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***Decastronema kotori* gen. nov., comb. nov.:**
a mat-forming cyanobacterium on Cretaceous carbonate platforms
and its modern counterparts

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Lee SEONG-JOO³

Abstract: The fossil renamed here was first described in 1959 as *Aeolisaccus kotori* RADOIČIĆ, a new species of a problematic fossil worm, *Aeolisaccus* ELLIOTT. In 1975 DE CASTRO recognized the true relationships of this microbial fossil: a cyanobacterium related closely to the modern genus *Scytonema*. The fossil is common in the sediments of the Mesozoic carbonate platforms of southern Europe. This contribution confirmed DE CASTRO's interpretation, determined, using the high resolution of the SEM, the extent to which these fossils have preserved their original architecture, and investigated their presumed modern counterparts among the abundant mat-forming species of *Scytonema* on the intertidal flats of Andros Island, a part of the Bahama carbonate platform. The systematic affinities of the fossil and the environments it inhabited were reconstructed by comparing the morphology of the fossils to that of their modern counterparts, along with their respective sedimentary contexts. Based on these comparisons, we conclude that the organism lived in a peritidal environment and was buried and fossilized in the shallow waters of an ancient carbonate platform. A formal transfer of the fossil to a new genus of fossil cyanobacteria thereby designated as *Decastronema* gen. nov. is proposed, honoring the contribution of Prof. Piero DE CASTRO to paleontology.

Key Words: *Aeolisaccus kotori*, *Decastronema* n. gen., carbonate platforms, cyanobacteria, diagenesis, microbial fossil, Cretaceous

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Résumé : *Decastronema kotori* n. gen., n. comb., une cyanobactérie des tapis algaires des plates-formes carbonatées crétacées, et ses analogues modernes.- Ce fossile fut décrit pour la première fois en 1959 sous le nom d'*Aeolisaccus kotori* RADOIČIĆ, une nouvelle espèce rapportée à un vers fossile énigmatique, *Aeolisaccus* ELLIOTT. En 1975, DE CASTRO reconnaissait la nature microbienne de ce fossile, une cyanobactérie très proche du genre actuel *Scytonema*. Cet organisme est fréquemment observé dans les séries carbonatées des plates-formes mésozoïques d'Europe méridionale. Le présent travail confirme l'interprétation de DE CASTRO. L'examen au MEB montre la remarquable préservation de l'architecture originelle de ces fossiles et permet d'établir des rapprochements significatifs avec les nombreuses espèces modernes de *Scytonema* participant à l'élaboration des tapis algaires dans les zones d'estran de l'île d'Andros sur la plate-forme carbonatée des Bahamas. L'établissement des affinités taxinomiques des fossiles et la reconstitution des environnements qu'ils occupaient ont été effectués en comparant les morphologies des fossiles et de leurs analogues actuels, et en prenant également en considération leurs contextes sédimentaires respectifs. En s'appuyant sur ces comparaisons, nous concluons que ces organismes vivaient dans un milieu péritidal et qu'ils ont été enfouis, puis fossilisés en eaux peu profondes sur une plate-forme carbonatée ancienne. Nous proposons le transfert formel de ce fossile dans un nouveau genre de cyanobactéries fossiles créé pour cette occasion et nommé *Decastronema* n. gen. en l'honneur du Professeur Piero DE CASTRO pour sa contribution à la paléontologie.

Mots-Clefs : *Aeolisaccus kotori*, *Decastronema* n. gen., plates-formes carbonatées, cyanobactéries, diagenèse, fossile microbien, Crétacé

Sažetak: *Decastronema kotori* gen. nov., comb. nov., cijanobakterija mikrobijskih pokrova na karbonatnim platformama Krede i njeni recentni predstavnici.- Fossil, koji proučavamo je izvorno je opisan 1959. godine pod imenom *Aeolisaccus kotori* RADOIČIĆ, kao nova vrsta problematičnog fosila, *Aeolisaccus* ELLIOTT. DE CASTRO revidira sistematsku pripadnost tog fosila 1975.

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godine i priključuje ga u skupinu cijanobakterija srodnu recentnom rodu *Scytonema*. Fosil je obilno zastupljen u sedimentima mesozojskih karbonatnih platforma južne Europe. Ovim je radom potvrđena sistematska pripadnost fosila po DE CASTRU, i to primjenom elektronske mikroskopije (SEM) u proučavanju njegove gradje. Nadalje se istražuju mogući recentni srodnici među vrstama roda *Scytonema*, koje grade mikrobijske pokrove u zoni plime i oseke na otoku Androsu (dio Bahamske karbonatne platforme). Sistematska pripadnost i sredina u kojoj je mezozojski fosil živio je rekonstruirana na osnovi usporedbe morfologije fosilnih i recentnih oblika u okviru sedimentarnog konteksta. Na toj osnovi zaključujemo, da je organizam obitavao u mediolitoralnoj zoni, te da je prenesen, odložen i sačuvan u talozima plitkog mora drevne karbonatne platforme. Predlažemo formalni prijenos fosila k novom rodu fosilnih cijanobakterija pod nazivom *Decastronema* gen. nov. a u čast profesoru Pieru DE CASTRU za njegove znanstvene doprinose u području paleontologije.

Ključne riječi: *Aeolisaccus kotori*, *Decastronema* gen. n., karbonatne platforme, cijanobakterije, diageneza, mikrobijski fosil, Kreda

Introduction

The fossil record of cyanobacteria is rich throughout more than 2000 Ma of Proterozoic time (SCHOPF & KLEIN, 1992), but is comparatively poor for the 543 Ma of the Phanerozoic Era. Many Proterozoic microfossils are preserved in early diagenetic silica deposits. In contrast, most fossils of algae and cyanobacteria recovered from Phanerozoic strata are preserved in carbonates. Many of them were calcified during or immediately after their lifetime, which facilitated their preservation (RIDING, 1991; PENTECOST, 1991).

Many Proterozoic cyanobacterial microfossils have morphotypically close modern counterparts, thus testifying to an early

evolutionary origin, early ecological specialization and conservative maintenance of the basic adaptive properties of this group of the original oxygenic phototrophs (KNOLL *et alii*, 1986; KNOLL & GOLUBIC, 1992).

This contribution examines a microfossil common in southern Europe in the Cretaceous strata of Periadriatic carbonate platforms, which are sedimentologically comparable with the modern carbonate platforms of Florida and the Bahamas (see D'ARGENIO, 1970; D'ARGENIO *et alii*, 1975; RADOIČIĆ & D'ARGENIO, 1999; VLAHOVIĆ *et alii*, 2002). The fossil was originally described as *Aeolisaccus kotori* and attributed to the problematic fossil worm: *Aeolisaccus* ELLIOTT, 1958 (RADOIČIĆ, 1959). Based on a detailed study of morphology and the morphometric



Figure 1: Area of study, the frame of the map sheet Orahovac and the outlines of the countries superimposed on NASA World Wind satellite image.

evaluation of a large fossil population, DE CASTRO (1975) provided a convincing re-interpretation of this fossil as the remains of an ancient cyanobacterium. In support of his interpretation, DE CASTRO invoked possible modern counterparts among mat-forming cyanobacteria, described by MONTY (1967) from the mudflats on Andros Island, Bahamas. Here we investigate the state of preservation of this fossil using high resolution SEM imaging to compare it with the modern mat-forming *Scytonema* populations of Andros Island, Bahamas. Based on confirmed cyanobacterial identity, and in accordance with the rules of zoological nomenclature, we propose to change the name of this fossil.

