems on Earth came to use almost exclusively left-handed amino acids (L-enantiomers).

FOOD SCIENCE

Reviews on carbon isotopes in food technology¹¹², NMR isotopic analysis¹¹³, GC studies of flavours and fragrances¹¹⁴ and authentication¹¹⁵ have been published. Isotopes are mainly used in the food science area to distinguish natural versus synthetic origin of ingredients. Indeed, cheap, synthetic, petroleumderived compounds have usually different isotope values than their costly, natural counterparts. Several studies highlight the use of both chirospecific analysis and ¹³C/¹²C determination by CG-C-IRMS for the authentication of fruit lactones^{11,116-119}. Schmidt reported ¹³C/¹²C values of various synthetic and natural compounds such as carvone, almond benzaldehyde, and cinnamon aldehyde¹²⁰. Breaking the molecules into pieces prior isotope analysis, these authors also showed that intramolecular isotope values are heterogeneous. For instance, the aglycone moieties of glycosides is ¹³C-depleted relative to the carbohydrate counterpart, as a consequence of different biosynthetic pathways. Braunsdorf et al. further studied ¹³C/¹²C values of flavour compounds from fruit oil, e.g. limonene and octanal, in relation to their metabolic pathways^{121,122}. Breas et al. highlighted the value of GC-C-IRMS for the rapid and convenient analysis of vanillin and mint hexenol¹²³. Karl et al. performed chirospecific measurements of isotope ratios of 2-methylbutanoates from apple flavours¹²⁴.

Site specific natural isotope fractionation measured by nuclear magnetic resonance (SNIF-NMR) allows to determine isotope ratios at atomic sites of a substance^{38-40,125-126}. This ability can be particularly useful for the authentication of synthetic and natural substances having close or similar molecular isotope values. Indeed, though the molecular isotope value of two substances of similar structure may be equal, their intramolecular distribution of isotopes can be different, owing to their different biosynthetic sources. Danho et al. the reported intramolecular D/H distribution of caffeine from both synthetic and natural origins¹²⁷. Hanneguelle et al. reported strikingly different intramolecular D/H values of linalool from plants and fossil materials¹²⁸. Martin et al. showed that D/H of wine ethanol can be used to sort out varieties according to production and vintage¹²⁹. Martin et al. further carried a systematic investigation of the water cycle in the wine ecosystem¹³⁰.

FORENSIC SCIENCE

Isotopic analysis can bring information on the geographical origin of drugs^{30,31,131-133}. For instance, bearing in mind that heroin is prepared by acetylation of natural morphine, Besacier et al. proposed GC-C-IRMS analysis to locate the geographical origin of heroin and of the acetic anhydride used by the drug trafficker³¹.

HUMIC SUBSTANCES

Humic substances are complex, partly macromolecular, yellow-brownish organic materials produced during biomass decomposition in soils, in waters and in sediments. Their precise sources, their molecular structure and their transformation pathways are still poorly known¹³⁴. Therefore ¹³C analysis and molecular characterisation is a promising approach to study this organic puzzle. By isotopic comparison of various organic components, e.g. linear alkanes, from crop soil and plant Lichtfouse et al. assessed the endogenous origin of humic substances from crop soils¹³⁵. A combined approach using microscopy, pyrolysis, structural characterisation and ¹³C/¹²C analysis showed than soil humic substances are formed in part by selective preservation of resistant, aliphatic biopolymers¹³⁶⁻¹³⁷. Alternative pathways of humic substances from the same soil sample can occur into two compartments, a free, extractable compartment and an older, humin-bound compartment^{27,28}. Filley et al. developped ¹³C-tetramethylamonium hydroxide (TMAH) chemolysis to track microbial modification of lignin in terrestrial environments¹³⁸. More specifically, ¹³C-TMAH thermochemolysis of fungi-degraded lignin followed by gas chromatograhy-mass spectrometry (GC-MS) analysis enabled to determine the amount of hydroxylated aromatic components in degraded lignin residues¹³⁸.

MICROBIOLOGY

A large number of microbial species living in natural media remain unidentified, notably because they can not be easily grown as pure strains in the laboratory. Here, several recent reports show that investigations coupling ¹³C molecular tools and molecular genetic approaches open unexplored research areas¹³⁹⁻¹⁴¹. Boschker et al. used ¹³C labelling of polar lipid fatty acids (PLFAs) to identify the bacteria involved in two important biogeochemical processes: sulphate reduction coupled to oxidation in estuarine sediments, and methane oxidation in a freshwater sediment¹³⁹. Hinrichs et al. further demonstrated the consumption of sedimentary methane by archaebacteria using two lines of evidence: extreme ¹³C-depletion of archaebacterial lipid biomarkers (-100‰) and 16S rRNA sequencing¹⁴⁰. A very promising stable-isotope tool for microbial ecology has also been set up by Radajewski et al.¹⁴¹. Indeed, since ¹³C-DNA can be separated from ¹²C-DNA by centrifugation, it is possible to identify microorganisms living in complex media, e.g. soils, and growing on specific ¹³C-labelled sources, e.g. methanol¹⁴¹. A such technique should be applied, for instance, to the identification of strains involved in organic pollutant degradation.

