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Heterologous expression of a plant uracil transporter in yeast: improvement of plasma membrane targeting in mutants of the Rsp5p ubiquitin protein ligase.

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3 1 **Heterologous expression of a plant uracil transporter in yeast: improvement of**
4
5 2 **plasma membrane targeting in mutants of the Rsp5p ubiquitin protein ligase**

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27 10 **Running title:** Functional expression of a plant transporter in yeast

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18 **Abstract:**

19 Plasma membrane proteins involved in transport processes play a crucial role in cell
20 physiology. On account of these properties, these molecules are ideal targets for
21 development of new therapeutic and agronomic agents. However, these proteins are
22 of low abundance, which limits their study. Although yeast seems ideal for expressing
23 heterologous transporters, plasma membrane proteins are often retained in
24 intracellular compartments. We tried to find yeast mutants potentially able to improve
25 functional expression of a whole set of heterologous transporters. We focused on
26 *Arabidopsis thaliana* ureide transporter 1 (AtUPS1), previously cloned by functional
27 complementation in yeast. Tagged versions of AtUPS1 remain mostly trapped in the
28 endoplasmic reticulum and were able to reach slowly the plasma membrane. In
29 contrast, untagged AtUPS1 is rapidly delivered to plasma membrane, where it
30 remains in stable form. Tagged and untagged versions of AtUPS1 were expressed in
31 cells deficient in the ubiquitin ligase Rsp5p, involved in various stages of the
32 intracellular trafficking of membrane-bound proteins. *rsp5* mutants displayed further
33 plasma membrane stabilization of untagged AtUPS1, and improved steady state
34 amounts of tagged versions of AtUPS1. *rsp5* cells are thus powerful tools to solve the
35 many problems inherent in heterologous expression of membrane proteins in yeast,
36 including ER retention.

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3 38 **Introduction:**
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8 40 Systematic sequencing of the genomes of complex organisms (e.g., the plant
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10 41 *Arabidopsis thaliana*, certain parasites, and man) has revealed a great number of
11
12 42 genes that are likely to code for membrane transporters [1]. Many of these proteins
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14 43 are assigned to this functional category on the sole basis of sequence similarity to
15
16 44 known transport proteins. Their biochemical properties are thus unknown and
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18 45 information regarding substrate specificity is often lacking. To possess such
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20 46 knowledge would be important both scientifically and industrially, because transport
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22 47 proteins (channels, pumps, and carriers) play a crucial role in cell physiology. This is
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24 48 illustrated by the numerous genetic diseases that are caused by defective transport
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26 49 systems. Furthermore, cell surface membrane transporters may be ideal targets for
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28 50 the development of new therapeutic or agronomic agents. Biological systems are
29
30 51 thus required for the functional expression of each transport protein and for the study
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32 52 of its biochemical properties by means of quick, simple tests. The yeast
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34 53 *Saccharomyces cerevisiae* expression system best meets these requirements. It is
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36 54 easy to use and inexpensive, its "transportome" has been extensively analyzed *in*
37
38 55 *silico* [2,3] and the function of over 150 yeast transport proteins has been identified
39
40 56 [4]. Moreover, the construction of a complete collection of strains deleted for each of
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42 57 the 6200 yeast genes [5] led to an impressive number of *S. cerevisiae* mutants
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44 58 deficient in the transport of compounds as diverse as inorganic ions, metabolites, and
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46 59 drugs. The phenotypes of these mutants are easily identified after growth on solid
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48 60 media and have been used extensively in complementation tests, notably to clone
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50 61 and characterize heterologous transporters.
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4 62 Hundreds of plant transporters have been successfully cloned by
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6 63 complementation in yeast [6-9]. The yeast system has also been used in biochemical
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8 64 approaches, for example, to analyze the functional properties of plant H⁺-ATPases
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10 65 [10]. Nevertheless, although yeast seems ideal for expressing plasma membrane
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12 66 proteins including transporters and receptors, investigators often found some of
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14 67 these proteins to be inactive in yeast, sometimes because the protein did not fold
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16 68 properly and/or was not delivered to the plasma membrane but accumulated in a
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18 69 secretory compartment (endoplasmic reticulum (ER), Golgi apparatus, secretory
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20 70 vesicles), sometimes even leading to formation of karmellae [11]. On the other hand,
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22 71 some heterologous proteins correctly delivered to the plasma membrane have been
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24 72 described to undergo rapid endocytosis and turnover, resulting in low plasma
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26 73 membrane levels [12].

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32 74 A few attempts to improve functional expression in yeast of heterologous
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34 75 plasma membrane transporters have been reported, but they often focused on only
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36 76 one given transporter [13,14]. Attempts to identify yeast mutants that would
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38 77 potentially improve the functional expression of a whole set of plasma membrane
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40 78 proteins and more specifically transporters, are still lacking. Finding such mutants for
41
42 79 improvement of the functional expression of several plant and mammalian
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44 80 transporters was the objective of an European network, EFFEXPORT ("Engineering
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46 81 yeast for efficient expression of heterologous membrane transporters"). We report
47
48 82 here data we obtained within EFFEXPORT in the case of the *Arabidopsis Thaliana*
49
50 83 ureide transporter (AtUPS1), already known to be functionally expressed in *S.*
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52 84 *cerevisiae* [15]. AtUPS1, was identified by functional complementation of a yeast *dal4*
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54 85 *dal5* mutant [15] defective in uptake of allantoin. AtUPS1 belongs to a superfamily of
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56 86 plant transporters with five members in Arabidopsis. Analysis of hydrophobicity

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3 87 predicts 10 putative transmembrane domains, with N- and C-termini predicted to
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5 88 protrude into the extracellular space. UPS proteins display a conserved central
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7
8 89 domain between predicted transmembrane helices 5 and 6, which contains a
9
10 90 consensus sequence for a P loop, also designated as a « Walker A » motif for ATP
11
12 91 binding [15]. Expression in yeast and *Xenopus* oocytes allowed to demonstrate that
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14
15 92 *AtUPS1* mediates uptake of allantoin and related metabolites including uracil [15,16].
16
17 93 The fate of *AtUPS1* can thus be compared to that of the endogenous uracil
18
19 94 permease (*Fur4p*), a well-known yeast transporter, the trafficking of which has been
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21
22 95 studied extensively (reviewed in [17]). *Fur4p*, which belongs to a family of five
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24 96 homologous proteins in *S. cerevisiae* [2], also consists of ten transmembrane spans,
25
26 97 with cytoplasmic oriented N- and C- termini [18]. Like most yeast plasma membrane
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28 98 proteins, it displays plasma membrane ubiquitylation, catalyzed by the *Rsp5p*
29
30 99 ubiquitin protein ligase, a modification triggering its internalization and subsequent
31
32 100 vacuolar degradation [19]. The same ubiquitin ligase, the sole member of the family
33
34 101 of *Nedd4* ubiquitin ligases in yeast [20], was also demonstrated to be required for
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36 102 direct Golgi-to-vacuole trafficking of a number of plasma membrane transporters,
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38 103 including *Fur4p* [17,21], *i.e.* proteins routed to a direct degradation pathway
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40 104 bypassing the plasma membrane under certain nutrient/substrate conditions. *Rsp5p*
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42 105 was also described to be required for Golgi-to-vacuole traffic of misfolded plasma
43
44 106 membrane proteins misrouted to the vacuole [22]. In latter two cases, these proteins
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46 107 were retargeted to plasma membrane in *rsp5* mutants. *rsp5* mutants thus display
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48 108 increased steady state amounts of many plasma membrane transporters (reviewed
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50 109 in [17]), and were good candidates for potential improvement of functional expression
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52 110 of heterologous transporters. However, in addition to its role in trafficking of
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54 111 membrane proteins [17], *Rsp5p* has many other functions, including essential
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3 112 functions [23,24]. Hence, only defined *rsp5* mutants may be used for optimization of
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5 113 functional expression of heterologous proteins. Two viable *rsp5* mutants that were
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8 114 described to be affected for several of the trafficking functions of Rsp5p appeared
9
10 115 interesting: *npi1* mutant, with altered *RSP5* promoter, leading to a 10-fold reduction in
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12 116 the steady state amount of this protein [25], and *rsp5ΔC2*, lacking Rsp5p C2 domain
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14 117 involved in localization of the enzyme at plasma membrane and endosomes [26].
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19 118 We report here an analysis of the fate of AtUPS1, as compared to that of
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21 119 Fur4p, in wild type and *rsp5* mutant cells, and show that *npi1* and *rsp5ΔC2* cells
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23 120 improved functional expression of the plant transporter.
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28 29 122 **Results**