Materials and Methods

Materials

R. RADOIČIĆ collected the fossiliferous rocks for this study in the Upper Cretaceous outcrop of Mt. Grebnik, Mirdita Zone, Serbia and Montenegro (Fig. 1). Fossil cyanobacterial fragments occur in great numbers in the lens-shaped deposits of fine-grained mudstone, wackestone and packstone immediately above paleokarstic bauxite deposits (Fig. 2). More were obtained from the collection of P. DE CASTRO, University of Naples, Italy. S. GOLUBIĆ collected modern specimens of *Scytonema* on the mud flats (with ponds and mangroves) on the west coast of Andros Island, Bahamas, mainly in the area bounded by the tidal creeks Polamasola and Three Creeks. The collected specimens were preserved both air-dried and in solution of 3% formaldehyde in environmental water.

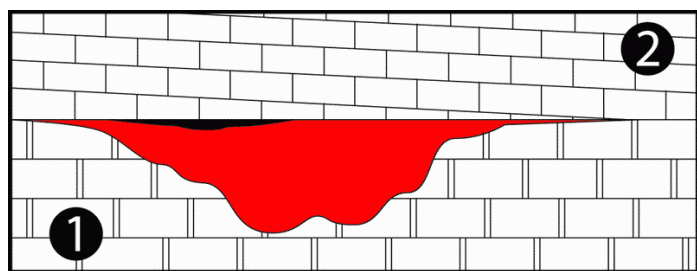


Figure 2: A simplified presentation of the geological setting of the *Decastronema* sampling site above the bauxite palaeokarst horizon of the Metohia, Mirdita zone, internal Dinarides. 1. Karstified Lower Cenomanian limestone: *Pseudorhapydionina dubia* (DE CASTRO), *Rotalia mesogeensis* TRONCHETTI, *Nummoloculina* sp. and other miliolids; 2. Three types of Santonian or Lowermost Campanian limestones were recognized: a) limestone with numerous Fe ooids and *Rotorbinella scarsellai* TORRE, discorbids, and rare *Clypeina dusanbrstinae* RADOIČIĆ; b) limestone with dispersed fertile ampullae of *Neomeris* and c) Limestone with foraminifera *Accordiella conica* FARINACCI, *Pseudocyclamina sphaeroidea* GENDROT, *Dicyclina* and *Nezzazatinella* sp.; red – bauxite; black - lentiform level, in the lower part with layers of densely packed and well sorted fragments of *Decastronema kotori*.

Preparations and Microscopy

Petrographic thin sections were prepared from compact fine-grained Cretaceous limestone cut both perpendicular to and parallel with to the bedding plane. They were examined using both transmitted and cross-polarized light microscopy. Other sections were subsequently polished, briefly etched with dilute HCl, washed in deionized water, dried, coated with gold-palladium and studied using scanning electron microscopy.

Specimens of modern *Scytonema* were washed in deionized water prior to analysis and mounted on slides for transmitted light microscopy. Selected samples were gradually dehydrated in an ethanol series and critical-point-dried using liquid CO₂. Whole and fragmented specimens were observed by SEM.

The identity of the fossil

The fossil was originally described (RADOIČIĆ, 1959) as minute thick-walled cylindrical bags closed at one end, ca 500 to 780 µm long, with 32 to 80 µm external and 10 to 24 µm internal diameters. It was ascribed to the genus *Aeolisaccus* ELLIOTT, 1958, under the name *A. kotori* RADOIČIĆ, 1959. A supplementary description with photomicrographs was published later (RADOIČIĆ, 1972) showing calcareous tubes with thick walls composed of a series of inserted conical units. DE CASTRO (1975) examined and measured large populations of this fossil in shallow-water Senonian carbonates from the Apenninic Carbonate Platform. He recognized that the peculiar architecture of the walls was comparable to that of modern scytonematacean cyanobacteria and deduced that this fossil was an ancient cyanobacterium.

When observed in petrographic thin sections under transmitted light the fossils appear as cylindrical tubules, each characterized by a dark wall and a clear lumen. The walls are cylindrical and smooth internally, but uneven externally (Fig. 3A). They are composed of funnels that diverge outward, and often have thin and undulating margins (Fig. 3B). This unique architecture can be examined by optical sectioning, or reconstructed from numerous transverse, oblique and longitudinal cuts through the fossils in thin section. The three-dimensional aspect of the series of inserted funnels is deducible from transitions between saggital and tangential sections (Figs. 3C-E). The fossil tubules, of different lengths and randomly oriented, are scattered in the sediment in densities ranging up to 100 tubules mm³. Such a distribution suggests that they were deposited as fragments, possibly after some transportation. They were never seen in growth position.

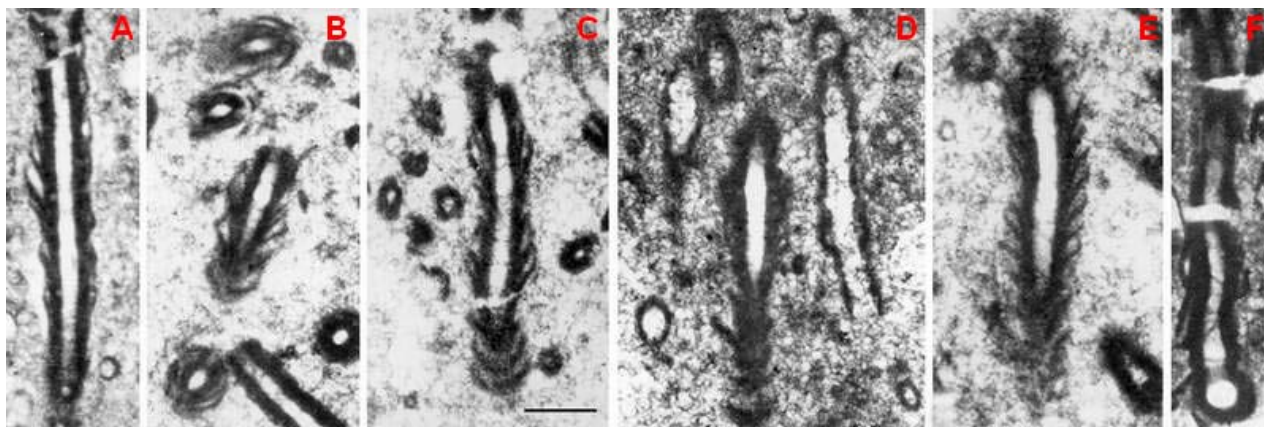


Figure 3: Petrographic thin section of *Decastronema kotori* (DE CASTRO) combinatio nova: (A) A filament in longitudinal section; note the thinning and bending of diverging outer layers of the wall; (B) Fragmented filaments of varied orientation in transverse and oblique sections; note the conspicuously clear lumina; (C–E) Filaments in nearly longitudinal section, becoming tangential at the lower end where the filaments curve out of the plane of the section; (F) Filament with "terminal chamber", representing a combination of longitudinal and transversal section at the position of a false branch (modified from DE CASTRO, 1975). Scale bar in C is 50 μm long for all pictures.