ORGANIC GEOCHEMISTRY

Following the traditional use of isotopes in geosciences², the applications of CG-C-IRMS in biogeochemistry¹⁰ have been blooming early and rapidly, as shown by the special issue edited by Schoell and Hayes⁹. Nonetheless, geochemists have been able to measure isotope values of individual substances⁴²⁻⁵⁷ prior the advent of GC-C-IRMS. Indeed, the fractionation of sedimentary mixtures into pure substances has been performed, for instance by HPLC of amino acids^{142,143} and of chlorophyll-derived

porphyrins¹⁴⁴⁻¹⁴⁶. Compound-specific isotope analysis has been applied mainly to source identification, palaeoreconstruction and petroleum geochemistry.

Freeman et al. analysed extracts from the Eocene Messel shale by GC-C-IRMS and found strikingly ¹³C-depleted values for bacterial hopanes, suggesting their derivation from methanotrophs living at the time of deposition¹⁴⁷. Similarly depleted values were observed in the Green River shale by Collister et al.¹⁴⁸, thus allowing to bring some light of the biological sources and food webs of the palaeoenvironment⁸³. Lichtfouse and Collister further observed an isotopic correlation between fossil *n*-alkanes and fossil *n*-fatty acids, thus confirming the production of petroleum alkanes by decarboxylation of fatty acids¹⁴⁹. Collister et al. reviewed *n*-alkane literature and proposed on isotopic grounds various bacterial, planktonic and plant sources for Green River sedimentary *n*-alkanes¹⁵⁰.

Kennicutt and Brooks observed that even-carbon number sedimentary *n*-alkanes are ¹³C-enriched relative to their odd-carbon counterparts¹⁵¹. By comparison of ¹³C/¹²C values of *n*-alkanes from leaf and lake sediments, Rieley inferred the plant origin of long-chain sedimentary *n*-alkanes¹⁵². Nonetheless other studies suggested possible alternative sources^{153,154}. Jones et al. reported ¹³C/¹²C values of *n*-alcohols, sterols and an hopanoid alcohol from surface sediments¹⁵⁵. Kenig et al. presented molecular and isotopic evidence for the presence in sediments of branched alkanes derived from insect waxes¹⁵⁶. Hartgers et al. assessed the derivation of ¹³C-enriched sedimentary carotenoid derivatives from photosynthetic green sulphur bacteria¹⁵⁷ because the biomass of these organisms is isotopically labelled, being enriched in ¹³C. Moers et al. observed unusually heavy ¹³C/¹²C values (-5‰) of monosaccharides extracted from surface sediments¹⁵⁸.

Organic sulphur is playing a major role in the formation and transformation of sedimentary organic matter^{159,160}. Here, isotopic analysis of molecular fossils released by desulfurization from sedimentary macromolecules brought some light on their biological sources, e.g. bacteria, algae, and higher plants¹⁶¹⁻¹⁶⁵. In particular, it is shown that free and S-bound substances have different ¹³C/¹²C values, suggesting their derivation from different sources.

One of the most fascinating field of application of carbon isotope analysis is the reconstruction of palaeoenvironments, as shown by Schidlowski who inferred life occurrence 3,800 millions years ago using the biosynthetic fingerprint of carbonates versus organic matter¹⁶⁶. Using ¹³C/¹²C ratios of fossil *n*-alkanes, Logan et al. brought some light of the water-column processes ruling the transition from anoxic, bacteria-enriched to oxic, algae-enriched waters during the Proterozoic aeon (2,500-540 Myrs)¹⁶⁷. The isotopic analysis of fossil photosynthetic pigments revealed the activity of photosynthetic sulphur bacteria in the Black Sea for substantial periods of time in the past, as shown by Sinninghe Damsté et al.¹⁶⁸. Jasper and Hayes found that ¹³C/¹²C ratios of fossil algal alkenones recorded atmospheric CO₂ levels during the late quaternary (0-100 kyrs)¹⁶⁹. Freeman and Hayes explained the principles of calculation of ancient CO₂

levels, monitoring for example the ¹³C/¹²C values of porphyrins, the chlorophyll fossils¹⁷⁰. In order to further understand ancient biogeochemical cycles, investigations of modern water-column processes are under way¹⁷¹.

Petroleum geochemists have extensively used carbon isotopes¹⁷²⁻¹⁷⁵ because the processes implied in the formation and migration of petroleum in sedimentary rocks are still obscure. Bjorøy et al. found clear ¹³C/¹²C differences for *n*-alkanes versus acyclic isoprenoids in North Sea oils¹⁷⁶. Schoell et al. showed that fossil biomarkers from Miocene tar-like petroleum can be related to their biological sources according to their ¹³C/¹²C¹⁷⁷. Lichtfouse and Collister inferred the formation of petroleum *n*-alkanes by decarboxylation of *n*-fatty acids¹⁴⁹. Gõni and Eglinton investigated the ¹³C/¹²C ratios of aliphatic and phenolic substances released by pyrolysis of petroleum precursors¹⁷⁸. Simoneit and Schoell confirmed the generation of PAHs from hydrothermal petroleum by breakdown of sedimentary kerogen, the macromolecular fraction of sedimentary organic matter¹⁷⁹. Lichtfouse et al. found that ¹³C/¹²C ratios of *n*-alkanes of deep rocks and petroleums from Paris basin fall in a narrow range¹⁸⁰, suggesting that the carbon of only a restricted number of living organisms is contributing to petroleum genesis.