30 123

31 124 **AtUPS1 mediates high affinity uracil transport in yeast**

32 125 As outlined above, when expressed in yeast, AtUPS1 transports [¹⁴C] labelled
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34 126 allantoin with high affinity and potentially other heterocyclic compounds as suggested
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36 127 by competition studies [15]. Moreover, AtUPS1 mediates uracil uptake when
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38 128 expressed in *Xenopus* oocytes [16]. In order to monitor the intracellular fate of
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40 129 AtUPS1 in yeast, *UPS1* was cloned in a multicopy plasmid under the control of the
41
42 130 galactose-inducible GAL promoter. We defined the characteristics of uracil uptake
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44 131 mediated by GAL-*UPS1*, as compared to uracil uptake mediated by the endogenous
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46 132 Fur4p cloned under the control of the same promoter. *fur4Δ* cells grown on galactose
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48 133 expressing either GAL-*UPS1* or GAL-*FUR4* displayed high sensitivity to 5-fluorouracil
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50 134 (5FU), a toxic analog of uracil: cells expressing the transporters were unable to grow
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52 135 on plates containing 1μM 5FU, whereas cells transformed with a control plasmid
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3 136 grew normally (Fig 1A). To determine the uracil transport properties of AtUPS1
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6 137 quantitatively, radiotracer uptake studies were performed using [¹⁴C] uracil. [¹⁴C]
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8 138 uracil uptake mediated by GAL-*UPS1* expressed in *fur4Δ* cells growing exponentially
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11 139 and fully induced on galactose was linear for at least 3 min, concentration-
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13 140 dependent, and displayed saturation kinetics with an apparent K_m of 6 μM , close to
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16 141 that observed in parallel for cells expressing GAL-*FUR4* (7.5 μM). *fur4Δ* cells
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18 142 expressing GAL-*UPS1* grown overnight in galactose containing media displayed an
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20 143 activity 50% of cells expressing GAL-*FUR4*, *i.e.*, more than 30-fold that of
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22 144 chromosomal encoded *FUR4* (Fig. 1B), thus providing a sensitive assay to follow
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25 145 AtUPS1 intracellular fate.
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30 147 **Insight into AtUPS1 intracellular trafficking in yeast**

32 148 Inducibility of AtUPS1 synthesis after galactose induction provides a useful
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34 149 tool for monitoring plasma membrane delivery, as previously demonstrated for the
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36 150 yeast Fur4p [27]. We measured uracil uptake activity after galactose induction of
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38 151 GAL-*UPS1* and GAL-*FUR4* in *fur4Δ* cells in parallel (Fig. 2A). In both cases, activity,
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40 152 as a measure of plasma membrane targeting, was detectable after 30 min induction.
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42 153 The increase in uracil uptake activity was linear for one hour with a similar slope for
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44 154 cells expressing the yeast and the plant transporters. Hence the heterologous
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46 155 AtUPS1 appears to be recognized efficiently by the yeast secretory machinery. We
47
48 156 then compared the fate of AtUPS1 and Fur4p along the endocytic pathway using an
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50 157 experimental condition known to trigger rapid Fur4p internalization and subsequent
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52 158 vacuolar degradation, *i. e.* the inhibition of protein synthesis by addition of
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54 159 cycloheximide [28] (Fig. 2B). Cycloheximide was added to exponentially growing
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56 160 *fur4Δ* cells induced overnight in galactose for expression of either AtUPS1 or Fur4p.
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3 161 Uracil uptake activity of cells expressing Fur4p decreased rapidly ($t_{1/2} = 45$ min),
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5 162 whereas uracil uptake activity of cells expressing AtUPS1 did not display a decrease
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8 163 for four hours. Hence, once delivered to the plasma membrane, the plant AtUPS1
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10 164 was remarkably stable, and did not undergo obvious endocytosis after inhibition of
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12 165 protein synthesis.
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17 167 **C-terminally tagged versions of AtUPS1 are retained in the ER**

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20 168 The monitoring of uracil uptake activity can afford insight into the intracellular
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22 169 fate of AtUPS1, since it provides information about the plasma membrane located
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24 170 transporter. This information, however, remains limited, and does not indicate the
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26 171 fraction of plasma membrane-delivered protein versus potential intracellular pools. In
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28 172 the absence of available antibodies, we decided to monitor the fate of tagged
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30 173 versions of AtUPS1. C-terminally tagged versions of AtUPS1 were constructed, first
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32 174 with a GFP-tag, a powerful tool for monitoring the intracellular fate in yeast of plasma
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34 175 membrane proteins of heterologous [14] or endogenous origin, including that of
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36 176 Fur4p [29-31]. *fur4Δ* cells transformed with a multicopy plasmid carrying *GAL-UPS1-*
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38 177 *GFP* displayed high 5FU sensitivity after growth on galactose (Fig. 3A). Strikingly,
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40 178 *fur4Δ* cells transformed with multicopy plasmid carrying *GAL-UPS1* tagged with the
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42 179 smaller HA epitope displayed intermediate 5FU sensitivity after growth on galactose,
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44 180 indicating that a smaller tag did not improve AtUPS1 functionality. According to these
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46 181 plate assays, C-terminally tagged AtUPS1 thus appeared functional, notably GFP-
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48 182 tagged AtUPS1. However, *fur4Δ* cells transformed by the multicopy plasmid carrying
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50 183 *GAL-UPS1-GFP* and fully induced displayed only a very low level of uracil uptake
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52 184 activity (0.04 nMol/min/A₆₀₀), an activity 50 fold lower than that observed in the case
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54 185 of induced cells transformed by a plasmid carrying untagged AtUPS1 (not shown).
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3 186 The use of a smaller tag (HA) or another promoter (CYC1) did not improve uracil
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5 187 uptake activity.
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8 188 The GFP -or HA- tags may inhibit transport activity or impair plasma
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10 189 membrane delivery of tagged transporter due to folding problems. We checked GFP
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12 190 fluorescence in time course experiments using the multicopy plasmid carrying the
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14 191 *GAL-UPS1-GFP* (Fig. 3B), or the multicopy plasmid *GAL-Fur4-GFP* as a control.
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16 192 Galactose induction of *Fur4-GFP* led, after 30 min, to observation of small internal
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18 193 compartments, likely Golgi/secretory vesicles. 30 min later, plasma membrane
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20 194 staining was clearly evidenced, often in a polarized fashion, with intense staining of
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22 195 small buds (Fig. 3B). The distribution of *AtUPS1-GFP* was strikingly different. After
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24 196 30 min of galactose induction of *AtUPS1-GFP* expression, a perinuclear staining was
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26 197 clearly observable. 30 min later, or after overnight induction, the perinuclear staining
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28 198 was still present and a discontinuous staining at/or underneath the plasma
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30 199 membrane has appeared. This pattern is typical of the yeast ER. Intense staining of
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32 200 lines or spots, as if intracellular membranes had formed aggregates were also
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34 201 visualized (Fig. 3B). Indeed, electron microscopy of the ultrastructural morphology of
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36 202 cells fully induced for expression of *AtUPS1-GFP* revealed proliferation, hanks of ER
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38 203 membranes (Fig. 3D), as often described in the case of cells overexpressing ER-
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40 204 retained proteins [11]. This altered morphology did not resulted from the
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42 205 overexpression of a membrane protein as such: overnight overexpression of
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44 206 endogenous *FUR4* from the same multicopy, GAL-inducible plasmid, did not lead to
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46 207 altered morphology (not shown).
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55 208 ER-retention of GFP-tagged *AtUPS1* apparently did not result from the mere
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57 209 overexpression of this protein. Perinuclear/ER staining was also evidenced upon
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59 210 expression of *AtUPS1-GFP* from a centromeric plasmid under the control of the mild
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3 211 strength *CYC1* promoter, leading to a steady state protein level about 4-fold lower
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5 212 than that observed after two hours galactose induction of GAL-*UPS1-GFP* (Fig. 3B
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7 213 and C). Furthermore, when analyzed by sucrose gradient fractionation, AtUPS1-GFP
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9 214 expressed from the CEN *CYC1*-plasmid or from the 2 μ GAL-inducible plasmid
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11 215 displayed exactly the same pattern with major pool in internal fractions (data not
12
13 216 shown).