Longer tubules are sometimes branched, that is the clear central core (lumen) along with the inner layers of the wall penetrates laterally through the outer wall to form a branch. Several cases of this unusual 'inside out' branching pattern were seen and illustrated by DE CASTRO (1975, Pl. 5, figs. 1-9), who recognized them as false branching of the type characteristic of the filaments in the family Scytonemataceae.

State of preservation

The SEM images of polished and lightly etched preparations that expose the fossil structures in low relief show morphological properties the same as those visible in light microscopy (Fig. 4A). At a higher resolution the fossils appear to be constructed mainly of minerals. At magnifications ranging from 300 to 4000x the SEM images indicate that preservation of wall structure was accomplished in two ways.

In the more common mode of preservation, the fossil structure is expressed entirely by mineral grains (crystallites) with sharp euhedral boundaries (Figs. 4B-C). The walls of the tubules are comprised of very fine micrite with isodiametric grains ranging between 0.3 and 0.8 μm in diameter. The layers of the wall are made up of alternating relatively coarser (0.4-0.8 μm) and finer (0.3-0.5 μm) grains of micrite. Each layer is characterized by a different and uniform grain size of micrite with sharp euhedral boundaries (Fig. 4G). In cross sections of the filaments the layers are concentric (Fig. 4B), in longitudinal sections V-shaped (Fig. 4C). The lumen of the tubules is filled with larger grains 10-15 μm in diameter.

The second mode of preservation was found in specimens with walls of a reddish, rust-color when viewed with an incident light microscope. Under SEM, these walls evince divergent layers (Figs. 4A & D-F) like those of the other mode.

At higher magnifications the walls are not granular, but spongy with a fine, submicron level porosity (Fig. 4H). The reddish color is due to ferric oxides and hydroxides (limonite). The wall structure in these specimens is less affected by acid so it produces deeper relief as, for example, the conical shape of the layers in the three-dimensional display (Fig. 4I).

In both types of preservation, the lumen of the tubules is filled with relatively large micritic and microsparitic grains (cf. Figs. 4B & D), up to 20 μm wide and 40 μm long, commonly conforming to the internal diameter of the tubule (Fig. 4E). The clarity of the lumen observed in thin sections by light microscopy (Fig. 3) is explained by the greater conductance of light in the relatively large calcitic grains that occupy the lumen (Figs. 4E-F). Their large size suggests that the tubes were empty when the grains were deposited. The clear interiors of the tubes are in marked contrast to the dark appearance of the walls which is caused by light diffraction and scattering in the numerous small crystals.

The two modes of preservation are indistinguishable under transmitted light microscopy. Therefore, it is unlikely that the iron accumulated during the lifetime of the organism. The wall may have been enriched in iron postdepositionally by chelation with the organic matter associated with the sheath. Ferrous iron in solution may have leached from the nearby bauxite deposits, then transported to overlying beds in anoxic ground water and later oxidized to limonite.

The carbonate matrix surrounding the fossils is an irregular network of unsorted carbonate grains of different sizes with streaks of finer grains meandering around islets of larger sparitic grains (Figs. 4A & F). The size of the grains comprising these islets often show a progressive centripetal increase, a characteristic

of pore fills. This suggests that the fossil was deposited in a fine-grained porous sediment (possibly calcareous mud) and later diagenetically altered by cementation and pore filling. The mudstone - wackestone matrix of the fossils presumably was laid down in an oxidized low-energy shallow marine environment on an ancient carbonate platform. This interpretation is consistent with the lack of preserved organic matter and the absence of reduced minerals.

The abundance of these fossils ranges widely. They may have been distributed over a large area of shallow sea or accumulated in minor depressions on a mud flat. The site in the Mirdita zone of Metohija contains high concentration of fossils in a lens-shaped accumulation. Its position immediately above terrigenous bauxite deposits suggests early transgression. We assume that prior to deposition the fossils had been transported by tidal currents.