By setting up ¹⁴C dating of trace organics at the molecular level using preparative GC, Eglinton et al. have opened a novel vista in organic geochemistry^{60,61}. Indeed, the assessment of relationships between molecules occurring in complex media such as soils and sediments have so far mainly relied on structural grounds. Now, ¹⁴C dating provides an independent way to unravel the biological sources of sedimentary organic molecules. For instance, in a 4-7 cm marine sediment of bulk organic matter age of 880 years BP, the large age difference observed between the C₂₅ *n*-alkane (1340 years) and the C₃₁ *n*-alkane (500 years) can be explained by a higher contribution of fossil carbon in the former⁶¹. These results are in agreement with the fossil fuel origin of short-chain linear alkanes found in soils⁸⁹. It is thus expected that ¹⁴C molecular dating will find applications into various fields, notably for environmental issues because human activities have introduced large amounts of fossil fuel ¹⁴C-(almost)free carbon in ecosystems (pollutants, plastics, solvents, drugs, etc.).

SOIL SCIENCE

Soil organic matter has many agronomic functions such as water retention, plant-nutrient sequestration, and holding of minerals^{91,181}. As in any complex ecosystem, the behaviour of soil organic molecules is far from being understood. Most soil organic substances still remain unidentified. Here, isotopic studies at the molecular level appear as a promising way to unravel the multiple pathways of organic matter transformation. For instance, the long-term turnover of plant wax *n*-alkanes¹⁷ in soil has been monitored using a 23-year natural labelling experiment by cultivation of maize^{19,20}. The plant origin of long-chain *n*-alkanes¹⁸², *n*-alcohols⁸² and *n*-fatty acids⁸², and the microbial origin of short-chain fatty acids⁷⁵ have been assessed using both natural and artificial ¹³C-labelling experiments. Huang et al. studied the ¹³C/¹²C values

of lipids of stratified organic soils¹⁸³. They found that hopanoid were ¹³C-enriched by 4‰ relative to plant *n*-alkanes, suggesting a source of heterotrophic bacteria feeding on carbohydrates or proteins. A 23-year plant decomposition experiment further revealed the absence of isotopic fractionation of *n*-alkanes, thus validating their use as proxies for palaeoenvironmental reconstruction and as tracers of carbon flow in soils and sediments¹⁸⁴.

SPORT

The detection of drugs used to improve sport performances is difficult because administrated substances are usually occurring at trace levels in urine and in blood. It is even more difficult when the drugs, e.g. steroids, are already naturally biosynthesized by the body. Here, similarly to aroma authentication, natural and synthetic homologues can be isotopically distinguished because the carbon of the synthetic substance is of petroleum origin. For instance, the ¹³C/¹²C analysis of urinary steroids to assess the misuse of testosterone has been set up^{15,185-189}. Aguilera et al. further extended this technique to the detection of hydrocortisone used for horseracing¹⁹⁰.

CONCLUSION

The recent development of mass spectrometric and NMR techniques allowing the determination of isotopic composition at the molecular and atomic levels has opened novel research fields into various scientific disciplines. Such techniques are already used for industrial applications, e.g. petroleum research and food autentication. Since there is a growing need for tracers to resolve various issues related to complex systems, it is expected that isotopic analysis at the molecular and atomic levels will become widely used. ¹⁴C-molecular dating, GC-C-IRMS (¹³C, ²H, ¹⁵N), and intramolecular isotopic analysis by SNIF-NMR look very promising. Molecular investigations undertaken at the frontiers of several fields, e.g. chemistry, geology and biology¹⁹¹, could be particurlarly fruitful.

ACKNOWLEDGEMENTS

This article is dedicated to John Hayes, Dawn Merritt, Kate Freeman, Jim Collister, David Hollander, Glenn Hieshima, John Jasper, Lisa Pratt, John Rupp and all other fellows from Indiana University, Bloommington IN, with whom I have had a good time and hard work during my post-doctoral stay in 1990. I also thank Dr. Nick Ostle, Institute of Terrestrial Ecology for his kind invitation to present a conference on CSIA at the annual meeting of the stable isotope mass spectrometry users group (SIMSUG), Grange-Over-Sands, UK (18th-19th January 2000).

REFERENCES

1. H. Craig, *Geochim. Cosmochim. Acta* **12**, 133-149 (1957). Carbon isotopic compositions are expressed in per mil. relative to the Pee Dee Belemnite carbonate standard $\delta^{13}C = [(^{13}C/^{12}C \text{ sample } -$

 $^{13}C/^{12}Cstd)/(^{13}C/^{12}Cstd)$], where $^{13}C/^{12}Cstd = 0.0112372$.

- 2. H. Craig, Geochim. Cosmochim. Acta 3, 53-92 (1953).
- 3. M. Sano, Y. Yotsui, H. Abe, and S. Sasaki, *Biomed. Mass Spectrom.* 3, 1-3 (1976).
- 4. D.E. Matthews and J.M. Hayes, Anal. Chem. 50, 1465-1473 (1978).
- 5. A. Barrie, J. Bricout, J. Koziet, Spectros. Int. J. 3, 259-268 (1984).
- 6. J.T. Brenna, Acc. Chem. Res. 27, 340-346 (1994).
- 7. E. Lichtfouse and H. Budzinski, Analusis 23, 364-369 (1995).
- 8. W. Meier-Augenstein, J. Chrom. A 842, 351-371 (1999).

9. M. Schoell and J.M. Hayes (Eds.), Compound-specific isotope analysis in biogeochemistry and petroleum research, *Org. Geochem.* **21**, 561-827 (1994).

- 10. J.M. Hayes, K.H. Freeman, B.N. Popp and C.H. Hoham, Org. Geochem. 16, 1115-1128 (1990).
- 11. A. Mosandl, U. Hener, H.-G. Schmarr and M. Rautenschlein, *J. High Resol. Chrom.* **13**, 528-531 (1990).

