

# An hierarchical artificial neural network system for the classification of transmembrane proteins

Claude Pasquier, Stavros Hamodrakas

### ▶ To cite this version:

Claude Pasquier, Stavros Hamodrakas. An hierarchical artificial neural network system for the classification of transmembrane proteins. Protein Engineering, Design and Selection, 1999, 12 (8), pp.631-634. hal-00170718v3

## HAL Id: hal-00170718 https://hal.science/hal-00170718v3

Submitted on 3 Mar 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

#### SHORT COMMUNICATION

# An hierarchical artificial neural network system for the classification of transmembrane proteins

#### C.Pasquier and S.J.Hamodrakas<sup>1</sup>

Faculty of Biology, Department of Cell Biology and Biophysics, University of Athens, Panepistimiopolis, Athens 15701, Greece

<sup>1</sup>To whom correspondence should be addressed

This work presents a simple artificial neural network which classifies proteins into two classes from their sequences alone: the membrane protein class and the non-membrane protein class. This may be important in the functional assignment and analysis of open reading frames (ORF's) identified in complete genomes and, especially, those ORF's that correspond to proteins with unknown function. The network described here has a simple hierarchical feedforward topology and a limited number of neurons which makes it very fast. By using only information contained in 11 protein sequences, the method was able to identify, with 100% accuracy, all membrane proteins with reliable topologies collected from several papers in the literature. Applied to a test set of 995 globular, water-soluble proteins, the neural network classified falsely 23 of them in the membrane protein class (97.7% of correct assignment). The method was also applied to the complete SWISS-PROT database with considerable success and on ORF's of several complete genomes. The neural network developed was associated with the PRED-TMR algorithm (Pasquier,C., Promponas, V.J., Palaios, G.A., Hamodrakas, J.S. and Hamodrakas, S.J., 1999) in a new application package called **PRED-TMR2.** A WWW server running the PRED-TMR2 software is available at http://o2.db.uoa.gr/PRED-TMR2 Keywords: membrane proteins/neural network/prediction/protein structure

#### Introduction

The number of protein sequences stored in public databases (78 197 in SWISS-PROT release 37, 178 773 in TrEMBL; Bairoch and Apweiler, 1998) is considerably larger than that of known protein structures (9129 in PDB; Sussman *et al.*, 1998): a gap that will continue to increase, as the experimental determination of the three-dimensional structure of a protein is a time consuming process compared with the time needed for the determination of the protein sequence. This is especially true for transmembrane proteins which are difficult to solve by X-ray crystallography.

Usually, the structure of a new protein having homologies above a certain level to another sequence of known structure can be predicted with reasonable accuracy (Persson and Argos, 1994; Rost *et al.*, 1994, 1995). However, the majority do not belong to this ideal case. For this set of proteins, prediction methods that do not depend on sequence alignments but using solely information contained in a sequence itself are necessary.

A number of methods or algorithms designed to locate the transmembrane regions in proteins, without the need for multiple-sequence alignment information, have been developed—for example, von Heijne (1992), Cserzo *et al.* (1997) and Pasquier *et al.* (1999), to mention just a few. However, these algorithms focus on the localization of transmembrane segments in known integral membrane proteins and are not suited to the discrimination of membrane proteins from non-membrane proteins.

Recently, we have published the PRED-TMR method in an attempt to improve the fine localization of transmembrane segments, by coupling a hydrophobicity analysis with a detection of potential termini (starts and ends) of transmembrane regions (Pasquier *et al.*, 1999). Now we have extended this application with a pre-processing stage, represented by an artificial neural network, which attempts to classify proteins into either membrane or non-membrane proteins.

Several applications of neural networks to the prediction of transmembrane segments or secondary structure prediction can be found in the literature (Reczko, 1993; Rost et al., 1994; Fariselli and Casadio, 1996; Aloy et al., 1997; Diederichs et al., 1998). Most of them use a local encoding for each amino acid and produce as output a classification for the amino acid in the middle of the input window. When an hierarchical feed-forward topology is used (the connectivity graph contains no loop), each network output is independent of the results obtained by previous processing. This causes little or no problem when the output of the network consists of continuous values (coordinates for example; Diederichs et al., 1998). However, when a threshold parameter is used for the choice of binary output, the absence of correlation between the possible structure of adjacent residues frequently results in incoherent topologies (a transmembrane segment composed of only one residue for example). This problem can be solved by designing recurrent neural networks which use additional information obtained with the processing of previous patterns (Reczko, 1993) or by building a system of cascading neural networks (Rost et al., 1994; Fariselli and Casadio, 1996). Nevertheless, these techniques are not appropriate for the correct classification between membrane and non-membrane proteins because they are too focused on one-residue topology prediction.