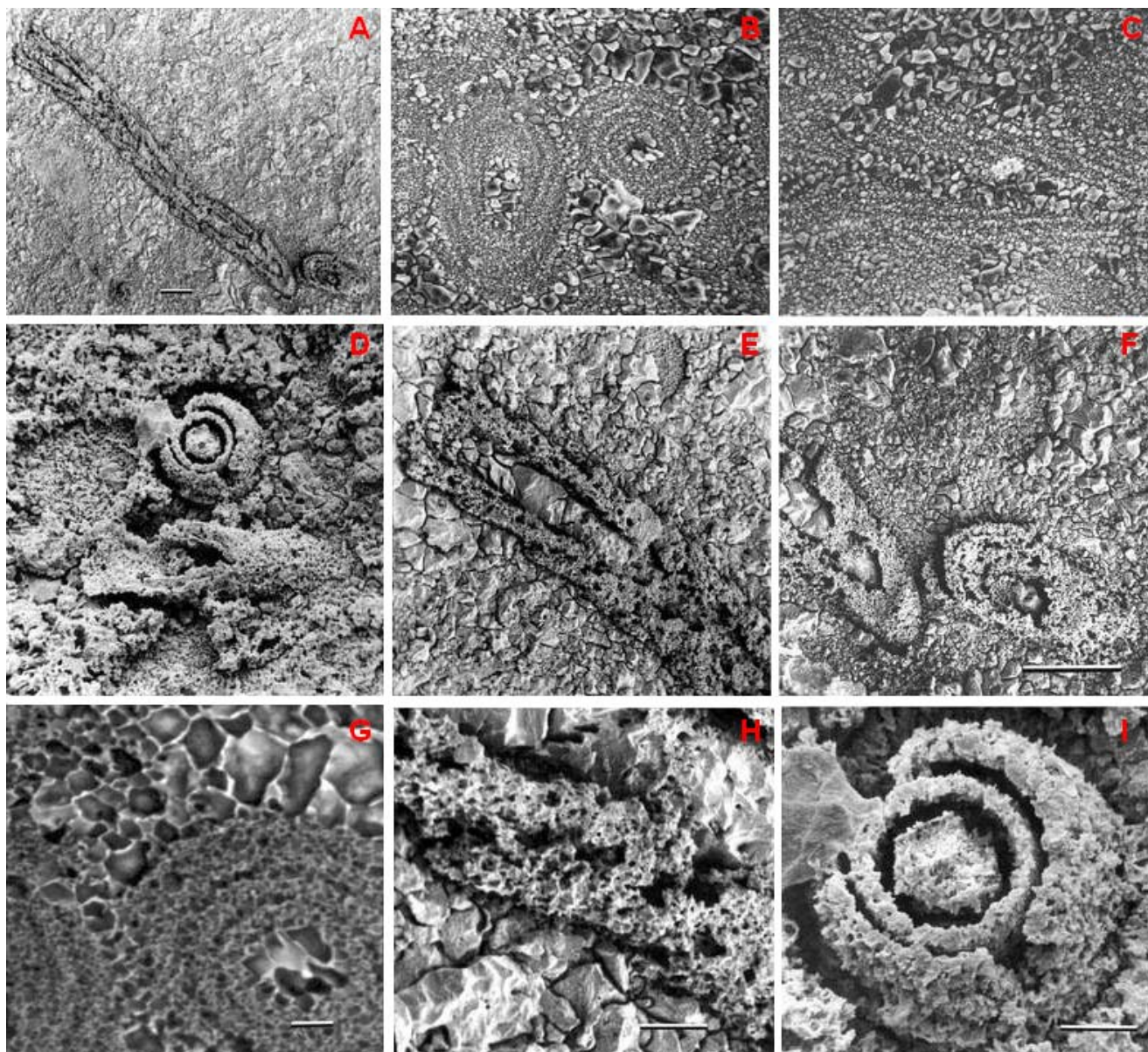


Figure 4: SEM images of *Decastronema kotori* on polished slightly etched rock surfaces: (A) View of a filament almost 1 mm long with upward diverging wall layers; (B-C) Oblique, transverse and longitudinal sections through filaments showing a concentric, upward divergent layering of the wall; the wall is comprised of micritic grains, whereas the lumen is filled with microsparite similar to that occupying pore spaces in the surrounding sediment; (D) Two fossil filaments etched from the matrix by prolonged application of acid. The wall appears spongy for it consists of a Fe-enriched non-crystalline material; (E-F) A similar etching of filaments in longitudinal and oblique sections; (G) Enlargement of a negative print of B, as negative, showing grain relationships in the micritically preserved sheath; (H) Detail of E, showing the spongy Fe-enriched texture of the wall; note the fine submicron-size porosity; (I) Detail of D: The upper filament, a nearly transverse section viewed from below, shows the core and two layers of the wall (comp. with Fig. 5B). The scale bar in A is 50 μ m long; the scale bar in F is 50 μ m long valid for B-F; the scale bars in G-I are 10 μ m long.

The modern counterparts

DE CASTRO (1975) recognized the unique architecture of the fossil, specifically the size, false branching and divergent layering of the wall, as characteristic of cyanobacterial sheaths. In search of the biological affinity of the fossil organism and its possible modern counterpart, DE CASTRO referred to MONTY'S (1967) study of modern microbial mats on Andros Island, Bahamas and considered among others a mat-forming cyanobacterium identified as *Scytonema myochrous*. Earlier, BLACK (1933) had described from that island two species with similar characteristics, but from a predominantly freshwater habitats: *Scytonema crustaceum* and *S. androsense*. However, DE CASTRO thought the fossil more closely related to the modern genus *Tolypothrix* because of the prevalence of single-sided false branching

We have studied the modern mat-building cyanobacteria on the mud flats of the west

coast of Andros Island, which is influenced by marine tidal circulation. Two species of *Scytonema* are common there. One inhabits a narrow range of the intertidal zone on the seaward side of the barrier beach, whereas the other covers vast surfaces around mangrove bushes on the ponded mud flats behind the barrier beach. The morphology of the mud-flat-dwelling species of *Scytonema* is closer to that of the Cretaceous fossil. The similarities include divergent sheath layering as well as the prevalence of single sided false branching, which like that of *Tolypothrix* is related to its upright growth (Fig. 5A). The filaments consist of thick sheaths harboring much narrower cellular trichomes in their core. The outer margins of V-shaped layers show externally as barely visible rings. The relationship between cellular trichome and layered sheath visible in fractured specimens (Figs. 5B-C) is similar to the architecture of the walls of the Cretaceous fossil (compare Fig. 4I).

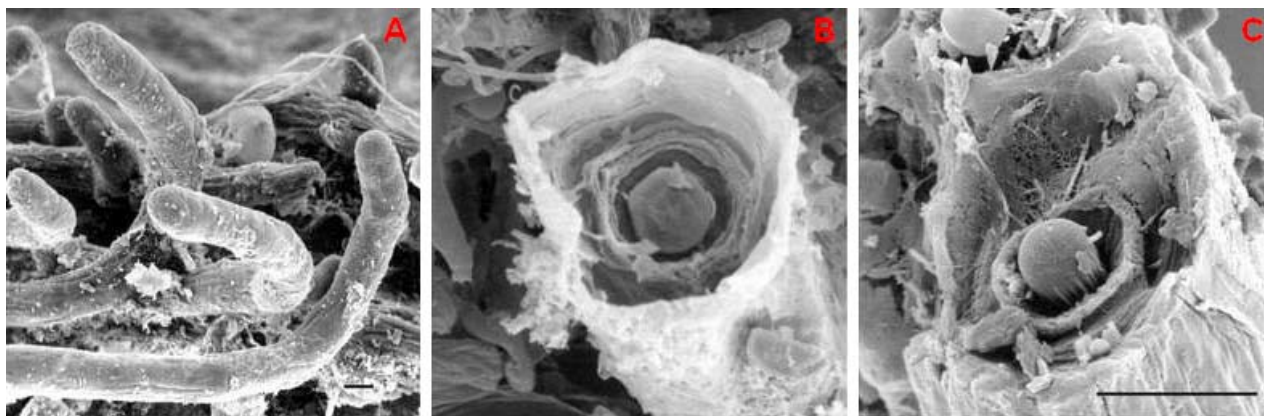


Figure 5: SEM of critical-point-dried specimens of Recent (modern) *Scytonema* sp. from the mats covering intertidal carbonate mud flats on the west coast of Andros Island, Bahamas. (A) Intact, upward curving filaments; the surface shows no appreciable carbonate precipitation. (B) View down a fractured filament of *Scytonema* showing the concentric arrangement of layered sheath enveloping a trichome cf. Fig. 4I. (C) Oblique view of another fractured filament with a still turgescient cell in the center of concentric sheath layers. Note the reticulate texture of the polysaccharide in the upper part of the picture. Scale bars are 10 μ m long; the bar in C is valid for B.

Thick envelopes and sheaths produced by copious amounts of extracellular polymeric substances (EPS) are common, particularly in subaerially growing cyanobacteria. However, the characteristic sheath architecture with open, upwardly divergent layering is particular to the section *Myochrotes* within the genus *Scytonema* (BORNET et FLAHAULT, 1887). Trichomes of *Scytonema* have the highest rates of cell division and sheath excretion at their tips, thus forming apical meristems; both decrease basipetally as the cells grow older. The newly excreted portion of the sheath encloses several actively growing cells below the apex (Fig. 6A). Due to apical growth, cell division and the accompanying production of EPS move in an acropetal direction. Consequently, the envelopes assume the asymmetrical shape of inverted cones (Fig. 6B). The angle of these cones is a function of the rate at which the trichome elongates: it is more

divergent when the growth is slow, and almost parallel in fast-growing aquatic forms (BHÂRADWÂJA, 1934). The pressure of the growing trichomes causes consecutive bursting and opening of envelopes at the trichome tip so the continuing growth produces a series of characteristic, funnel-shaped collars (Fig. 7, below). This feature has a potentiality for preservation and thus for recognition in the fossil record (see GOLUBIC & BARGHOORN, 1976). In addition, nitrogen fixation in these organisms, performed by specialized cells called heterocysts, stimulates growth in the middle of the trichome, forming intercalary meristems. Differentiated heterocysts do not produce EPS and act as anchoring points for trichomes, restricting their gliding inside the sheaths (GOLUBIC *et alii*, 1996). As a consequence, the trichomes protrude through the old sheath and continue to grow as false branches.



Figure 6: Filament tips of modern *Scytonema* sp. under light microscope showing the development of divergent sheath collars: (A) The tip of the trichome excretes layers of EPS that first stretch and then break as the trichome extends. (B) Sheath production is most intense at the tip and less basipetally. The lower parts of the trichome are enveloped by a series of inserted cones. Note the thinning of envelope margins in both pictures. The scale bar in A is 10 μm for A and 30 μm for B.