12. M.J. Rennie, W. Meier-Augenstein, P.W. Watt, A. Patel, I.S. Begley and C.M. Scrimgeour, *Biochem. Mass Spec.* **24**, 927-932 (1996).

- 13. W. Meier-Augenstein, Curr. Opin. Clin. Nutr. Metab. Care 2, 465-470 (1999).
- 14. J.P. Jasper, *Pharmac. Techn.* 5 p., oct. 1999.

15. M. Becchi, R. Aguilera, Y. Farizon, M.-M. Flament, H. Casabianca and P. James, *Rapid Commun. Mass Spectrom.* **8**, 304-384 (1994).

16. G. Rieley, J.W. Collister, B. Stern and G. Eglinton, *Rapid Commun. Mass Spectrom.* **7**, 488-491 (1993).

- 17. J.W. Collister, G. Rieley, B. Stern, G. Eglinton and B. Fry, Org. Geochem. 21, 619-627 (1994).
- 18. R.P. Evershed, K.I. Arnot, J. Collister, G. Eglinton and S. Charters, Analyst 119, 909-914 (1994).
- 19. E. Lichtfouse, Tetrahedron Lett. 36, 529-530 (1995).
- 20. E. Lichtfouse, Naturwissenschaft. 84, 22-23 (1997).

21. B. Sherwood Lollar and T.A. Abrajano, Jr (Eds.) Compound-specific isotope analysis: tracing organic contaminant sources and processes in geochemical systems. *Org. Geochem.* **30**, 721-871 (1999).

- 22. M. Usaki, K. Yamada and R. Ishiwatari, Geochem. J. 27, 385-389 (1993).
- 23. V.P. O'Malley, T.A. Abrajano Jr and J. Hellou, Org. Geochem. 21, 809-822 (1994).
- 24. E. Lichtfouse and T. Eglinton, Org. Geochem. 23, 969-973 (1995).
- 25. H.S. Dempster, B. Sherwood Lollar and S. Feenstra, Env. Sci. Technol. 31, 3193-3197 (1997).
- 26. E. Lichtfouse, S. Dou, S. Houot and E. Barriuso, Org. Geochem. 23, 845-847 (1995).
- 27. E. Lichtfouse, P. Wehrung and P. Albrecht, Naturwissenschaft. 85, 449-452 (1998).
- 28. E. Lichtfouse, Analusis/Eur. J. Anal. Chem. 27, 385-386 and 442-444 (1999).

- 29. M.H. Engel, S.A. Macko and J.A. Silfer, *Nature* 348, 47-49 (1990).
- 30. M. Desage, R. Guilluy, J.L. Brazier, H. Chaudron, J. Girard, H. Cherpin and J. Jumeau, *Anal. Chim. Acta* **247**, 249-254 (1991).
- 31. F. Besacier, R. Guilluy, J.L. Brazier, H. Chaudron-Thozet, J. Girard and A. Lamotte, *J. Forensic Sci.* **42**, 429-433 (1997).
- 32. D.H. Chace and F.P. Abramson, Anal. Chem. 61, 2724-2730 (1989).
- 33. D.A. Merritt, J. M. Hayes, J. Am. Soc. Mass Spectrom. 5, 387-397 (1994).
- 34. C.M. Scrimgeour, I.S. Begley and M.L. Thomason, *Rapid Commun. Mass Spectrom.* **13**, 271-274 (1999).
- 35. A.L. Sessions, T.W. Burgoyne, A. Schimmelmann and J.M. Hayes, *Org. Geochem.* **30**, 1193-1200 (1999).
- 36. R.J. Caimi and J.T. Brenna. J. Mass Spectrom. 30, 466-472 (1995).
- 37. R.J. Caimi and J.T. Brenna. Anal. Chem. 65, 3497-3500 (1993).
- 38. G.J. Martin, X.Y. Sun, C. Guillou and M.L. Martin, *Tetrahedron* 41, 3285-3296 (1985).
- 39. G.J. Martin, M.L. Martin and B.L. Zhang, Plant Cell Env. 15, 1037-1050 (1992).
- 40. V. Caer, M. Trierweiler, G.J. Martin and M.L. Martin, Anal. Chem. 63, 2306-2313 (1991).
- 41. T.I. Eglinton, L.I. Aluwihare, J.E. Bauer, E.R.M. Druffel and A.P. McNichol, *Anal. Chem.* **68**, 904-912 (1996).
- 42. P.H. Abelson and T.C. Hoering, Proc. Nat. Acad. Sci. 47, 623-632 (1961).
- 43. E. Bengsch, J.Ph. Grivet and H.-R. Schulten, Z. Naturforsch. 36b, 1289-1296 (1981).
- 44. P.L. Parker, Carnegie Inst. Yearb. 61, 187-190 (1961-62).
- 45. P.L. Parker, Geochim. Cosmochim. Acta 28, 1155-1164 (1964).
- 46. E.A. Vogler and J.M. Hayes, J. Org. Chem. 44, 3682-3686 (1979).
- 47. E.A. Vogler and J.M. Hayes, Carbon isotopic compositions of carboxyl groups of biosynthesized fatty
- acids. In Adv. Org. Geochem. 1979, Eds. A.G. Douglas and J.R. Maxwell, Pergamon, 697-704 (1980).
- 48. E.A. Vogler and J.M. Hayes, Int. J. Appl. Rad. Isot. 29, 297-300 (1978).
- 49. K.D. Monson and J.M. Hayes, J. Biol. Chem. 255, 11435-11441 (1980).
- 50. K.D. Monson and J.M. Hayes, Geochim. Cosmochim. Acta 46, 139-149 (1982).
- 51. W.G. Meinschein, G.G.L. Rinaldi, J.M. Hayes and D.A. Schoeller, *Biomed. Mass Spectrom.* **1**, 172-174 (1974).
- 52. G. Rinaldi, W.G. Meinschein and J.M. Hayes, Biomed. Mass Spectrom. 1, 412-414 (1974).
- 53. G. Rinaldi, W.G. Meinschein and J.M. Hayes, *Biomed. Mass Spectrom.* 1, 415-417 (1974).
- 54. D.H. Welte, Naturwissenschaft. 56, 133-134 (1969).