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17 217 Hence, the GFP tag at the C-terminus of AtUPS1 obviously triggers ER
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19 218 retention of the transporter. The low uracil uptake activity of cells expressing this
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21 219 transporter can only be observed after long expression periods (3-4 hours, cf Fig.
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23 220 6B), and likely corresponds to the low fraction of protein finally reaching the plasma
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25 221 membrane. With such folding problems, overexpression is a way to increase steady
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27 222 state plasma membrane expression: both 5FU sensitivity and uracil uptake activity
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29 223 were improved in the case of AtUPS1-GFP expression from a multicopy plasmid and
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31 224 strong GAL promoter compared to expression from a centromeric plasmid under the
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33 225 control of a lower strength promoter.
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41 227 **Intracellular fate of AtUPS1 carrying an internal myc tag in yeast: slow ER exit** 42 43 228 **but final plasma membrane delivery**

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46 229 We checked whether insertion of a small myc-tag inside the central loop
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48 230 containing the Walker A motif would be a better way to study AtUPS1 intracellular
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50 231 trafficking. Cells expressing internally myc tagged AtUPS1 from a multicopy
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52 232 galactose inducible plasmid displayed high sensitivity to 5FU (Fig. 4A). Cells
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54 233 transformed with the multicopy GAL-*UPS1^{myc}* plasmid grown overnight in galactose
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56 234 displayed a relatively high level of uracil uptake activity: 1 nMol/min/A₆₀₀, *i. e.* 70%
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58 235 that observed in the case of untagged AtUPS1 (Fig. 4B), suggesting that the myc tag
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3 236 at this position had only a small impact on AtUPS1 function. However, this tag had a
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5 237 clear influence on AtUPS1 intracellular trafficking. Induction experiments showed that
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8 238 uracil uptake activity appeared slowly: 4-6 hours were necessary before we were
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10 239 able to measure any detectable uptake activity (Fig. 4C). Aliquots withdrawn at
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12 240 several time points after galactose induction were analyzed by protein gel blots using
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14
15 241 a specific anti-myc antibody. AtUPS1^{myc} appeared on gels as a band of apparent
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18 242 molecular mass of about 36 kDa, *i. e.* slightly below the expected molecular mass
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20 243 deduced from predicted protein sequence (44 kDa) (Fig. 4E), as often observed for
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23 244 very hydrophobic proteins. A comparable pattern was observed after overnight
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25 245 galactose induction of AtUPS1^{myc} expressed from either a centromeric or a multicopy
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28 246 plasmid (data not shown).

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30 247 In order to check the intracellular location of AtUPS1^{myc} derived species, cells
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32 248 transformed with GAL-UPS1^{myc} were induced by addition of galactose, aliquots
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35 249 withdrawn periodically were fixed and analyzed by immunofluorescence using a
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37 250 monoclonal anti-myc antibody. One hour induction was sufficient to observe a
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39 251 specific signal, mostly perinuclear, supported by simultaneous DAPI staining (Fig.
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42 252 5A). To obtain better resolution of the cell surface staining (potentially corresponding
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44 253 to either ER or plasma membrane), cells were examined by confocal microscopy.
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47 254 The lower background in optical slices made it possible to show that all cells
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49 255 displayed perinuclear staining, together with discontinuous regions of cell surface
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51 256 staining (Fig. 5B). Because cell surface was not homogeneously stained, we can
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53 257 conclude that the main signal corresponds to ER staining and that a plasma
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55 258 membrane localisation cannot be detected by this approach. The low rate of uracil
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57 259 uptake activity thus likely resulted from low ER exit rates (Fig. 4C) The finding that
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60 260 the activity of fully induced cells is in a similar range as that of cells expressing

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3 261 untagged AtUPS1, which was rapidly targeted to plasma membrane, may suggest
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5 262 that after slow rates of ER exit, AtUPS1^{myc} finally reached the plasma membrane.
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7 263 Further indication that AtUPS1^{myc} stored in internal compartments could finally reach
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9 264 plasma membrane was provided by the observation of some increase in uracil
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11 265 uptake activity after stopping transporter synthesis by the addition of CHX (Fig. 4D).
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13 266 Once targeted at the plasma membrane, AtUPS1^{myc} was rather stable, as judged
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15 267 from the extreme stability of uracil uptake activity for over 3 hours after this transient
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17 268 uracil uptake activity increase following CHX addition. However, some low rate
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19 269 endocytosis likely occurred as compared to the incredibly stable untagged AtUPS1
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21 270 (Fig. 4D).
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32 273 **Mutations in the Rsp5 ubiquitin protein ligase improve functional expression of** 33 34 274 **tagged and untagged AtUPS1 in yeast**

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36 275 Despite the difficulties encountered in tagging AtUPS1 at different positions,
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38 276 the high sensitivity of the two functional tests, 5FU sensitivity and uracil uptake
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40 277 measurements, provided suitable tools to check whether yeast mutants could
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42 278 improve the steady state levels of functional tagged or untagged AtUPS1 versions.
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44 279 We used two viable *rsp5* mutants, *npi1* and *rsp5ΔC2*, that display delayed
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46 280 endocytosis of several cargoes, including Fur4p. Accordingly, they displayed
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48 281 increased 5FU sensitivity as a result of the plasma membrane stabilization of
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50 282 endogenous chromosomal or plasmid encoded Fur4p (not shown). We checked the
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52 283 fate of tagged and untagged versions of AtUPS1 in *npi1* and *rsp5ΔC2* mutant cells.
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58 284 For this purpose, *FUR4* was deleted in wild type, *npi1* and *rsp5ΔC2* cells, and
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60 285 the resulting strains were transformed with multicopy plasmids carrying either *UPS1*

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3 286 under the control of the constitutive PGK promoter (plasmid pFL61-*UPS1*), or
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5 287 galactose inducible *UPS1*, *UPS1-GFP* and *UPS1^{myc}*. Transformed cells grown in
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8 288 glucose, or galactose containing media in the case of plasmids with a GAL promoter
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10 289 displayed identical growth in the absence of 5FU (Fig. 6A), indicating that neither the
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12 290 mutations, nor the expression of the plant transporters impaired growth. 5FU
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14 291 sensitivity of transformants was strikingly enhanced in both *rsp5ΔC2* and *npi1* cells
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16 292 expressing the tagged and untagged plant transporter when compared to wild type
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18 293 cells, with slightly stronger effect promoted in all cases by the *npi1* mutation
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20 294 (compare the size of isolated colonies) (Fig. 6A). This suggests higher amounts of
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22 295 plasma membrane tagged or untagged transporters in the mutants. In the case of the
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24 296 untagged transporters, the enhancement in 5FU sensitivity was evidenced after
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26 297 synthesis from PGK or *GAL10* promoter at different 5FU concentrations (Fig. 6A),
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28 298 indicating that *rsp5* mutations interfered with trafficking, rather than with galactose-
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30 299 driven expression. For untagged AtUPS1 which is rapidly delivered to the plasma
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32 300 membrane and very stable in wild type cells, the increase in functional transporter
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34 301 activity observed in *rsp5* α mutants may result from inhibition of some direct Golgi to
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36 302 vacuole targeting, or from protection against a possible low basal endocytosis, for
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38 303 instance when cells reached stationary phase, a likely situation for cells grown
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40 304 several days on plates.
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48 305 In the case of the myc-tagged version of AtUPS1, we monitored the fate of the
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50 306 transporter after galactose induction by western blots (Fig 6C), and uracil uptake
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52 307 measurements (Fig. 6B). In agreement with fluorescent data, AtUPS1^{myc} protein was
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54 308 already detectable in wild type cells at early time points after induction (30-60 min)
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56 309 but uracil uptake was readily measurable only at far later time points (4 hours). In
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58 310 *npi1* cells, AtUPS1^{myc} was detectable in higher amounts at all time points and uracil
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3 311 uptake activity appeared at least one hour earlier compared to wild type cells (Fig.
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5 312 6B). *rsp5ΔC2* mutation also resulted in more rapid appearance of uracil uptake
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7 313 activity. Strikingly, even if the activity was lower for cells expressing AtUPS1-GFP
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9 314 compared to AtUPS1^{myc}, the rate of appearance of uptake was very similar. For both
10
11 315 types of tagged transporters uracil uptake activity was thus 2-3 fold higher in mutant
12
13 316 cells than in wild type cells after 4 hour induction. Since the major pool of both
14
15 317 AtUPS1-GFP and AtUPS1^{myc} seemed to be in the ER, both *npi1* and *rsp5ΔC2*
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17 318 mutations apparently lead to accelerated ER exit.
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320 Discussion:

321 The use of a regulable promoter and the high sensitivity of uracil uptake
322 measurements enabled us to obtain crucial information about the fate of untagged
323 AtUPS1 uracil transporter in yeast. AtUPS1 is rapidly delivered to the plasma
324 membrane, with kinetics indistinguishable from those of endogenous yeast
325 transporter. Hence, the yeast secretory pathway efficiently handles the heterologous
326 plant transporter. It is possible that such property was the reason for the successful
327 cloning by functional expression of so many plant transporters when using yeast as a
328 host system [6,7]. Plasma membrane delivered AtUPS1 exhibited striking stability,
329 notably when compared to the endogenous yeast uracil permease especially
330 susceptible to stress-induced endocytosis [28]. This was true for both untagged
331 transporter and for internally tagged AtUPS1^{myc} (not shown). This differential stability
332 between yeast and plant uracil transporters may possibly reflect fundamental
333 differences in their endocytic processes. Endocytosis in yeast is dependent on prior
334 ubiquitylation of plasma membrane cargoes by the Rsp5p ubiquitin protein ligase
335 [17]. A subset of mammalian proteins undergo ubiquitin-dependent endocytosis,

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3 336 sometimes involving ubiquitin-protein ligases of the Nedd4/Rsp5 family [32]. In
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6 337 contrast, although plants display numerous ubiquitin protein ligases, they do not
7
8 338 seem to have Rsp5p orthologs [33]. The extreme stability of AtUPS1 in yeast could
9
10 339 result from the absence or low accessibility of ubiquitylation sites that Rsp5p can
11
12 340 recognize, at least under the experimental conditions we tested. One possible reason
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15 341 could be the luminal orientation of both AtUPS1 N- and C-termini.
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17 342 In addition to obtaining information on intracellular trafficking of the plant
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19 343 AtUPS1 in yeast based on uptake measurements, we tried to gain new insights into
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21 344 the biochemical properties of this transporter. Unfortunately, tagging the transporter,
22
23 345 either at the N-terminus (not shown) and C-terminus or inside its central loop, lead to
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25 346 folding problems, often resulting in ER retention of most of the protein, preventing us
26
27 347 from using these tagged versions to obtain a judicious biochemical characterization
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29 348 of this transporter. A marked difference, however, was observed between internal
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31 349 tagging, and tagging at the N- and C-termini, which was far more deleterious.
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33 350 Internally tagged transporter displayed delayed ER exit, but finally reached the
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35 351 plasma membrane in a fully functional state, and fully induced cells expressing this
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37 352 version of the transporter displayed activity closely resembling that of cells
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39 353 expressing untagged transporter. In contrast, cells expressing C-terminally tagged
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41 354 transporter displayed 40-fold less uracil uptake activity compared to cells expressing
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43 355 untagged transporters, probably because of a very low percentage of the protein
44
45 356 exiting the ER. In latter case, some improvement could be achieved by increased
46
47 357 levels of expression (stronger promoter, multicopy versus centromeric plasmid). The
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49 358 deleterious effect of C-terminal tags could result from the unusual structure of the
50
51 359 transporter. Hydrophobicity plot analysis of proteins of the UPS family suggested an
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53 360 external orientation of both termini [15] a situation somewhat rare for transporters.
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3 361 Most yeast transporters, for instance, display cytoplasmic oriented termini, as
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5 362 predicted from hydrophobicity plot analysis or from experimental data [2]. Many of
6
7 363 these transporters were studied after tagging, most often at their C-terminus, a
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9 364 modification that did not induce any major trafficking problems. The observation that
10
11 365 N- and C-terminal tags impaired AtUPS1 folding is compatible with an external
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13 366 orientation, but experimental data are required to prove this. One obvious lesson
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15 367 from these observations is that tagging at the positions and with the tags used in the
16
17 368 present study may probably influence the fate of UPS proteins in plant cells in a
18
19 369 similar way as in yeast cells. Indeed, previous studies attempting to study the
20
21 370 subcellular location of N- or C-terminal fusions of UPS proteins with GFP by transient
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23 371 expression in Arabidopsis protoplasts showed that these fusion proteins did not
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25 372 reach the plasma membrane (Schmidt, A., Baumann, N. and Desimone, M.,
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27 373 unpublished data). This suggests that yeast may also represent a useful
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29 374 experimental system to decipher where tags can be introduced in heterologous
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31 375 transporters.
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39 376 The distinctive behaviour of the various versions of AtUPS1, one correctly
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41 377 targeted to the plasma membrane, and two retained in the ER to various extents,
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43 378 provided the opportunity to check how to improve the steady state plasma membrane
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45 379 amount of heterologous transporters with distinct intracellular fates. We checked
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47 380 specifically the influence of mutations in Rsp5p. This ubiquitin protein ligase is
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49 381 involved in various stages of intracellular trafficking of membrane-bound proteins,
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51 382 including ER-associated degradation, plasma membrane internalization, Golgi to
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53 383 vacuolar trafficking, and sorting to multivesicular bodies [17]. Limiting or preventing
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55 384 latter three trafficking steps results in elevated steady state levels of yeast
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57 385 transporters. Two specific viable mutants were used, *npi1*, with decreased amounts
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3 386 of Rsp5p [25] and *rsp5ΔC2*, lacking the C2 domain of Rsp5p [34], dispensable for
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6 387 viability, but critical for Rsp5p trafficking functions [26,34-38]. Strikingly, both mutants
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8 388 improved steady state amounts of functional, plasma membrane targeted, tagged
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10 389 and untagged versions of AtUPS1, as judged from increased 5FU sensitivity. In the
11
12 390 case of untagged transporter, increased 5FU sensitivity was observed after
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15 391 expression of the transporter under the control of the constitutive PGK promoter, or
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17 392 the inducible *GAL10* promoter, indicating that the mutations interfered as expected
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19 393 with trafficking rather than with expression.
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21

22 394 In the case of the untagged version of AtUPS1, it was difficult to further
23
24 395 explore the mechanism of this improvement. Plasma membrane AtUPS1 was so
25
26 396 stable in wild type cells submitted to CHX treatment that further stabilisation in *rsp5*
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28 397 cells could not be detected in this type of experiment. The improvement in plasma
29
30 398 membrane steady state amounts of AtUPS1 in *npi1* and *rsp5ΔC2* cells might possibly
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32 399 arise from a low basal endocytosis of AtUPS1, undetectable in CHX chase
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34 400 experiments, but possibly occurring once cells reach stationary phase, as is the case
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36 401 for cells grown on plates. Alternatively, the increase in plasma membrane AtUPS1 in
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38 402 these *rsp5* mutants could result from a decreased direct Golgi to vacuole targeting of
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40 403 a fraction of the transporter.
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46 404 Mutations in *RSP5*, either *npi1* or *rsp5ΔC2*, had a similar impact on the
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48 405 intracellular fate of tagged versions of AtUPS1 at both the C-terminus or in the
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50 406 intracellular loop. These two variants of AtUPS1 were mainly located in the ER, with
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52 407 possibly exit in only very limited amounts (AtUPS1-GFP), or more important final
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54 408 amounts (AtUPS1^{myc}) thus leading to uracil uptake activity similar to that displayed by
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56 409 cells expressing untagged transporters. In both cases, the rate of plasma membrane
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58 410 delivery of uracil uptake activity was greatly delayed, with activity measurable after 3-
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3 411 4 hours induction instead of 30 min for AtUPS1 –i. e. long after detection of the
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6 412 protein on Western blots- and the two *rsp5* mutations reduced this delay. At first
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8 413 glance, this effect can be attributed to an effect of Rsp5p on ER exit. Among its many
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10 414 functions, Rsp5p was described to be involved in ubiquitylation followed by
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12 415 proteasome degradation of several misfolded soluble and membrane-bound proteins
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15 416 retained in the ER, notably under conditions of saturation of other ER-associated
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17 417 degradative (ERAD) pathways [39]. Tagged forms of AtUPS1 might be partly
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19 418 susceptible to such an Rsp5p-dependent ER-associated degradation, and inhibition
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21 419 of this degradation in *npi1* and *rsp5ΔC2* mutants would result in more rapid ER exit of
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23 420 transporters escaping this degradation to some extent. It is also possible that a small
24
25 421 number of misfolded tagged transporters undergo ER to Golgi trafficking, followed by
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27 422 Golgi to vacuole sorting. Rsp5p was also shown to be involved in Golgi to vacuole
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29 423 trafficking of some misfolded mutant plasma membrane proteins [22] or of yeast
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31 424 transporters displaying direct vacuolar targeting under specific nutrient conditions
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33 425 [17]. In both cases in *rsp5* mutants, these proteins are directed to plasma membrane
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35 426 [22,40]. A small fraction of tagged AtUPS1 could undergo such direct vacuolar
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37 427 trafficking, a process inhibited in *rsp5* mutants. In support of a partial direct Golgi to
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39 428 vacuole trafficking of AtUPS1^{myc}, we observed that *pep4* mutant cells, deficient for
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41 429 vacuolar protease activities, displayed higher amounts of the 36 kDa AtUPS1^{myc}
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43 430 species than wild type cells even after short induction times, as did *vps23Δ* mutants,
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45 431 impaired in Golgi to vacuole trafficking (data not shown). The increase in plasma
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47 432 membrane amounts of functional forms of tagged AtUPS1 in *rsp5* mutants, attested
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49 433 in exponentially growing cells by increased uracil uptake activity, and on plates by
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51 434 increased 5FU sensitivity might also result from an inhibition of Rsp5p-dependent
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3 435 processes at three levels: ERAD, Golgi-to-vacuole trafficking and plasma membrane
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5 436 internalization.
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8 437 Rsp5p plays a key role in trafficking of yeast plasma membrane proteins. The
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10 438 present study shows that several viable *rsp5* mutants display increased plasma
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12 439 membrane amounts of a plant transporter, rapidly targeted to the plasma membrane.
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14 440 Strikingly, within the same european network (EFFEXPORT), other laboratories
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16 441 observed, in viable *rsp5* mutants, improved functional expression of several
17
18 442 heterologous transporters, including several NH_4^+ transporters (Rh family) of animal
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20 443 origin (Marini A and André B, personal communication), and increased plasma
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22 444 membrane amounts versus internal fractions of mammalian Na^+/H^+ antiporters
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24 445 (Flegegova, H, Haguenaer-Tsapis, R and Sychrova, H. Biochem. Biophys. Acta, in
25
26 446 press). In addition, these mutants optimize plasma membrane delivery of tagged
27
28 447 versions of the plant AtUPS1, stacked in the ER as a result of folding problems. ER
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30 448 retention is one of the major problems encountered in the case of expression of
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32 449 heterologous plasma membrane proteins in yeast. The increase in steady state
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34 450 plasma membrane amounts of heterologous plant and mammalian transporters,
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36 451 upon expression in viable *rsp5* mutants, show that these cells could be powerful tools
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38 452 to solve the many problems inherent in heterologous expression of membrane
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40 453 proteins, including ER retention.
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51 52 455 **Materials and methods**