The presence of these same properties in the fossil *Decastronema* filaments, in particular the collar-like opening of sheath layers in conjunction with false branching, as well as an overall similarity in shape and size demonstrates unquestionably that DE CASTRO's interpretation and identification of this organism as an ancient cyanobacterium is valid.

Using light microscopy, DE CASTRO (1975) saw in longitudinal sections of *Decastronema* tubules, faint dark lines across the lumen. He interpreted these lines as the possible cross walls of the cellular trichomes, which would imply cell lengths of up to 30 μm . Our use of cross-polarized light microscopy and SEM images resolved these lines as grain boundaries between microsparitic grains that filled the tubes, often in a single row (Figs. 4D-E). At the level of resolution used in this study no remains of trichome cells or cross walls separating them were found.

The meristematic (*i.e.* rapidly dividing) cells in modern *Scytonema* are short and wide. As the rate of division slows, the cells grow longer and narrower, accompanied by a loss in turgescence. The consequence is a wide variability in the dimensions of the cells, which is consistent with the considerable variability of the internal diameters of *Decastronema* tubules.

The appearance of one-sided 'closure' reported in the original description (RADOIČIĆ, 1959) exists also in modern *Scytonema*, but is limited to growing tips and young false branches. This condition is explained by the life cycle and mode of reproduction of these

filamentous cyanobacteria, during which trichomes fragment to produce and release short segments called hormogonia, which then open and leave the sheath. A similar feature was described as a 'terminal chamber' in *Decastronema* and interpreted as a possible heterocyst (DE CASTRO, 1975, Pl. 6, Fig. 1). Actually, it represents a section through a curved or falsely branched filament of which a part was sectioned longitudinally and the other part transversally (Fig. 3F).

The habitat of the coastal *Scytonema* of Andros Island is an intertidal mudflat, flooded by meandering tidal creeks and periodically flushed by seawater during storms. Freshwater comes from rain and the drainage of water. Thus, salinity fluctuates from brackish to hypersaline. The organisms endure periodic shortages of water and an excessive solar illumination. Their thick sheaths may slow down the loss of water; they also contain the dark pigment scytonemin which provides a modicum of protection against damage from excessive light and UV radiation (GARCIA-PICHEL, 1998).



Figure 7: Two species of *Scytonema* growing on cliffs wetted by freshwater. They show distinction in size, sheath construction and the ability to support carbonate precipitation. The smaller one (left) is *Scytonema julianum* with sheaths encrusted with calcite crystals, the larger one (right) is *S. myochrous* producing divergently layered non-calcifying sheaths. Scale bar is 10 μm .

We consider this organism to be the closest modern counterpart of *Decastronema*. The fossil must have had ecological requirements very much like those of the modern species. Therefore we propose a similar habitat for the Cretaceous form.

Taxonomic revision

In 1975 DE CASTRO determined that the microfossil he was studying was misidentified and that its true taxonomic affinity is among the cyanobacteria. This called for a new nomenclatural combination with an assignment

to a different genus. However, in view of the remaining uncertainties in the identification of the fossil, DE CASTRO hesitated "to transfer the species *Aeolisaccus kotori* from a 'genus' to which it certainly does not belong to another to which it might not belong." (...) "For the moment," he concluded, "it seemed preferable to indicate the species under study by the name assigned to it by RADOIČIĆ, at least until further information permits to specify the existing doubts".

Here we confirm DE CASTRO's interpretation of *Aeolisaccus kotori* as fossilized sheaths of an ancient scytonematacean cyanobacterium. Our detailed investigations give us added confidence, so we propose a separate generic identity for this now well-defined microbial fossil, placing it in the Phylum (Division) Cyanobacteria, Order Nostocales, Family Scytonemataceae as:

***Decastronema*, new genus**

Etymology: In honor of Professor Piero DE CASTRO, of the University of Naples, Italy, for his valuable contributions to paleontology.

Diagnosis: Tubular filaments with bright core and dark walls, comprised of divergent and externally tapering layers arranged like a stack of inserted cones. The layers appear concentric in transverse and V-shaped in longitudinal section.

Iconotype: Figs. 3B-C & E

Type species: *Decastronema kotori* (RADOIČIĆ) *combinatio nova*

Description: Filaments and filament fragments with bright lumen and dark walls (as observed in transmitted light in petrographic

thin sections), preserved mostly as tubules with walls comprised of divergent and externally tapering layers (Figs. 3A-F). The remains of the walls are preserved in a distinctive grain arrangement (Figs. 4B-C & G) or by iron-rich, spongy textures (Figs. 4A, D-F & H-I), rust-colored in polished slabs and petrographic thin sections. The tubules of *D. kotori* have an internal diameter of 14.2 ± 4.05 (330) μm expressed as Mean \pm Standard Deviation (n). The walls appear dark and thick in petrographic thin sections, with an external diameter averaging 55.6 ± 12.5 (330) μm (recalculated after DE CASTRO, 1975), characterized by divergent layers that in three dimensions form as a stack of inserted funnels. The outer margins of the divergent layers are often thinned and curved (Figs. 3A-C). Most tubules are short, randomly distributed fragments open at both ends. Larger fragments are often branched, which in well-preserved specimens can be recognized as being one-sided false branches -- the core and inner layers of the wall protrude through a lateral break and perforate the outer layer of the main filament to form a branch.

Decastronema kotori is interpreted as the fossil remains of a large filamentous microorganism, which produced and dwelt in tubular sheaths made of extracellular polymeric substances (EPS, polysaccharides with some polypeptides). The architecture of the sheaths is consistent with the unique mode of growth, development and differentiation in a small group of modern species among the heterocystous cyanobacteria assigned to the genus *Scytonema* of the section *Myochrotes*.