55. D.H. Welte, Determination of C13/C12 isotope ratios of individual higher *n*-paraffins from different petroleums. In *Adv. Org. Geochem. 1968* (Eds. P.A. Schenck and I. Havenaar), Pergamon, 269-277.
56. E.M. Galimov and V.G. Shirinskiy, *Geochim. Int.* 12, 157-180 (1975).

- 57. E.M. Galimov, L.A. Kodina and V.N. Generalova, *Geochim. Int.* 13, 9-13 (1975).
- 58. S.N. Dudd and R.P. Evershed, *Science* 282, 1478-1481 (1998).
- 59. A.W. Stott, R.P. Evershed and N. Tuross, Org. Geochem. 26, 99-103 (1997).
- 60. T.I. Eglinton, L.I. Aluwihare, J.E. Bauer, E.R.M. Druffel and A.P. McNichol, *Anal. Chem.* **68**, 904-912 (1996).
- 61. T.I. Eglinton, B.C. Benitez-Nelson, A. Pearson, A.P. McNichol, J.E. Bauer and E.R.M. Druffel, *Nature* **277**, 796-799 (1997).
- 62. S. Tissot, S. Normand, R. Guilluy, C. Pachiaudi, M. Beylot, M. Laville, R. Cohen, R. Mornex and J.P. Riou, *Diabetologia* **33**, 449-456 (1990).
- 63. Y. Khalfallah, S. Normand, S. Tissot, C. Pachiaudi, M. Beylot and J.P. Riou, *Biol. Mass Spectrom.* 22, 707-711 (1993).
- 64. J. Koziet, P. Gross, G. Debry and M.J. Royer, Biol. Mass Spectrom. 20, 777-782 (1991).
- 65. T. Preston and C. Slater, Proc. Nutrit. Soc. 53, 363-372 (1994).
- 66. K.E. Yarasheski, K. Smith, M.J. Rennie and D.M. Bier, Biol. Mass Spectrom. 21, 486-490 (1992).
- 67. T. Preston, K.C.H. Fearon, D.C. McMillan, F.P. Winstanley, C. Slater, A. Shenkin and D.C. Carter, *British J. Surg.* **82**, 229-234 (1995).
- 68. W. Meier-Augenstein, D. Rating, G.F. Hoffmann, U. Wendel, U. Matthiesen and P. Schadewaldt, *Isotopes Environ. Health Stud.* **31**, 261-266 (1995).
- 69. Z.K. Guo, A.H. Luke, W.P. Lee and D. Schoeller, Anal. Chem. 65, 1954-1959 (1993).
- 70. B. Deruaz, D. Deruaz, A. Bannier, M. Desage and J.L. Brazier, Analusis 22, 241-248 (1994).
- 71. K.J. Goodman and J.T. Brenna, Anal. Chem. 64, 1088-1095 (1992).
- 72. N. Brossard, C. Pachiaudi, M. Croset, S. Normand, J. Lecerf, V. Chirouze, J.P. Riou, J.L. Tayot and M. Lagarde, *Anal. Biochem.* **220**, 192-199 (1994).
- 73. S.T. Brookes, K.S. Craig and S.C. Cunnane, Biochem. Soc. Trans. 22, 164S (1994).
- 74. G. Rieley, Analyst 119, 915-919 (1994).
- 75. E. Lichtfouse, G. Berthier, S. Houot, E. Barriuso, V. Bergheaud and T. Vallaeys, *Org. Geochem.* 23, 849-852 (1995).
- 76. C.M. Scrimgeour, I.S. Begley and M.L. Thomason, *Rapid Commun. Mass Spectrom.* **13**, 271-274 (1999).
- 77. J.M. Hayes. Marine Geol. 113, 111-125 (1993).
- 78. J. Fang, T.A. Abrajano, P.A. Comet, J.M. Brooks, R. Sassen and I.R. MacDonald, *Chem. Geol. Isot. Geosc. Sect.* **109**, 271-279 (1993).
- 79. M. O'Leary, Phytochem. 20, 553-567 (1981).
- 80. E. Lichtfouse, S. Derenne, A. Mariotti and C. Largeau, Org. Geochem. 22, 1023-1027 (1994).
- 81. M.J. Lockheart, P.F. Van Bergen and R.P. Evershed, Org. Geochem. 26, 137-153 (1997).
- 82. E. Lichtfouse, Naturwissensch. 85, 76-77 (1998).

83. R.E. Summons, L.L. Jahnke and Z. Roksandic, Geochim. Cosmochim. Acta 58, 2853-2863 (1994).

- 84. R.E. Summons, P.D. Franzmann and P.D. Nichols, Org. Geochem. 28, 465-475 (1998).
- 85. M.T.J. Van der Meer, S. Schouten and J.S. Sinninghe Damsté, Org. Geochem. 28, 527-533 (1998).
- 86. M.-Y. Sun, Org. Geochem. 31, 199-209 (2000).