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55 56 457 *Yeast strains and growth conditions*

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58 458 Yeast strains were transformed as described by Gietz et al [41]. Cells were grown at
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60 459 30°C in minimal medium (YNB) containing 0.67% yeast nitrogen base without amino

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3 460 acids (BD bioscience, NJ, USA), and supplemented with appropriate nutrients. The
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5 461 carbon source was 2% glucose, or 2% galactose plus 0.05% glucose as indicated in
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7 462 the figure legends. Galactose induction was performed on cells grown overnight in
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9 463 2% raffinose plus 0.05% glucose up to an $A_{600nm}=0.5$. Galactose (2%) was then
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11 464 added to the medium.
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14 465 The disruption of *FUR4* gene was achieved by ORF replacement with long flanking
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16 466 homology regions to the KanMX4 cassette corresponding to the strategy described
17
18 467 by Wach [42].
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23 24 469 *Growth tests in the presence of 5-fluorouracil*

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26 470 Cells, prototroph for uracil, were cultured overnight in minimal medium containing
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28 471 glucose and spotted on plates containing minimal medium with galactose to induce
29
30 472 expression of *FUR4* or *UPS1* variants and supplemented with various concentrations
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32 473 of 5FU (Sigma-Aldrich, Lyon, France). The first drop contained $3 \cdot 10^4$ cells and each
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34 474 subsequent drop was diluted six-fold compared to the prior drop.
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40 476 *Construction of plasmids*

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42 477 DNA manipulations, including restriction analysis and ligations, were performed
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44 478 essentially as described by Maniatis *et al.* [43].
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47 479 The control plasmid p195gF-GFP (pRT208) expressing *FUR4*-GFP under the control
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49 480 of the *GAL10* promoter was constructed by cloning a *Pst*I/*Bam*H1 fragment (*FUR4*-
50
51 481 GFP) from pFL38gF-GFP [30] into p195gF [28] at the *Pst*I/*Bam*HI site.
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54 482 The pGAL-*UPS1* (pRT205) plasmid was constructed by insertion of a *Bam*HI/*Eco*RI
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56 483 fragment encoding *UPS1* (with pFL61-*UPS1* as a template [15]) in the *Bam*HI/*Eco*RI
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58 484 site of pYEF1 [44].
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3 485 To construct the plasmid GAL-*UPS1-GFP* (pRT206) we first built the pNBT29
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5 486 plasmid, which contains the yeast enhanced GFP under the control of the GAL10
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7 487 promoter on a multicopy plasmid. For this purpose, a BamHI/NotI fragment encoding
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9 488 GFP was obtained by PCR using the pUG35 plasmid as a template [45] and
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11 489 introduced at the BamHI/NotI site of pYEF1 [44]. Then a BamHI/ClaI fragment
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13 490 encoding AtUPS1 was amplified by PCR using pFL61-*AtUPS1* [15] as a template,
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15 491 and introduced at the BamHI/ClaI site of pNBT29 in frame with the coding sequence
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17 492 of GFP thereby creating a GFP C-terminally tagged version of AtUPS1. To construct
18
19 493 the *CYC1-UPS1-GFP* (pRT207) plasmid we cloned the BamHI/EcoRI fragment
20
21 494 containing *UPS1-GFP* at the BamHI/EcoRI site of the p416-CYC1 plasmid [46].
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23 495 The pGAL-*UPS1-HA* (pRT204) plasmid was obtained by insertion in the ClaI site of
24
25 496 the pYEF2 [44], in frame with the coding sequence of the HA tag, of a fragment
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27 497 encoding *UPS1* obtained by restriction of the plasmid GAL-*UPS1-GFP*.
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29 498 To construct the plasmid pGAL-*UPS1^{myc}* (pRT203) two fragments of *UPS1* were
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31 499 amplified separately by PCR using pFL61-*UPS1* as a template. One fragment
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33 500 contained at start an EcoRI and an XbaI site, the 5'- portion of the *UPS1* coding
34
35 501 sequence (from ATG to the position 549) and a BamHI site at the end. The other
36
37 502 fragment contained a BamHI site at the start, the coding sequence for the c-myc
38
39 503 epitope, the 3'- portion of *UPS1* (from position 550 to the stop codon), and an EcoRI
40
41 504 and a XhoI site at the end. Both fragments were sequentially cloned in pDR199 [47]
42
43 505 using the EcoRI/BamHI sites for the first fragment and BamHI/and XhoI for the
44
45 506 second. After sequencing, the complete c-myc tagged *UPS1* sequence was obtained
46
47 507 by restriction and subcloned into the Xba I / Xho I sites of the CEN plasmid p416-
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49 508 GAL [46]. Afterwards, the multicopy plasmid pGAL-*UPS1^{myc}* was obtained by
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3 509 subcloning a SacI/XhoI fragment of the p416-GAL-*UPS1^{myc}* into the SacI/XhoI sites
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5 510 of the p426-GAL plasmid [46].
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10 512 *Measurement of uracil uptake.*

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12 513 Uracil uptake was measured in exponentially growing cells as previously described.