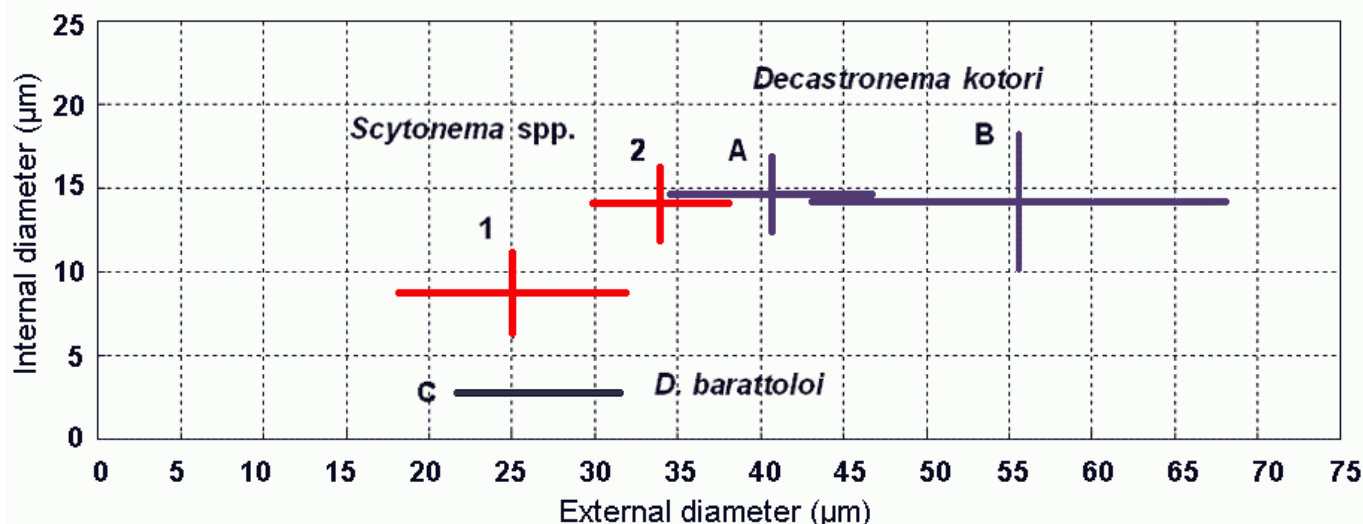


Figure 8: Plot of the size relationships between external diameters (x) vs. internal diameters (y) of filaments in fossil populations of *Decastronema* (A-C) and Recent populations of *Scytonema* from Andros Island, Bahamas (1-3). Mean values plot at the point of intersection in each of the cross diagrams, with arms extended for one standard deviation on each side of the mean. The total range of data for each population is inside a field of two standard deviations on each side of the mean.

A comparison of the fossil with the modern representatives of that genus reveals that both employ a unique and complex mechanism to construct the upwardly divergent sheath of extracellular polymers and false branches. That mechanism involves cellular polarity, differentiation among cells to perform specialized functions and the localization of zones of active growth (meristems). Intercalary N-fixing heterocysts anchor the trichomes in the sheaths, so that their further growth produces false branches. Several fossil and modern taxa of similar morphology exist. Their representative populations range widely in size (Fig. 8).

D. kotori is common in Santonian and lower Campanian sediments, often extraordinary abundant, usually in association with *Thaumatoporella parvovesiculifera* RAINIERI, less commonly with small foraminifera. It has also been reported in rocks ranging in age from Aptian to Paleocene (DE CASTRO, 1975) and found widely distributed in the Apennines (DE CASTRO, 1975), Dinarides (Adriatic Islands, Dalmatian coast, the external Dinaridic domains; RADOIČIĆ, 1959; GUŠIĆ & JELASKA,

1990) and Helenides (FLEURY, 1980).

Comment: The transfer of *Aeolisaccus kotori* RADOIČIĆ to the new genus *Decastronema* is based on a correction of the attribution of this particular fossil. The legitimacy of the genus *Aeolisaccus* and its type species *A. dunningtoni* (ELLIOTT, 1958) is not questioned here. But other species of that genus have been reclassified as foraminifera, e.g. *A. tintinniformis* MIŠIĆ, 1971. *A. ampliformis* PANTIĆ, 1972, has been assigned tentatively to genus *Erlandia*, and the systematic position of *A. inconstans* (RADOIČIĆ, 1967) and *A. gracilis* PANTIĆ, 1972, remains uncertain (see ZANINETTI, 1976). We consider the properties of *Decastronema kotori* unique and defined well enough to merit recognition as a discrete genus regardless of the taxonomic treatment of other species of *Aeolisaccus*. That *Decastronema* is related to the modern *Scytonema* is very probable. However, because of objective limitations imposed by fossilization on the criteria for taxonomic identification, we do not recommend that the names of extant biotaxa be used for fossils.

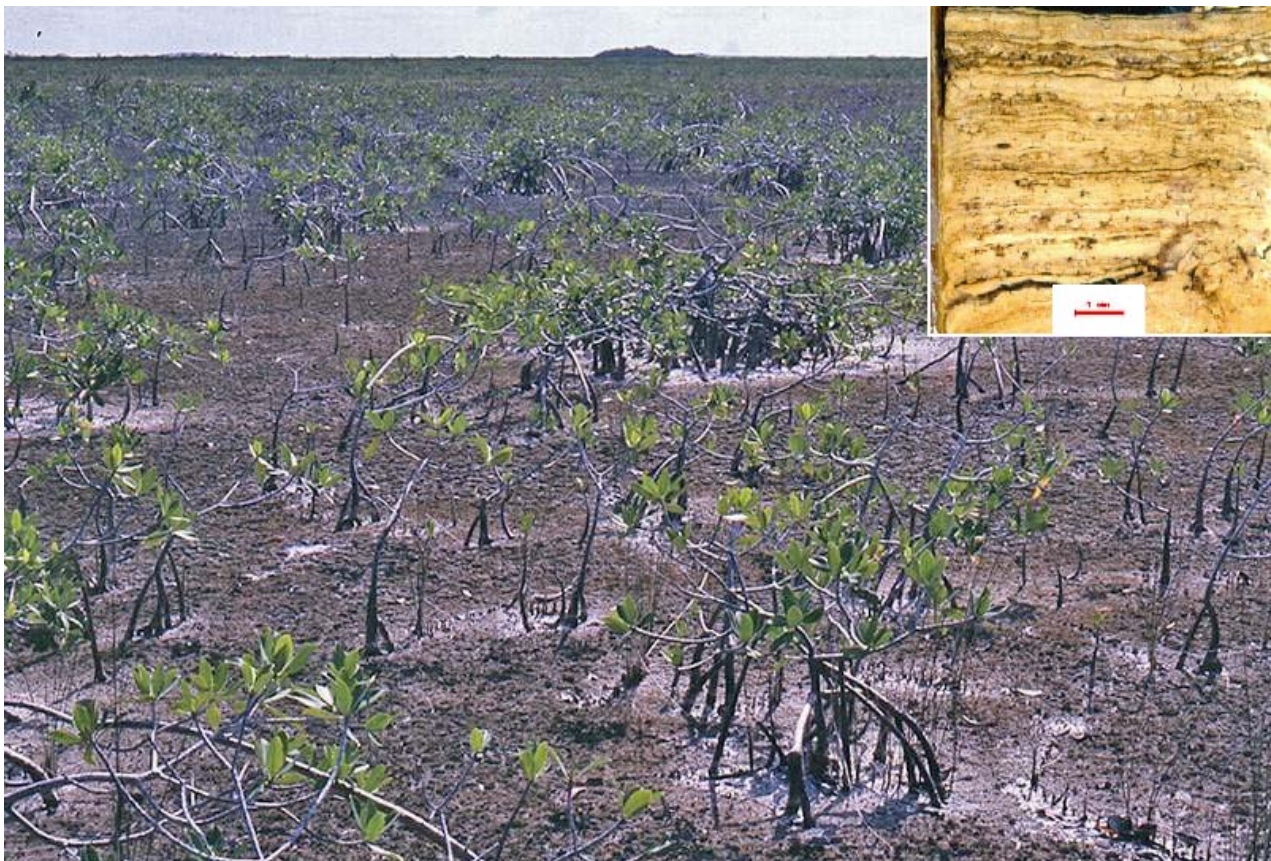


Figure 9: *Scytonema* mats around mangrove bushes on Intertidal flats of Andros Island, Bahamas form a dense brown cover over white carbonate mud. Inset: Core taken from the same area showing dark *Scytonema* fragments interlayered by carbonate mud.