This paper presents an artificial neural network which does not predict the exact location of transmembrane segments, but produces instead a unique output showing whether an analyzed part of a sequence is related to a transmembrane region or not.

#### Materials and methods

#### Information gathering

Eleven proteins with known topologies were used for the training of the network: six transmembrane proteins containing a total of 19 transmembrane segments [CB21\_PEA, GPT\_GRILO, LECH\_HUMAN, FCE2\_HUMAN (SWISS-PROT codes), 1PRCH, 1PRCL (NRL3D codes)], two fibrous proteins [CH16\_DROME and ELS\_CHICK (SWISS-PROT codes)] and three globular ones [ADH1\_CHICK, ANGI\_CH-ICK, CONA\_CANEN (SWISS-PROT codes)]. The sequences

used, and other information concerning the application, are available on our web site at http://o2.db.uoa.gr/PRED-TMR2/ Results/).

Five different test data sets of transmembrane proteins with reliable topologies were collected from the literature. Test set 1 includes 64 sequences from the set of Rost *et al.* (1995) the sequences 2MLT, GLRA\_RAT, GPLB\_HUMAN, IGGB\_STRSP and PT2M\_ECOLI, which were not found in the public databases, were not used), sets 2 and 3 of 48 and 83 proteins respectively were taken from Rost *et al.* (1996), set 4 comprises the 44 sequences used by Cserzo *et al.* (1997) and set 5 is composed of 92 sequences from Fariselli and Casadio (1996).

A test data set of globular proteins was extracted from the Protein Data Bank (PDB), using the list of non-homologous sequences of PDBSELECT (Hobohm *et al.*, 1994). The 25% threshold list was used, excluding entries of membrane- and lipid-associated proteins (1AIJ, 1ALY, 1AR1, 1ATY, 1BEH, 1BHA, 1BQU, 1BXM, 1FTS, 1IXH, 1JDW, 1KZU, 1LGH, 1LML, 1NKL, 1OCC, 1PRC, 1QCR, 1SQC, 1TLE, 1×DT, 1YST, 2CPS, 2MPR, 2OMF, 2POR, 7AH1). This set of watersoluble proteins consists of 995 sequences.

#### *Calculation of amino acid residue transmembrane propensities (potentials)*

As described by Pasquier *et al.* (1999), a propensity for each residue to be in a transmembrane region was calculated using the formula

$$P_i = \frac{F_i^{\mathcal{I}M}}{F_i},\tag{1}$$

where  $P_i$  is the propensity value (transmembrane potential) of residue type *i* and  $F_i^{\text{TM}}$  and  $F_i$  are the frequencies of the *i*th type of residue in transmembrane segments and in the entire SWISS-PROT database respectively. Values above 1 indicate a preference for a residue to be in the lipid-associated structure of a transmembrane protein, whereas propensities below 1 characterize unfavorable transmembrane residues. The propensity values for the 20 amino acid residues are given in Table I.

#### Neural network topology and training parameters

The neural network used here has a multi-layer feed-forward (MLFF) topology. It consists of an input layer, one hidden layer and an output layer. Each of the units in the input layer are connected to all of the units in the hidden layer. The units in the hidden layer are then connected to all of the units in the output layer. This is a 'fully-connected' neural network where each unit *i* of a given layer is connected to each unit *j* of the next layer (Figure 1). The strength of each connection is given by a weight  $w_{ij}$ . The state *s* of each unit in the input layer is assigned directly from the input data, whereas the states  $s_i$  of higher layers *j* are computed by the sigmoid function

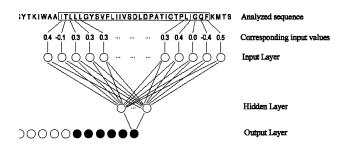
$$s_{j} = \frac{1}{1 + e^{-} \left( w_{j0} + \sum_{i=1}^{n} W_{ij} S_{i} \right)} , \qquad (2)$$

where  $w_{i0}$  is a bias from the states  $s_i$  of lower layers.