87. R. Ishiwatari, M. Uzaki and K. Yamada, Org. Geochem. 21, 801-808 (1994).

88. K.M. Rogers and M.M. Savard, Org. Geochem. 30, 1559-1569 (1999).

89. E. Lichtfouse, G. Bardoux, A. Mariotti, J. Balesdent, D.C. Ballentine and S.A. Macko, *Geochim. Cosmochim. Acta* **61**, 1891-1898 (1997).

90. E. Lichtfouse, S.E. Apitz and M. Nanny (Eds.) The biogeochemistry of polycyclic aromatic hydrocarbons: sources interactions, biodegradation, toxicity and analytical developments. *Org. Geochem.*30, 873-969 (1999).

91. E. Lichtfouse (Ed.) Soil pollutants. Analusis 25, M16-M72 (1997).

92. V.P. O'Malley, R.A. Burke and W.S. Schlotzhauer, Org. Geochem. 27, 567-581 (1997).

93. D.C. Ballentine, S.A. Macko, V.C. Turekian, W.P. Gilhooly and B. Martincigh, *Org. Geochem.* **25**, 97-104 (1996).

94. C. McRae, C.-G. Sun, C.E. Snape, A.E. Fallick and D. Taylor, Org. Geochem. 30, 881-889 (1999).

95. E. Lichtfouse, H. Budzinski, Ph. Garrigues and T.I. Eglinton, Org. Geochem. 26, 353-359 (1997).

96. H.H. Richnow, A. Eschenbach, B. Mahro, R. Seifert, P. Wehrung, P. Albrecht and W. Michaelis, *Chemosphere* **36**, 2211-2224 (1998).

97. R.A. Harrington, S.R. Poulson, J.I. Drever, P.J.S. Colberg and E.F. Kelly, *Org. Geochem.* **30**, 765-775 (1999).

98.R.U. Meckenstock, B. Morasch, R. Warthmann, B. Schink, E. Annweiler, W. Michaelis and H.H. Richnow, *Environ. Microbiol.* **1**, 409-414 (1999).

99. Y. Zeng, H. Mukai, H. Bandow and Y. Nojiri, Anal. Chim. Acta 289, 195-204 (1994).

100. M.E. Conrad, A.S. Templeton, P.F. Daley and L. Alvarez-Cohen, Org. Geochem. 30, 843-859 (1999).

101. A. Jackson and J. Pardue, Org. Geochem. 30, 787-792 (1999).

102. D. Rowe and K. Muehlenbachs, Org. Geochem. 30, 861-871 (1999).

103. H. Dayan, T. Abrajano, N.C. Stuchio and L. Winsor, Org. Geochem. 30, 755-763 (1999).

104. L.J. Heraty, M.E. Fuller, L. Huang, T. Abrajano Jr and N.C. Sturchio, *Org. Geochem.* **30**, 793-799 (1999).

105. B. Sherwood Lollar, G.F. Slater, J. Ahad, B. Sleep, J. Spivack, M. Brennan and P. MacKenzie, *Org. Geochem.* **30**, 813-820 (1999).

106. L.G. Stehmeier, M.McD. Francis, T.R. Jack, E. Diegor, L. Winsor and T.A. Abrajano Jr, *Org. Geochem.* **30**, 821-833 (1999).

107. B.A. Bergamaschi, M.S. Fram, C. Kendall, S.R. Silva, G.R. Aiken and R. Fujii, *Org. Geochem.* **30**, 835-842 (1999).

108. R.F. Dias and K.H. Freeman, Anal. Chem. 69, 944-950 (1997).

109. S. Epstein, R.V. Krishnamurthy, J.R. Cronin, S. Pizzarello and G.U. Yuen, *Nature* **326**, 477-479 (1987).

110. J.A. Silfer, M.H. Engel, S.A. Macko and E.J. Jumeau, Anal. Chem. 63, 370-374 (1991).

111. M.H. Engel and S.A. Macko, *Nature* **389**, 265-268 (1997).

112. H.W. Krueger and R.H. Reesman, Mass Spectrom. Rev. 1, 205-236 (1982).

- 113. G. Martin, Ann. Fals. Exp. Chim. 85, 249-270 (1992).
- 114. A. Mosandl, J. Chrom. 624, 267-292 (1992).

115. J.L. Brazier, R. Guilluy and S. Dautraix, Spectra Analyse 186, 26-29 (1995).

116. A. Bernreuther, J. Koziet, P. Brunerie, G. Krammer, N. Christoph and P. Schreier, *Z. Lebensm. Unters. Forsch.* **191**, 299-301 (1990).

117. M. Bigois, H. Casabianca, J.B. Graff, B. Philit, P. Jame and C. Perrucchietti, *Spectra Analyse* **181**, 19-22 (1994).

118. H. Casabianca and J.B. Graff, J. Chrom. A 684, 360-365 (1994).

119. U. Hener, S. Faulhaber, P. Kreis and A. Mosandl, *Pharmazie* 50, 60-62 (1995).

120. H.-L. Schmidt, M. Butzenlechner, A. Rossmann, S. Schwarz, H. Kexel and K. Kempe, *Z. Lebensm. Unters. Forsch.* **196**, 105-110 (1993).