The proposed revision does include those species of *Aeolisaccus* that have been identified as cyanobacteria. In addition to *Decastronema kotori*, this involves only one species: *Aeolisaccus barattoloi* DE CASTRO, 1989, which is reassigned here as:

***Decastronema barattoloi* (DE CASTRO)
comb. nov.**

This fossil shares most morphological characters with *D. kotori*. It too consists of hollow tubular segments, but is smaller and has thinner walls. The outer diameter is given as less than 33 µm. It ranges from the Maastrichtian, where it is most abundant, to the Danian, across the K/T boundary (DE CASTRO, 1989; BARATTOLO, 1998).

Comment: The existence of discrete populations of *Decastronema* with a broad spectrum of sizes is consistent with data on Cretaceous material from various sites and with the speciation of similar forms in comparable modern environments (Fig. 8).

Calcification and preservation potential of cyanobacterial sheaths

The fossil *Decastronema kotori* is a mineral replication of the exopolymer product of an ancient cyanobacterium. This fact raises a question regarding the nature of the process and the timing of sheath calcification and replacement.

An impressive fossil record shows that the potential for preservation of uncalcified cyanobacterial sheaths and envelopes is greater than that of the cells that produced them. The prokaryotic cells, enclosed during lifetime by an osmotically responsive plasma membrane, confined by cross-linked peptidoglycan walls, tend to collapse, shrivel and deform after death, while their sheaths contract in an orderly manner without change in shape as they dehydrate (GOLUBIC, 1980). Thus, Proterozoic cyanobacteria, embedded and preserved in silica, consist mainly of extracellular envelopes and sheaths (KNOLL & GOLUBIC, 1979; SCHOPF & KLEIN, 1992; SEONG-JOO & GOLUBIC, 1998; GOLUBIC & SEONG-JOO, 1999), that sometimes retain pigmentation that may have screened UV-rays (GOLUBIC & HOFMANN, 1976). This pigmentation may have been a significant protective agent during Proterozoic and Archaean times (GARCIA-PICHEL & CASTENHOLZ, 1991; GARCIA-PICHEL, 1998). However, when embedded in a carbonate, Proterozoic microbial fossils are rarely preserved in as much detail (see KNOLL & SEMIKHATOV, 1998) and suffer more commonly from diagenetic recrystallization that obliterates the original structure (HOFMANN, 1976).

In contrast, most Phanerozoic fossils exist as skeletal body parts preserved in carbonates.

Well preserved non-mineralized cyanobacterial fossils embedded in silica are exceptionally rare (e.g. TAYLOR *et alii*, 1997; CHACÓN-BACA *et alii*, 2002; KERP, 2002). Therefore, it is probable that the overall change in the fossil record from Proterozoic to Phanerozoic was to some extent influenced by a concurrent change in the predominant sedimentary environment.

Protists, animals and calcareous algae produce elaborate skeletal morphologies through enzymatically controlled intracellular or intercellular processes. Such precise cell-control systems are unknown in prokaryotes. Nevertheless, calcification can be promoted or inhibited by prokaryotic metabolic activities and products (CHAFETZ & BUCHINSKI, 1992; DÉFARGE & TRICHET, 1995). These biogenic influences can modify the mineralogy, size and arrangements of the precipitates, resulting in species-specific patterns (GOLUBIC & CAMPBELL, 1981; COUTÉ, 1982; OBENLÜNESCHLOSS, 1991; FREYET & VERRECCHIA, 1998). Among the possible mechanisms for the preservation of *Decastronema* by calcification are the following:

1. The sheaths were calcified during the life of the organism, along the lines of the model described for *Scytonema* in the waters of the Everglades, Florida (MERZ, 1992; MERZ-PREISS, 2000). Some, mainly freshwater and subaerial species, calcify (e.g. *Scytonema julianum*), while others (e.g. *S. myochrous*) do not (Fig. 7), even when growing together under the same conditions (GOLUBIC *et alii*, 2000). DE CASTRO selected as his model the modern *S. myochrous*, which was found calcified under supersaturated slightly brackish conditions on Andros Island (MONTY, 1967). Such calcification is often localized on segments of filaments and so may contribute to their fragmentation. Although this type of primary calcification may have occurred in *Decastronema*, no evidence for it has been retained.
2. *Decastronema* may have become calcified *post mortem* in conjunction with bacterial sheath degradation, as discussed by ARP *et alii* (1999), with the possible involvement of sulfate reduction and micrite replacement of EPS (VISSCHER *et alii*, 2000; PAERL *et alii*, 2001). However, in *Decastronema* the accuracy and precision with which the micritic grains outline the layers of EPS is markedly different from the clumpy, irregularly distributed micritic peloids associated with bacterial decay as illustrated by SPRACHTA *et alii* (2001) and DUPRAZ *et alii* (2004).
3. In *Decastronema* the close relationship between the patterns of distribution of micritic grains and the structural

properties of the sheath is in general agreement with the concept of 'organomineralization' *sensu* TRICHET & DÉFARGE (1995), in which an organic product guides the precipitate at the stage of crystal nucleation, while the preceding chemical induction of precipitation may be either abiotic or biogenic (GAUTRET *et alii*, 2004). BRIGGS (2003) discusses similar arrangements of uniform crystal size in diagenetic permineralization and preservation of soft-bodied tissues as analogous to the pixel resolution of computer images. This analogy is applicable to the wall texture of *Decastronema*. However, the permineralizations described by BRIGGS took place in the presence of high concentration of decaying organic matter. That environment makes it difficult to separate the effects of bacterial activities (induction) from those of bacterial products (templating). MORSE & WANG (1996) offer theoretical models of such processes and suggest that the formation of distinct fields of uniform-sized crystals may be either flux- or substrate-controlled. Both types of control may apply for the mineralization of cyanobacterial sheaths. BRAISSANT *et alii* (2003) showed that varying concentrations of polysaccharide and amino acids to form precipitates either with or without bacteria produced an array of discrete forms of crystals. Moreover, control of crystal size has also been achieved with artificial polymers (D'SOUZA *et alii*, 1999).

We conclude that in *Decastronema*, templating on the polysaccharide matrix of sheaths is the mechanism most likely to have determined the sites of crystal nucleation, for the patterns of calcification correlate closely with the sheath architecture. We suggest that calcification was a part of post-burial diagenetic mineralization (TURNER *et alii*, 2000; PRATT, 2001). In view of the known durability of cyanobacterial sheaths, it is conceivable that the pattern of grains may have persisted through more than one diagenetic recrystallization. This supposition is supported indirectly by the fact that a subset of fossil *Decastronema* from the same site showed permineralization with iron rather than by calcium enrichment (Figs. 4D-F).

The above interpretation does not preclude primary calcification that for Cretaceous microfossils may have been calcitic, because the ocean chemistry of the time was different from that of the modern ocean and calcification then favored calcite over aragonite (STANLEY & HARDIE, 1998). Primary calcite incorporated into the sheaths of *Decastronema* may have provided some initial stability to the organo-mineral relationship that was modified

subsequently.

RIDING (1994) noticed a positive correlation throughout the Phanerozoic between increases in the occurrence of calcified cyanobacteria and rises in abiotic carbonate precipitates. He proposed a model that distinguishes episodes of enhanced vs. reduced cyanobacterial calcification. The occurrences of *Decastronema* do not appear to conform to that model's prediction for these calcified filaments were most numerous during late Cretaceous times, a period during which the rate of calcification in cyanobacteria is said to have decreased. At that time massive skeletal calcification in plankton (coccolithoforids and foraminifera; KNOLL, 2003) and benthos (*e.g.* rudists), may have competitively lowered environmental carbonate saturation levels. However, such changes in seawater chemistry would have mattered less if the calcification of *Decastronema* occurred post-depositionally.