The network was trained using the backpropagation algorithm. During this process, a data set, describing the states  $s_i$  of the input units and their desired output value is presented to the network. The activations of the units of the network are

Table I. Propensity values and corresponding input used in the neural
network for the 20 amino acid residue types that belong to transmembrane
segments, calculated from the entire SWISS-PROT database

Residue		$P_i$	NN input
Phenylalanine	F	2.235	1.000
Isoleucine	Ι	2.083	0.929
Leucine	L	1.845	0.817
Tryptophan	W	1.790	0.791
Valine	V	1.756	0.775
Methionine	Μ	1.502	0.655
Alanine	А	1.383	0.599
Cysteine	С	1.202	0.514
Glycine	G	1.158	0.494
Tyrosine	Y	1.075	0.455
Threonine	Т	0.879	0.362
Serine	S	0.806	0.328
Proline	Р	0.597	0.230
Histidine	Н	0.395	0.135
Asparagine	Ν	0.389	0.132
Glutamine	Q	0.273	0.078
Aspartic acid	D	0.153	0.021
Glutamic acid	Е	0.131	0.011
Arginine	R	0.124	0.007
Lysine	Κ	0.108	0.000



**Fig. 1.** Schematic architecture of the neural network. Amino acids of the input sequence are converted to unique input values corresponding to the propensity for each amino acid to be located inside a transmembrane region (see Table I). Output of the network consists of values between 0 and 1. Values above 0.9 (shown in black on the figure) indicate a detection of a potential transmembrane segment.

then calculated, feeding forward layer-by-layer from the inputs to the output. Once the network output value has been produced, it is compared with the target output specified in the training data set. Following this comparison, a backwards adjustment of the weights (backpropagation) is performed in order to minimize the differences between the computed output and the desired output value. The algorithm is performed until the total error reaches a low enough value which means that the network comes to approximate the target values, given the inputs in the training set.

During the prediction phase, the neural network is fed with new input data that are not in the training set. By a simple feed-forward process, using the previously obtained weight, new output values are calculated and are taken as predictions of the network.

Applied to our classification problem, the idea is to use as input to the network a representation of a part of a sequence in order to obtain a unique output showing whether the analyzed segment is related to a transmembrane region or not.

The propensity values for the 20 amino acids given in Table I, which can be regarded as numeric representations of amino acids, are used to encode the input segment after being linearly transformed to lie within the range 0 to +1 (Figure 1). The output of the network consists of a unique value

between 0 and 1, which gives the propensity that the input segment is related to a transmembrane region. A high output value (greater than 0.9) is used to trigger the detection of a transmembrane segment. When at least one transmembrane region is detected, a protein is classified in the membrane protein class, otherwise it is put in the non-membrane protein class.

Experimentation determined the optimal size of the input layer to be 30. Training proteins were converted to input vectors by shifting a window of 30 residues successively through the sequence, i.e. the first segment contains amino acids from position 1 to 30, the *n*th segment encodes amino acids from position n to n+30. The training set was accordingly converted to 3140 input vectors of 30 values each.

Considering an input vector of 30 amino acids, we decided that it represents a significant transmembrane region if at least 10 amino acids in it belong to a transmembrane segment. This number was found by experimentation. Experimentation also determined the optimal number of hidden layers to be 1 and the number of neurons in this layer to be 2. The network was totally connected between adjacent layers (Figure 1).

The neural network was trained with the 3140 input vectors and their corresponding output values until convergence to a total error of less than 0.005.

#### Results

Using only information contained in 11 sequences from the training set, the neural network was able to generalize the processing of a test set with very good reliability. When applied to the five test sets of membrane proteins (see Materials and methods), the system gave a perfect prediction rating of 100% by classifying all the sequences in the membrane class. A total of 101 non-homologous proteins constituted the final test set (details are given at the Web address http://o2.db.uoa.gr/ PRED-TMR). Six proteins of the training set were included in the test set. Removing them from the test set, the neural network still predicted the remaining 95 proteins as membrane proteins (100% accuracy). For the test set of 995 globular proteins (see Materials and methods), the neural network predicted falsely 23 of them to be in the membrane class (97.7% of correct assignment). The proteins falsely classified were 1AGN, 1AMU, 1ARZ, 1AW8, 1BFD, 1BIB, 1BNK, 1CD1, 1DLC, 1FGJ, 1IHP, 1KVE, 1LXT, 1MAZ, 1NOX, 10VA, 1PS1, 1TAD, 1TAH, 1UAE, 1WER, 2ABK and 3R1R.

These results are good but they cannot be easily generalized to decide the predictive power of the method applied on real cases, like the classification of open reading frames (ORF's) identified in complete genomes. The test sets of membrane proteins indeed seem too limited and composed exclusively of proteins where reliable information about the location of transmembrane segments already exists. This does not necessarily reflect the composition of the complete genome. In addition, the proteins used in these sets contain only transmembrane  $\alpha$ -helices, which are easier to predict than transmembrane  $\beta$ -strand segments (Diederichs *et al.*, 1998).