- 121. R. Braunsdorf, U. Hener, S. Stein and A. Mosandl, Z. Lebensm. Unters. Forsch. 197, 137-141 (1993).
- 122. R. Braunsdorf, U. Hener and A. Mosandl, Z. Lebensm. Unters. Forsch. 194, 426-430 (1992).
- 123. O. Breas, F. Fourel and G.J. Martin, Analusis 22, 268-272 (1994).
- 124. V. Karl, A. Dietrich and A. Mosandl, *Phytochem. Anal.* 5, 32-37 (1994).
- 125. G. Martin, G. Remaund and G.J. Martin, Flavour Fragr. J. 8, 97-107 (1993).
- 126. G.J. Martin, M.L. Martin, F. Mabon and M.-J. Michon, Anal. Chem. 54, 2382-2384 (1982).

127. D. Danho, N. Naulet and G.J. Martin, Analusis 20, 179-184 (1992).

128. S. Hanneguelle, J.-N. Thibault, N. Naulet and G.J. Martin, J. Agric. Food Chem. 40, 81-87 (1992).

129. G.J. Martin, C. Guillou, M.L. Martin, M.-T. Cabanis, Y. Tep and J. Aerny, *J. Agric. Food Chem.* **36**, 316-322 (1988).

130. G.J. Martin, D. Odiot, V. Godineau, N. Naulet, R. Morlat, C. Asselin and J. Robichet, *Appl. Geochem.* **4**, 1-11 (1989).

131. J.L. Brazier, *Toxicorama* 4, 13-18 (1992).

132. J.H. Liu, W.F. Lin, M.P. Fitzerald, S.C. Saxena and Y.N. Shieh, J. Forensic Sci. 24, 814-816 (1979).

133. F. Mas, B. Beemsterboer, A.C. Veltkamp and A.M.A. Verweij, Forensic Sci. Int. 71, 225-231 (1995).

134. E. Lichtfouse and J. Lévêque (Eds.) Humic Substances, Eur. J. Anal. Chem. 27, 385-444 (1999).

135. E. Lichtfouse, S. Dou, C. Girardin, M. Grably, J. Balesdent, F. Behar and M. Vandenbroucke, *Org. Geochem.* **23**, 865-868 (1995).

136. E. Lichtfouse, C. Chenu, F. Baudin, C. Leblond, M. Da Silva, F. Behar, S. Derenne, C. Largeau, P. Wehrung and P. Albrecht, *Org. Geochem.* 28, 411-415 (1998).

137. E. Lichtfouse, C. Chenu and F. Baudin, Org. Geochem. 25, 263-265 (1996).

138. T.R. Filley, P.G. Hatcher, W.C. Shortle and R.T. Praseuth. Org. Geochem. 31, 181-198 (2000).

139. H.T.S. Boschker, S.C. Nold, P. Wellsbury, D. Bos, W. de Graaf, R. Pel, R.J. Parkes and T.E.

Cappenberg, Nature 392, 801-805 (1998).

140. K.-U. Hinrichs, J.M. Hayes, S.P. Sylva, P.G. Brewer and E.F. DeLong, Nature 398, 802-805 (1999).

141. S. Radajewski, P. Ineson, N.R. Parekh and J.C. Murrell, Nature 403, 646-649 (2000).

142. M.H. Engel and S.A. Macko, *Nature* **323**, 531-533 (1986).

143. P. Edgar Hare, M.L. Fogel, T.W. Stafford Jr., A.D. Mitchell and T.C. Hoering, *J. Archeol. Sci.* **18**, 277-292 (1991).

144. J.M. Hayes, R. Takigiku, R. Ocampo, H.J. Callot and P. Albrecht, Nature 329, 48-51 (1987).

145. R. Ocampo, H.J. Callot, P. Albrecht, B.N. Popp, M.R. Horowitz and J.M. Hayes, *Naturwissenschaft*. **76**, 419-421 (1989).

146. J.M. Hayes, B.N. Popp, R. Takigiku and M. Johnson, *Geochim. Cosmochim. Acta* **53**, 2961-2972 (1989).

147. K.H. Freeman, J.M. Hayes, J.-M. Trendel and P. Albrecht, *Nature* 343, 254-256 (1990).

148. J.W. Collister, R.E. Summons, E. Lichtfouse and J.M. Hayes, Org. Geochem. 19, 265-276 (1992).

149. E. Lichtfouse and J.W. Collister, Tetrahedron Lett. 33, 8093-8094 (1992).

150. J.W. Collister, E. Lichtfouse, G. Hieshima and J.M. Hayes, Org. Geochem. 21, 645-659 (1994).

151. M.C. Kennicutt II and J.M. Brooks, Org. Geochem. 15, 193-197 (1990).

152. G. Rieley, R.J. Collier, D.M. Jones, G. Eglinton, P.A. Eakin and A.E. Fallick, *Nature* **352**, 425-426 (1991).

153. N. Spooner, G. Rieley, J.W. Collister, M. Lander, P.A. Cranwell and J.R. Maxwell, *Org. Geochem.* **21**, 823-827 (1994).

154. E. Lichtfouse, S. Derenne, A. Mariotti and C. Largeau, Org. Geochem. 22, 1023-1027 (1994).

155. D.M. Jones, J.F. Carter, G. Eglinton, E.J. Jumeau and C.S. Fenwick, *Biol. Mass Spectrom.* **20**, 641-646 (1991).

156. F. Kenig, J.S. Sinninghe Damsté, J.W. De Leeuw and J.M. Hayes, *Naturwissenschaft.* **81**, 128-130 (1994).

157. W.A. Hartgers, J.S. Sinninghe Damsté, A.G. Requejo, J. Allan, J.M. Hayes and J.W. de Leeuw, *Nature* **369**, 224-227 (1994).