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14 514 Yeast culture (1 ml) was incubated with 5 μM [^{14}C] uracil (ICN biomedical Illkirch,

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16 515 France) for 20 sec at 30°C, then quickly filtered through Whatman GF/C filters, which

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18 516 were in turn washed twice with ice-cold water and counted for radioactivity. In the

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20 517 case of low uracil uptake activity, this basic protocol was slightly modified, with the

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22 518 use of two ml samples and incubation for 2 min at 30°C.
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29 520 *Michaelis-Menten kinetics*

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31 521 Uracil uptake activities measured at various substrate concentrations were fitted to a

32
33 522 hyperbola with SIGMA PLOT 5.0, V5 according to Michaelis-Menten kinetics.
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39 524 *Yeast cell extracts, SDS-PAGE and Western immunoblotting*

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41 525 Total protein extracts were prepared by the NaOH/Trichloroacetic acid (TCA) lysis

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43 526 technique as described in [28]. Proteins were separated by SDS-PAGE on Tricine

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45 527 gels and transferred onto nitrocellulose membranes. The membranes were probed

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47 528 with monoclonal antibodies against GFP (Roche Diagnostics Meylan, France), or

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49 529 myc (9E10 from Roche Diagnostics), or polyclonal antibody against Gas1p (a kind

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51 530 gift from H. Riezman). Primary antibodies were detected using horseradish

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53 531 peroxidase-conjugated anti-rabbit or anti-mouse IgG secondary antibody (Sigma-

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55 532 Aldrich, Lyon, France) revealed by ECL chemiluminescence (Amersham).
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3 534 *Immunofluorescence*
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5 535 Immunofluorescence was performed as described in [48] except that cells were
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8 536 permeabilized with 0.5% Triton X100. The primary antibody was the monoclonal anti-
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10 537 Myc (9E10 from Roche Diagnostics, Meylan, France) and the secondary antibody
11
12 538 was an FITC-conjugated goat anti-mouse-IgG (Jackson ImmunoResearch
13
14 539 Laboratories, Inc., West Grove, PA). For DNA staining, 1 µg/ml Diamin-Phenylindol-
15
16 540 Dihydrochlorid (DAPI) was used. Samples were viewed under an Olympus
17
18 541 microscope BY61 using FITC and DAPI filter sets. Image acquisition was performed
19
20 542 using a Spot charge-coupled device camera SPOT4.05 .
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24 543 For confocal analysis, cells were imaged using an inverted microscope (Leica, Inc.
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26 544 Wetzlar, Germany) and scanning was performed with a True Confocal Scanner LEICA
27
28 545 TCS 4D.
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34 547 *Electron microscopy*
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36 548 Yeast cells were fixed by adding 200 µl of 50% aqueous glutaraldehyde to 10 ml of
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38 549 growth medium for 10 min and then centrifuged at 5000 g for 10 min at 4°C. After
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40 550 fixation with fresh fixatives for 2 h at 4°C, cells were washed in 0.1 M cacodylate
41
42 551 buffer (pH 7.4) and in water. Subsequently, cells were treated with 1% KMnO₄ for 2 h
43
44 552 on ice, washed in water and re-suspended in 2% aqueous uranyl acetate for 1 h at
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46 553 4°C. Cells were dehydrated in a graded series of ethanol, infiltrated in a mixture of
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48 554 ethanol and Spurr's resin and embedded in Spurr's low viscosity media. Thin
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50 555 sections were cut, stained with lead citrate and examined in a Tecnai 12 electron
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52 556 microscope (Eindhoven, Netherlands).
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TABLE 1. List of strains

| Strain | Background | Genotype | Source |
|--------|----------------|--|------------|
| MF04 | Σ 1278b | <i>MATa ura3 trp1 FUR4::KanMX4</i> | This study |
| MF05 | Σ 1278b | <i>MATa ura3 trp1 rsp5ΔC2 FUR4::kanMX4</i> | This study |
| MF06 | Σ 1278b | <i>Mata ura3 trp1 npi1 FUR4::kanMX4</i> | This study |
| 27061b | Σ 1278b | <i>Mata ura3 trp1</i> | [19] |
| 27064b | Σ 1278b | <i>Mata ura3 trp1 npi1</i> | [19] |
| BY4741 | BY | <i>Mata leu2Δ met15Δ ura3Δ his3Δ</i> | Euroscarf |

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TABLE 2. List of plasmids

| Plasmid | Characteristics | Source |
|--|--|------------|
| pPGK- <i>UPS1</i> (PFL61- <i>UPS1</i>) | 2 μ , <i>URA3</i> prom. <i>PGK-UPS1</i> | [15] |
| pGAL (PYeF2) | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>Cter HA</i> | [44] |
| pGAL- <i>FUR4</i> (pFL38gF) | CEN, <i>URA3</i> , prom. <i>GAL10</i> , <i>FUR4</i> | [30] |
| p195gF | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>FUR4</i> | [28] |
| p195gF-GFP (pRT208) | 2 μ , <i>URA3</i> , prom. <i>GAL10-FUR4-GFP</i> | This study |
| pGAL- <i>UPS1-HA</i> (pRT204) | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>UPS1-HA</i> | This study |
| pGAL- <i>UPS1</i> (pRT205) | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>UPS1</i> | This study |
| pGAL- <i>UPS1-GFP</i> (pRT206) | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>UPS1-GFP</i> | This study |
| pCYC1- <i>UPS1-GFP</i> (pRT207) | CEN, <i>URA3</i> , prom. <i>CYC1</i> , <i>UPS1-GFP</i> | This study |
| pGAL- <i>UPS1^{myc}</i> (pRT203) | 2 μ , <i>URA3</i> , prom. <i>GAL10</i> , <i>UPS1-myc</i> | This study |

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3 563 **Figure 1: Comparison of uracil uptake activity of *Arabidopsis thaliana* UPS1**
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5 564 **expressed in yeast and endogenous uracil permease Fur4p.**

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8 565 A: 5-Fluorouracil (5FU) sensitivity. *fur4*Δ cells transformed with either pGAL (empty
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10 566 vector), pGAL-*UPS1* or pGAL-*FUR4* were grown on galactose containing plates
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12 567 supplemented or not with 1 μM 5FU (toxic analog of uracil).

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15 568 B: Uracil uptake activity of AtUPS1 compared to Fur4p. *fur4*Δ strains transformed
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17 569 with pGAL (white), pGAL-*UPS1*(grey) or pGAL-*FUR4* (black) were grown to
18
19 570 exponential phase in galactose containing medium and used for measurement of
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21 571 [¹⁴C] uracil uptake as described in Materials and Methods. Results are the average of
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23 572 four measures (two measures in two independent experiments).
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29 574 **Figure 2: Intracellular trafficking of AtUPS1 in yeast.**

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32 575 A: *fur4*Δ strains transformed with pGAL-*UPS1* (triangle) or pGAL-*FUR4* (circle) were
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34 576 grown with raffinose as a carbon source. Galactose was then added to induce
35
36 577 expression of *AtUPS1* and *FUR4*. The kinetics of plasma membrane delivery of
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38 578 AtUPS1 and Fur4p was determined by quantification of [¹⁴C] uracil uptake every 30
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40 579 minutes after galactose induction. Results at each time point are the average of two
41
42 580 independent measurements.
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46 581 B: *fur4*Δ strains transformed with pGAL-*UPS1* (triangle) or pGAL-*FUR4* (circle) were
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48 582 grown in galactose containing medium. Protein synthesis was inhibited by addition of
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50 583 cycloheximide (CHX) (100 μg/ml). Uracil uptake activity was measured at various time
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52 584 points (two measurements) after CHX addition. Results are shown as the percentage
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54 585 of initial activities.
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3 587 **Figure 3: Expression of C-terminally GFP tagged AtUPS1 promotes ER**
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5 588 **proliferation in yeast**
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8 589 A: *fur4*Δ cells transformed with pGAL, pGAL-*UPS1-HA*, pGAL-*UPS1-GFP* and
9
10 590 *pCYC1-UPS1-GFP* were tested for growth on plates containing or not 5-fluorouracil
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12 591 (0,75 μM).
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15 592 B and C: *fur4*Δ cells transformed with pCYC-*UPS1-GFP*, pGAL-*UPS1-GFP* or pGAL-
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17 593 *Fur4-GFP* were grown to mid exponential phase either in glucose, or in raffinose
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19 594 containing medium in the case of strains bearing a plasmid with a GAL promoter.
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21 595 Galactose was then added to induce *AtUPS1-GFP* or *Fur4-GFP* expression. At the
22
23 596 indicated times, after galactose addition or after growth in glucose containing media
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25 597 (in the case of pCYC-*UPS1-GFP*), cells were observed by fluorescence microscopy
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27 598 (B) and protein extracts were prepared, resolved by SDS PAGE and analysed by
28
29 599 Western immunoblotting using an anti-GFP antibody (C)
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31

32 600 D: *fur4*Δ cells expressing pGAL and pGAL-*UPS1-GFP* grown overnight in galactose
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34 601 containing media were processed for electron microscopy. White arrows indicate the
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36 602 endoplasmic reticulum and N the nucleus.
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43 604 **Figure 4: Intracellular fate of myc tagged version of AtUPS1**
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46 605 A: *fur4*Δ cells transformed with pGAL, pGAL-*UPS1*, and pGAL-*UPS1^{myc}* were tested
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48 606 for 5-fluorouracil (1 μM) sensitivity on plates.
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51 607 B: [¹⁴C] uracil uptake activity of *fur4*Δ cells transformed with pGAL-*FUR4*, pGAL-
52
53 608 *UPS1* and pGAL-*UPS1^{myc}* grown to mid exponential phase on galactose containing
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55 609 medium. Results are the average of two independent measures.
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58 610 C: *fur4*Δ strains transformed with pGAL-*UPS1* (triangle) or pGAL-*UPS1^{myc}* (square)
59
60 611 were grown to mid log phase in raffinose containing medium and galactose was then