Despite the errors contained in SWISS-PROT, it is thought that the annotations contained in this database can be used to automatically extract two sets of membrane and non-membrane proteins, which should be more representative of the composition of complete genomes. These sets can serve as a common test set that could be used for the rating and comparison of similar methods.

The set of membrane proteins extracted from the SWISS-

Table II.	Percentages of transmembrane	proteins	predicted by	/ PRED-TMR2
on seven	complete genomes			

Genome names	Percentage of TM proteins		
Escherichia coli	24.6		
Haemophilus influenzae	21.2		
Methanococcus jannaschii	19.8		
Mycoplasma genitalium	26.3		
Mycoplasma preumoniae	22.9		
Saccharomyces cerevisiae	28.0		
Synechocystis SP	26.5		

PROT database release 37 contains 10 743 entries. It has been built by selecting all the sequences containing the keyword 'TRANSMEMBRANE' and having at least one transmembrane segment annotated. All the remaining sequences in the database are not necessarily non-membrane proteins as some membrane sequences might not have been annotated yet. A reliable set was extracted by selecting, from the cluster of proteins not classified as transmembrane proteins, only sequences with a known three-dimensional structure (presence of the keyword '3D-STRUCTURE'). The set of non-membrane proteins contained 2280 sequences.

The neural network was applied to the sets of membrane and globular proteins collected. For the membrane proteins, it correctly classified 92.28% of them in the membrane protein set (9914 out of 10 743). Our neural network was also tested on proteins containing transmembrane  $\beta$ -strands (Diederichs *et al.*, 1998) and it produced disappointing results. For the globular water-soluble proteins, it correctly classified 93.38% of them (2129 out of 2280). The score obtained on this set of globular proteins is lower than the rating calculated on the set taken from PDBSELECT but it should be a good indicator of the validity of the extraction method. The ratio of 93% of correct assignment (both for membrane and non-membrane proteins) should be representative of the predictive power of the method when applied to complete genomes.

On the basis of these encouraging results, the neural network was associated with the PRED-TMR algorithm (Pasquier *et al.*, 1999) in a new application package called PRED-TMR2. This program can be used directly for the prediction of unknown proteins or on the ORF's predicted by the various genome projects.

It is true that this neural network does not provide evidence for the presence of N-terminal signal peptides. Methods exist capable of identifying signal peptides and predicting their cleavage sites (e.g. Nielsen *et al.*, 1997). If the set of membrane proteins (containing the keyword 'TRANSMEM') extracted from the SWISS-PROT database release 37, which contains 10 743 entries, is screened for the presence of signal peptides, 8558 sequences are found with no signal peptides. Of these, 7888 are correctly classified with the use of the neural network (92.17% accuracy).

PRED-TMR2 has been applied on seven complete genomes and on the entire content of the SWISS-PROT database. The percentage of membrane sequences predicted in each genome is given in Table II. The results range from 19.8% for *Methanococcus jannaschii* to 28% for *Saccharomyces cerevisiae*. Details of the results obtained can be downloaded together with the list of the transmembrane segment assignments from http://o2.db.uoa.gr/PRED-TMR2/Results/. The results have not been screened for the presence of N-terminal signal peptides.

#### Discussion

The prediction of transmembrane segments within proteins is a central problem of computational biology. A number of methods have been developed over the past 20 years. Some of them accomplish high accuracy and are available via the Internet [see, for example, Promponas et al. (1999) and references therein]. However, most of these methods are focused on the localization of transmembrane segments in known integral membrane proteins and produce a number of false segment detections when applied to globular watersoluble proteins. The rate of over-prediction is not well known as few works have been published on this subject. Two recent papers tackle the problem of identification of transmembrane proteins. Kihara et al. (1998) have tested their method on two sets of 89 transmembrane proteins collected from the literature and 928 globular proteins extracted from PDBSELECT. They announce a correct classification of 82 of the transmembrane proteins (92.13%) and of 836 of the globular ones (90.1%). Our neural network was found to perform slightly better than this method. Hirokawa et al. (1998) made the tests of their SOSUI system on a set of 92 transmembrane proteins listed by Fariselli and Casadio (1996) and 502 soluble proteins extracted from PDBSELECT and state that their system discriminated all sequences correctly, except for one in each set of data, resulting in an accuracy of more than 99%. Concerning the classification of transmembrane proteins, our method produced similar results as SOSUI: an accuracy of 100% was achieved on the same set and also on several other sets. For the globular proteins, SOSUI, with an incredible accuracy of 99.8%, seems to perform slightly better than our neural network, which was tested on a larger set (995 proteins) extracted from PDBSELECT with an accuracy of 97.7%. An execution of SOSUI on the 23 soluble proteins misclassified by our system results in three of them being assigned to transmembrane proteins (1bnk, 1cd1, 1kve). Even with these errors, the rating is still excellent and better than our method, assuming that all remaining sequences are correctly predicted by SOSUI.