158. M.E.C. Moers, D.M. Jones, P.A. Eakin, A.E. Fallick, H. Griffiths and S.R. Larter, *Org. Geochem.* **20**, 927-933 (1993).

159. J.S. Sinninghe Damsté and J.W. de Leeuw, Org. Geochem. 16, 1077-1101 (1990).

160. P. Adam, J.C. Schmid, B. Mycke, C. Strazielle, J. Connan, A. Huc, A. Riva and P. Albrecht, *Geochim. Cosmochim. Acta* 57, 3395-3419 (1993).

161. M.E.L. Kohnen, S. Schouten, J.S. Sinninghe Damsté, J.W. de Leeuw, D.A. Merritt and J.M. Hayes, *Science* **256**, 358-362 (1992).

162. M.E.L. Kohnen, S. Schouten, J.S. Sinninghe Damsté, J.W. de Leeuw, D. Merritt and J.M. Hayes, *Org. Geochem.* **19**, 403-419 (1992).

163. T.R. Filley, K.H. Freeman and P.G. Hatcher, Org. Geochem. 25, 367-377 (1996).

164. S. Schouten, J.S. Sinninghe Damsté, M.E.L. Kohnen and J.W. de Leeuw, *Geochim. Cosmochim. Acta* **59**, 1605-1609 (1995).

165. F. Kenig, J.S. Sinninghe Damsté, N.L. Frewin, J.M. Hayes and J.S. De Leeuw, *Org. Geochem.* 23, 485-526 (1995).

166. M. Schidlowshi, Nature 333, 313-318 (1988).

167. G.A. Logan, J.M. Hayes, G.B. Hieshima and R.E. Summons, *Nature* 376, 53-56 (1995).

168. J.S. Sinninghe Damsté, S.G. Wakeham, M.E.L. Kohnen, J.M. Hayes and J.W. de Leeuw, *Nature* **362**, 827-829 (1993).

169. J.J. Jasper and J.M. Hayes, *Nature* **347**, 463-464 (1990).

170. K.H. Freeman and J.M. Hayes, *Glob. Biogeoch. Cycles* 6, 185-198 (1992).

171. R.D. Pancost, K.H. Freeman and S.G. Wakeham, Org. Geochem. 30, 319-340 (1999).

172. S.R. Silverman and S. Epstein, Bull. Amer. Assoc. Petr. Geol. 42, 998-1012 (1958).

173. S.R. Silverman, J. Amer. Oil Chem. Soc. 44, 691-695 (1967).

174. Z. Sofer, Amer. Assoc. Petr. Geol. Bull. 68, 31-49 (1984).

175. M. Schoell, Org. Geochem. 6, 645-663 (1984).

176. M. Bjorøy, K. Hall, P. Gillyon and J. Jumeau, Chem. Geol. 93, 13-20 (1991).

177. M. Schoell, M.A. McCaffrey, F.J. Fago and J.M. Moldowan, *Geochim. Cosmochim. Acta* 56, 1391-1399 (1992).

178. M.A. Gõni and T.I. Eglinton, J. High Resol. Chrom. 17, 476-488 (1994).

179. B.R.T. Simoneit and M. Schoell, Org. Geochem. 23, 857-863 (1995).

180. E. Lichtfouse, P. Albrecht, F. Behar and J.M. Hayes, *Geochim. Cosmochim. Acta* 58, 209-221 (1994).

181. M. Schnitzer, *Soil Science* **151**, 41-58 (1991).

182. E. Lichtfouse, B. Elbisser, J. Balesdent, A. Mariotti and G. Bardoux, *Org. Geochem.* **22**, 349-351 (1994).

183. Y. Huang, R. Bol, D.D. Harkness, P. Ineson and G. Eglinton, Org. Geochem. 24, 273-287 (1996).

184. Y. Huang, G. Eglinton, P. Ineson, P.M. Latter, R. Bol and D.D. Harkness, *Org. Geochem.* **26**, 497-501 (1997).

185. M. Becchi, R. Aguilera, Y. Farizon, M.-M. Flament, H. Casabianca and P. James, *Rapid Commun. Mass Spectrom.* **8**, 304-308 (1994).

186. R. Aguilera, M. Becchi, C. Grenot, H. Casabianca and C.K. Hatton, J. Chrom. B 687, 43-53 (1996).

187. R. Aguilera, M. Becchi, H. Casabianca, C.K. Hatton, D.H. Catlin, B. Starcevic and H.G. Pope Jr, *J. Mass Spectrom.* **31**, 169-176 (1996).

188. C.H.L. Shackleton, A. Phillips, T. Chang and Y. Li, Steroids 62, 379-387 (1997).

189. R. Aguilera, D.H.Catlin, M. Becchi, A. Phillips, C. Wang, R.S. Swerdlorf, H.G. Pope and C.K. Hatton, *J. Chrom. B***727**, 95-105 (1999).

190. R. Aguilera, M. Becchi, L. Mateus, M.A. Popot, Y. Bonnaire, H. Casabianca and C.K. Hatton, *J. Chrom. B* **702**, 85-91 (1997).

191. E. Lichtfouse, Fossiles Moleculaires d'intérêt microbiologique, pétrolier, agronomique et environnemental. *L'Actualité Chimique*, avril, 5-19 (2000).