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3 612 added to induce expression of the transporter. The kinetics of plasma membrane
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5 613 delivery of AtUPS1 or Fur4p were determined by quantification of [¹⁴C] uracil uptake
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8 614 at various times after galactose induction.
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10 615 D: *fur4*Δ strains transformed with pGAL-UPS1 (triangle) or pGAL-UPS1^{myc} (square)
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12 616 were grown in galactose containing medium. Uracil uptake activity was measured at
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15 617 different time points after inhibition of protein synthesis by addition of CHX
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17 618 (100μg/ml). Results are shown as the percentage of initial activity.
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20 619 E: WT cells transformed with pGAL-UPS1^{myc} were grown to mid log phase in
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22 620 raffinose containing medium. Galactose was then added to induce transporter
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24 621 expression. Protein extracts were prepared at indicated times and proteins were
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26 622 resolved by SDS PAGE and analysed by Western immunoblotting using an anti-myc
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28 623 antibody to detect the transporter and an anti-Gas1p as a loading control.
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34 625 **Figure 5: Intracellular localization of UPS1^{myc} after galactose induction.**

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36 626 A: *fur4*Δ cells transformed with pGAL-UPS1^{myc} were grown overnight in raffinose
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38 627 containing media. Galactose was added during exponential growth phase. Aliquots
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40 628 were withdrawn at various time points, cells were fixed and processed for
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42 629 immunofluorescence. AtUPS1^{myc} was detected using an anti-myc antibody as
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44 630 described in Materials and Methods. The nuclei were stained using DAPI.
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46 631 Fluorescence was observed under an Olympus microscope.
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51 632 (Note that, as it is often the case for multicopy plasmid and galactose induction, only
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53 633 a subset of the cells within the population was stained with the anti-myc antibody)

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55 634 B: The same preparations as in (A) were also visualized using confocal microscopy.
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57 635 One section is presented for each time point.
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3 637 Figure 6: **Mutations in *RSP5* improve functional expression in yeast of tagged**
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5 638 **and untagged UPS1**

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8 639 A: *fur4* Δ , *rsp5* Δ *C2 fur4* Δ and *npi1 fur4* Δ cells transformed with pPGK-UPS1, pGAL-
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10 640 UPS1, pGAL-UPS1-GFP or pGAL-UPS1^{myc} were grown in glucose containing
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12 641 medium and spotted either on glucose (pPGK-UPS1) or on galactose containing
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14 642 plates (pGAL plasmids) with or without 5-fluorouracil as indicated. Different 5FU
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16 643 concentrations were used in order to differentiate optimally the growth of each of the
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18 644 tested strains with regard to the various plasmids: concentration was lower in the
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20 645 case of galactose-induced cells expressing untagged and myc-tagged AtUPS1 that
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22 646 display greater uracil uptake activities than cells expressing GFP-tagged AtUPS1, or
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24 647 untagged AtUPS1 under the control of the PGK promoter. In all cases, growth was
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26 648 tested using a whole range of 5FU concentrations, and a selection of representative
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28 649 plates is shown.

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34 650 B: *fur4* Δ (triangle), *rsp5* Δ *C2 fur4* Δ (square) and *npi1 fur4* Δ (diamond) transformed
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36 651 with pGAL-UPS1-GFP or pGAL-UPS1^{myc} were grown to mid log phase in raffinose
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38 652 containing media and the expression of the various transporters was induced by
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40 653 addition of galactose. Plasma membrane delivery of the transporters was assessed
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42 654 by measuring uracil uptake activity at different time points after galactose addition.

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45 655 C: Induction of AtUPS1^{myc} was monitored in *WT* and *npi1* cells by Western blot
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47 656 analysis of aliquots withdrawn at several time points after addition of galactose.
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49 657 Proteins were resolved by SDS polyacrylamide gel electrophoresis and analyzed by
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51 658 immunoblotting with anti-myc antibodies for quantification of transporter expression
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53 659 and anti-Gas1p as loading control.

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676 **References**

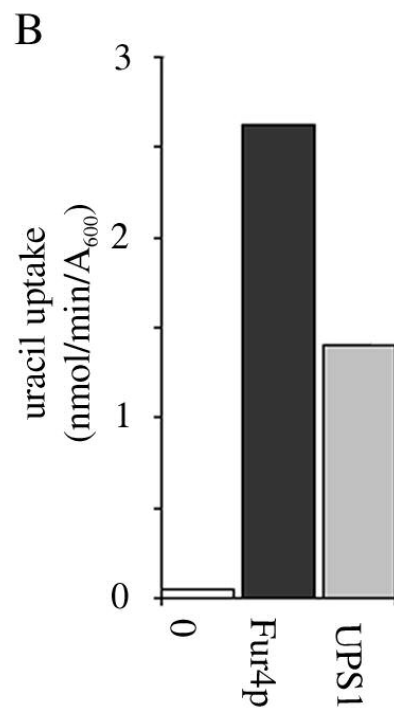
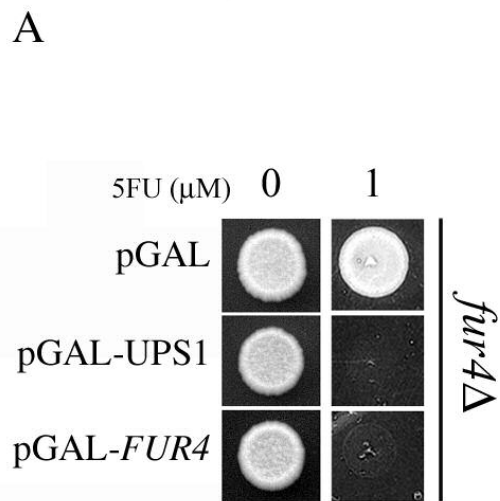
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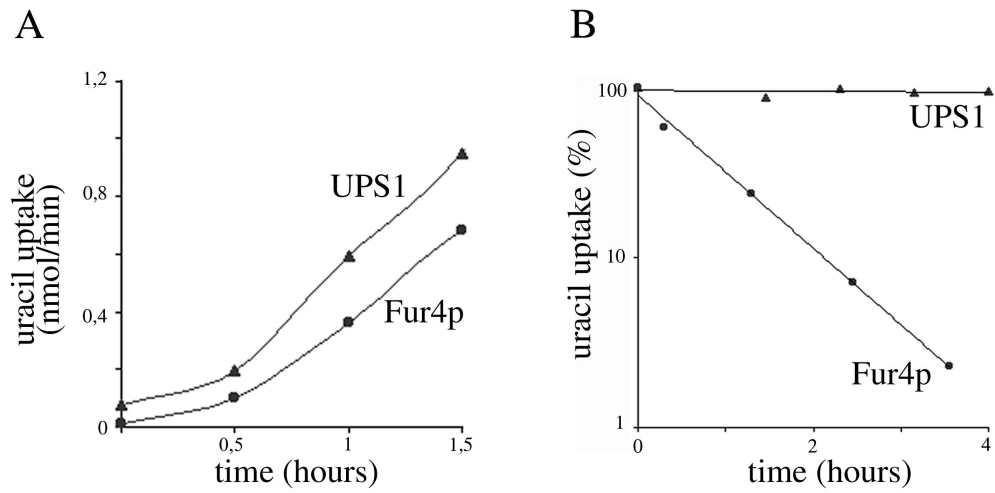
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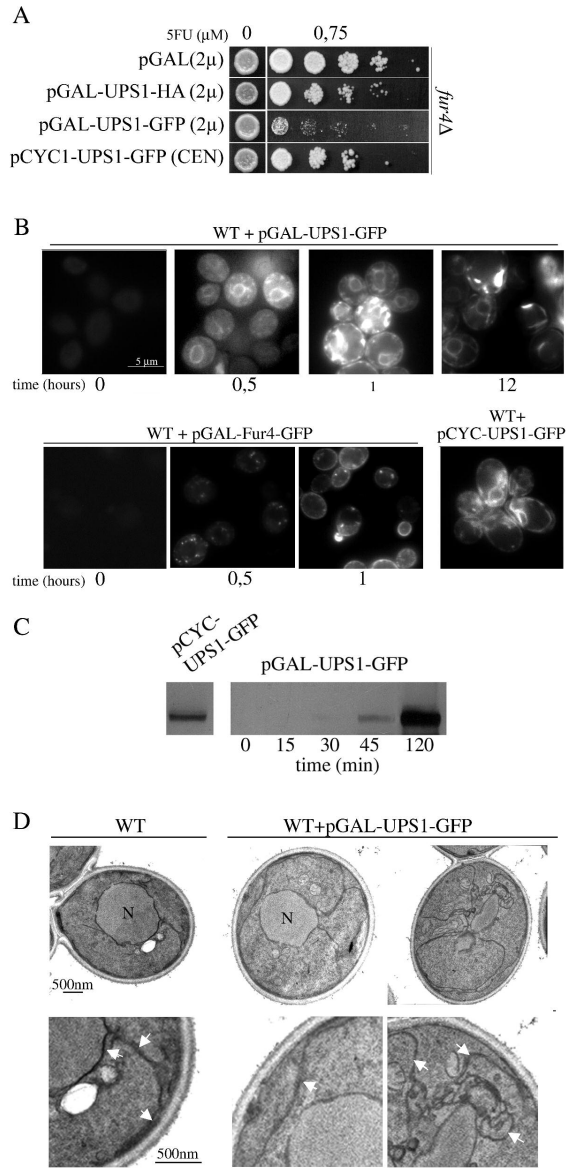
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Review