The systems above do not use neural network systems for the classification. We show here that a simple and very fast neural network system can be successfully applied to this kind of problem. The novelty in our network topology is the small number of neurons and connections required. Most of the neural network systems presented so far use the same local encoding for each amino acid in a sequence (Qian and Sejnowski, 1988; Reczko, 1993; Rost *et al.*, 1995; Fariselli and Casadio, 1996; Aloy *et al.*, 1997; Diederichs *et al.*, 1998), i.e. each residue is represented by a vector of 20 or 21 values. The input layer of the networks using this encoding must be 20 times the size of the input segment. In the case of a window of 30 amino acids, this represents 600 neurons. In our system, each amino acid is encoded with a unique value and only two neurons in the hidden layer are used.

It is known that the successful generalization of a prediction by a neural network requires a much larger number of cases that the number of weights adjusted during the training phase. With our architecture, the total number of connections associated with a weight is only 62 (60 to connect 30 input neurons to the hidden layer and 2 from this layer to the unique output). This allows one to successfully train the network with information on the topology of very few proteins. In our application, the number of cases (3140) is larger than the number of weights by a factor of 50. In addition, the simple feed-forward topology of the network and its limited number of connections allow proteins to be processed very quickly and could open the way for a new implementation able to handle longer segments of amino acids and, perhaps, complete sequences.

A WWW server running the PRED-TMR2 algorithm is freely available at http://o2.db.uoa.gr/PRED-TMR2/

#### Acknowledgements

The authors gratefully acknowledge the support of the EEC-TMR 'GENE-QUIZ' grant ERBFMRXCT960019.

#### References

- Aloy, P., Cedano, J., Olivia, B., Aviles, X. and Querol, E. (1997) CABIOS, 13, 231–234.
- Bairoch, A. and Apweiler, R. (1998) Nucleid Acids Res., 26, 38-42.
- Cserzo, K., Wallin, E., Simon, I., von Heijne, G. and Elofsson, A. (1997) Protein Engng, 10, 673–676.
- Diederichs, K., Freigang, J., Umhau. S., Zeth, K. and Breed, J. (1998) Protein Sci., 7, 2413–2420.
- Fariselli, P. and Casadio, R. (1998) CABIOS, 12, 41-48.
- Hirokawa, T., Boon-Chieng, S. and Mitaku, S. (1998) *BioInformatics*, 14, 378–379.
- Hobohm, U. and Sander, C. (1994) Protein Sci., 3, 522.
- Kihara, D., Shimizu, T. and Kanehisa, M. (1998) Protein Engng, 11, 961-970.
- Nielsen,H., Engelbrecht,J., Brunak,S. and von Heijne,G. (1997) *Protein Engng*, **10**, 1–6.
- Pasquier, C., Promponas, V.J., Palaios, G.A., Hamodrakas, J.S. and Hamodrakas, S.J. (1999) *Protein Engng*, **12**, 381–385.
- Persson, B. and Argos, P. (1994) J. Mol. Biol., 237, 182-192.
- Promponas, V.J., Palaios, G.A., Pasquier, C.M., Hamodrakas, J.S. and Hamodrakas, S.J. (1999) *In Silico Biology*, 3, 1–4.
- Qian, N. and Sejnowski, T.J. (1988) J. Mol. Biol., 262, 865-884.
- Reczko, M. (1993) SAR and QSAR in Environmental Research, 1, 153-159.
- Rost, B., Sander, C. and Schneider, R. (1994) CABIOS, 10, 53-60.
- Rost, B., Casadio, R., Fariselli, P. and Sander, C. (1995) Protein Sci., 4, 521-533.
- Rost, B., Fariselli, P. and Casadio, R. (1996) Protein Sci., 5, 1704–1718.
- Sussman, J.L., Ling, D., Jiang, J., Manning, N.O., Prilusky, J., Ritter, O. and Abola, E.E. (1998) Acta Crystallogr., 54, 1078–1084.
- von Heijne, G. (1992) J. Mol. Biol., 225, 487-494.

Received February 22, 1999; revised April 16, 1999; accepted May 17, 1999