Biocoenosis or thanatocoenosis

The known occurrences of *Decastronema* are randomly oriented filaments and filament fragments in shallow marine carbonates associated with marine calcareous algae and foraminifera, and commonly with the dasyclad *Thaumatoporella* (BARATTOLO & BIGOZZI, 1996). The fossils are in fine-grained limestone, its grains ranging from micrite to microsparite. This lithology and association suggests that the sedimentary environment was entirely marine, but very shallow and at extremely low energy levels (DE CASTRO, 1975). The variability in abundance and the random orientation of the *Decastronema* fragments suggest that the fossils were transported prior to deposition, and so are very probably an allochthonous element in the thanatocoenosis in which they are usually found. In addition, the fossil was never seen in growth position. Even when densely packed, the broken filaments were deposited as randomly oriented clasts. The Mirdita population, for example, accumulated in lens-shaped depressions near the contact between overlying back reef lagoonal carbonates and paleokarstic bauxite.

The comparison with modern *Scytonema* supports a subaerial habitat for the fossil. Today, thick-layered sheaths are characteristic of subaerial species of *Scytonema*, whereas the sheaths of submersed species are thin (BHĀRADWĀJA, 1934). So the habitat of *Decastronema* with its thick and divergent sheaths was probably subaerial.

Modern *Scytonema*, considered a likely counterpart of *Decastronema*, grows on intertidal flats. It forms extensive mats on the west coast of Andros Island, Bahamas, covering many square km around interspersed bushes of *Rhizophora mangle* (Fig. 9). Periodic storms often disrupt and displace these intertidal *Scytonema* mats and bury them in layers of

fine-grade carbonate mud. Short cores taken from the mud flats (Fig. 9, inset) show dark layers of dislocated and randomly oriented *Scytonema* filaments between layers of carbonate mud with a few complete colonies preserved (R.N. GINSBURG, personal communication). However, disrupted and fragmented *Scytonema* filaments are also exported by tides and distributed over a much larger area where these fragments are buried in a shallow subtidal setting (Fig. 10). So we assume that the Cretaceous *Decastronema* lived in an intertidal habitat, similar to that on the west coast of the Andros Island (Fig. 11) but after death was transported to adjacent shallow areas of the carbonate platform where it was buried and fossilized.

Changes in carbonate platforms and their microbiota

Here we have compared cyanobacteria of similar structure and comparable habitat separated by 60-70 million years of geological time. Andros Island is the largest of a number of emergent portions of the Bahama carbonate

platform, surrounded by extensive areas of shallow waters not more than a few meters deep. The sediment build-up on the platform, continuous since Mesozoic times, shows that its construction kept abreast of repeated changes in sea level. In southern Europe where *Decastronema* lived the development of carbonate platforms of similar age and lithology was interrupted by Alpine orogeny following the collision of African and Eurasian continental plates. The carbonate environments compared here may have had a common origin as parts of a series of Mesozoic platforms along the southern margins of the Tethys Sea (D'ARGENIO, 1970). The western portion of these platforms escaped destruction by Alpine orogenesis as the Atlantic Ocean widened, and continued to exist on the stable trailing end of the North American continent. Contemporaneous strata on both platforms were analyzed and found to be lithologically identical (D'ARGENIO *et alii*, 1975). Here we add paleontological support to the similarities in lithological and paleoenvironmental properties reported independently by others.



Figure 10: Aerial view of tidal creeks on the NW coast of Andros Island, Bahamas, rimmed by *Scytonema* cover (reddish brown) around bright carbonate mud in shallow ponds.



Figure 11: Westward view of the ponded intertidal mud flats of Andros Island, a possible analogue of the environmental setting of the Cretaceous *Decastronema*.

Stratigraphic vs. paleoenvironmental considerations

Decastronema kotori was found first in the Turonian rocks (Upper Cretaceous) of the external Dinarides (RADOIČIĆ, 1959). Subsequently it was found in strata ranging from uppermost Aptian to Paleocene, with a maximum abundance in the Senonian (reviewed by DE CASTRO, 1975). Although the organism survived the massive extinctions at the Cretaceous-Tertiary transition, it has not yet been encountered in earlier or later deposits. This is unusual for a member of cyanobacteria, which as a group diversified early in the Proterozoic and persisted over time while maintaining their conservative phenotypic properties. Heterocystous cyanobacteria, of which *Decastronema* is one, were reported from Devonian Rhynie chert (KIDSTON & LANG, 1921), but may have evolved during the Mesoproterozoic, between 1300 and 1500 million years ago, as suggested by the findings of the fossil *Archaeoellipsoides*, interpreted as akinetes of heterocystous cyanobacteria similar to *Anabaena* (GOLUBIC *et alii*, 1995).

The peculiar divergent layering of the sheath of *Scytonema* and *Decastronema* along with the false branching of filaments is a stage in the evolution of filamentous cyanobacteria during which the cells differentiate to assume different

forms and functions: The 'vegetative' trichome cells fix carbon and grow in localized meristematic zones, the heterocysts fix nitrogen and the akinetes store nutrients and act as resting spores. These properties in modern cyanobacteria serve as phylogenetic markers. The interpretation of the architecture of the Cretaceous fossil is consistent with the presence of these elements in modern heterocystous cyanobacteria.

The time constraints on the range of *Decastronema* may be environmental as well as evolutionary. If our conjectures based on comparisons with modern *Scytonema* are correct, the fossil lived on the intertidal flats adjacent to land. The intertidal areas are normally very narrow strips of the coast. Consequently it is possible that the fossil has been overlooked in stratigraphic surveys due to its narrowly defined ecological niche. Environments within the intertidal range are especially vulnerable to frequent changes in sea level that limit recording of the occurrence of intertidal biota to episodes that geologically are very short. Short-term sea level oscillations that follow MILANKOVIĆ cycles, as recorded in microstratigraphic analyses (LONGO *et alii*, 1994; D'ARGENIO *et alii*, 1992, 1997; BUONOCUNTO *et alii*, 2002), may have caused many repeated short exposures and submersions on carbonate platforms as well as significant lateral migration

of habitats and their biota. Each such change, although it has an ecologically dramatic impact, is no more than an instant on the geological time scale.

The long-term survival of shallow water biota on carbonate platforms requires that sedimentation be successful in matching subsidence. Their demise may be caused by drowning, as was the case with mid-oceanic platforms (CAMOIN *et alii*, 1998), or by major tectonic changes associated with continent collisions, as was the case with the Periadriatic carbonate platforms. Carbonate platforms such as the Bahamas that have existed for a long time are the exception. Their longevity is due to the relative stability of conditions on the trailing side of the continent.

Conclusion

Microorganisms on carbonate platforms have been subjected to significant long and short-term changes in environment. Prokaryotes such as cyanobacteria have responded by adaptations to environmental instabilities and have re-occupied their niches after catastrophic changes, including periods of major extinctions. Their record as fossils continues unbroken although it is accompanied by changes in the composition of successful but short-lived narrowly specialized eukaryotic microbiota, *e.g.* coccolithophorids, foraminifera and calcareous algae. We expect, therefore, that *Decastronema*-type fossils will eventually be found in peritidal settings throughout the Phanerozoic and perhaps earlier.

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