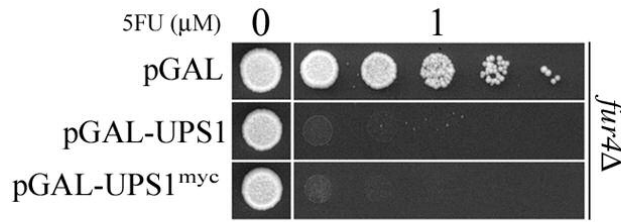


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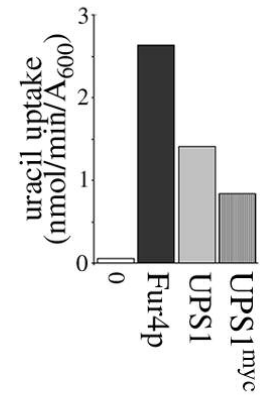


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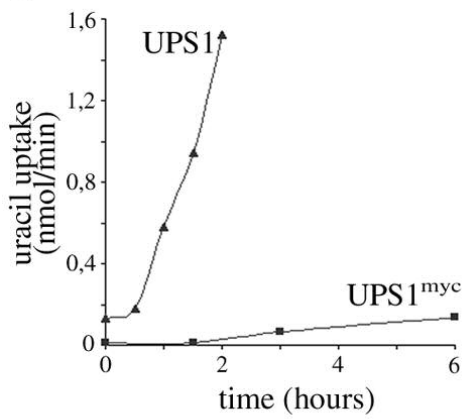
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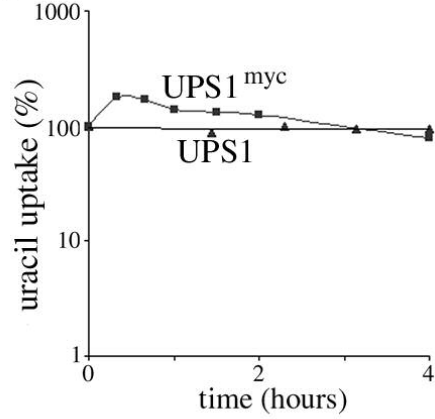
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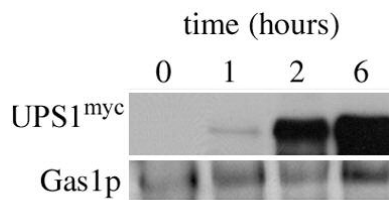
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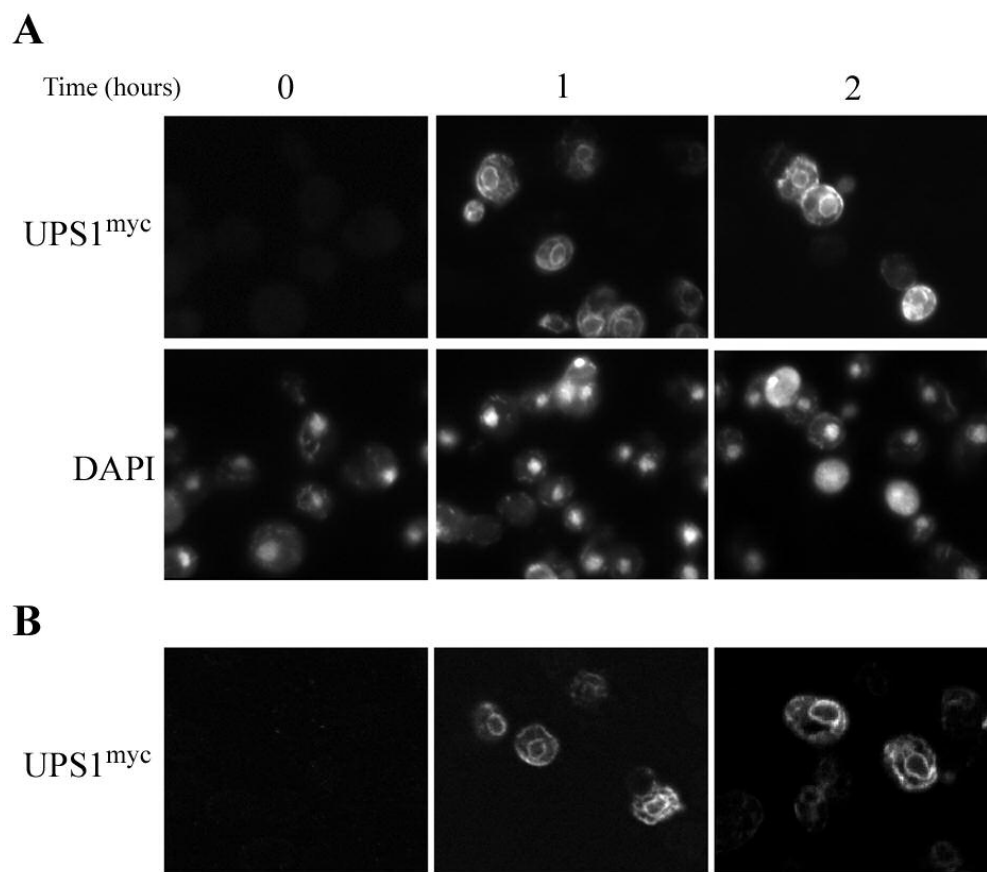


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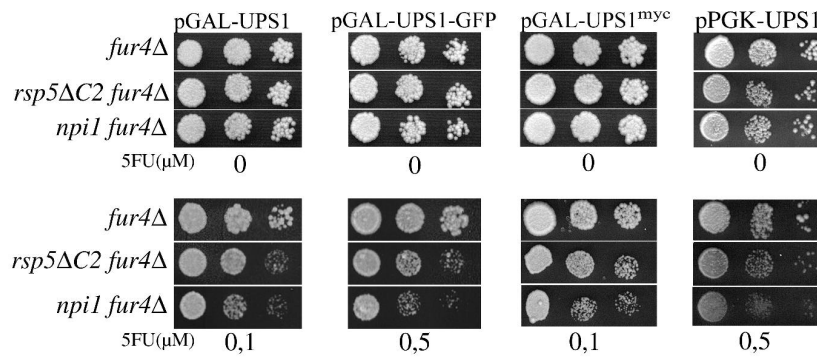
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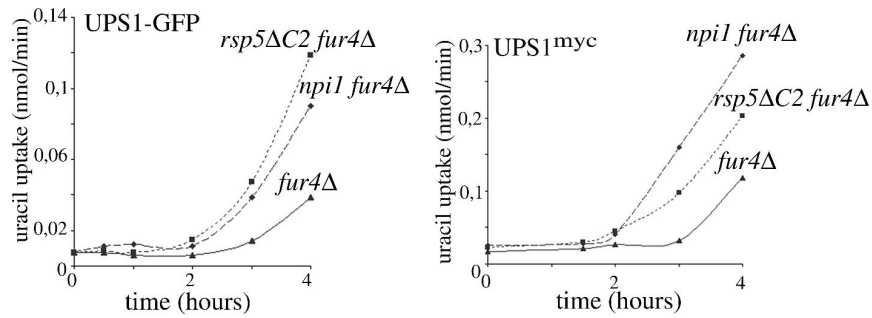


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