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Mimivirus Gene Promoters Exhibit an Unprecedented Conservation among all Eukaryotes

Karsten Suhre^{1*}, Stéphane Audic^{1*}, Jean-Michel Claverie^{1,2}

¹ Information Génomique & Structurale, CNRS UPR2589, IBSM, 13402 Marseille;

² University of Méditerranée, School of Medicine, 13385, Marseille, France

* These two authors contributed equally to this work.

Running head : Promoters are Uniquely Conserved in Mimivirus

SUMMARY

The initial analysis of the recently sequenced genome of *Acanthamoeba polyphaga* Mimivirus, the largest known double-stranded DNA virus, predicted a proteome of size and complexity more akin to small parasitic bacteria than to other nucleo-cytoplasmic large DNA viruses, and identified numerous functions never before described in a virus. It has been proposed that the Mimivirus lineage could have emerged before the individualization of cellular organisms in the 3 domains of life.

An exhaustive *in silico* analysis of the non-coding moiety of all known viral genomes, now uncovers the unprecedented perfect conservation of a AAAATTGA motif in close to 50% of the Mimivirus genes. This motif preferentially occurs in genes transcribed from the predicted leading strand and is associated with functions required early in the viral infectious cycle, such as transcription and protein translation. A comparison with the known promoter of unicellular eukaryotes, in particular amoebal protists, strongly suggests that the AAAATTGA motif is the structural equivalent of the TATA box core promoter element.

This element is specific to the Mimivirus lineage, and may correspond to an ancestral promoter structure predating the radiation of the eukaryotic kingdoms. This unprecedented conservation of core promoter regions is another exceptional features of Mimivirus, that again raises the question of its evolutionary origin.

INTRODUCTION

The recent discovery and genome analysis of *Acanthamoeba polyphaga* Mimivirus, the largest known DNA virus, challenged much of the accepted dogma regarding viruses [1,2]. Its particle size (>400 nm), genome length (1.2 million bp) and huge gene repertoire (911 protein coding genes) all contribute to blur the established boundaries between viruses and the smallest parasitic cellular organisms. Phylogenetic analysis placed the evolutionary origin of Mimivirus prior to the emergence of the extant eukaryotic kingdoms, raising the possibility that large DNA viruses may define a domain distinct from the three other domains of life, Eucarya, Archea and Bacteria [2]. The exceptionally large gene content of the Mimivirus genome, which includes key protein translation genes as well as a very diverse set of enzymes belonging to different metabolic pathways, is consistent with the hypothesis that Mimivirus (and other large DNA viruses) evolved from a free living organism through a genome reduction process akin to the one experienced by parasitic intracellular bacteria [3-5]. However, less radical scenarios have been proposed, such as the recruitment of numerous host acquired genes complementing a set of core genes common to all Nucleo-cytoplasmic large DNA viruses (NCLDVs) [6]. In the latter case, the acquired open reading frames would require to be transferred with its own promoter, or be put under the control of a suitable viral promoter to function properly. These questions about the origin of the Mimivirus genes prompted us to systematically analyze the DNA sequences immediately 5'upstream of each of Mimivirus ORFs in search for transcriptional motifs. Unexpectedly, this analysis revealed the presence of the perfectly conserved octamer AAAATTGA in the putative core promoter regions of near half of all Mimivirus genes. The same analysis, applied to the genomes of other large DNA viruses, confirmed that such an homogeneity of the core promoter sequences is a unique characteristics of Mimivirus.

RESULTS & DISCUSSION

A highly conserved motif in the 5'upstream regions of Mimivirus genes

The annotation of the Mimivirus genome sequence defines 911 predicted protein coding genes, plus an additional set of 347 less convincing unidentified reading frames (URFs).

Mimivirus intergenic regions have an average size of 157 nt with a standard deviation of 113 nt (excluding the two-tailed 5% most extreme data points). Starting from the predicted gene map, we searched for conserved motifs within the 150-nucleotide regions upstream of each of the predicted translation start codon (ATG) of the 911 genes. These sub-sequences were (i) searched for statistically over-represented short oligomers, and (ii) using the Gibbs sampler algorithm implemented in MEME [7].

In contrast to the common difficulty of extracting well conserved signals from eukaryotic promoters [8], our results were unexpectedly clear cut: we found that 403 of the 911 (45%) predicted Mimivirus genes exhibited a strictly conserved AAAATTGA motif within their 150-nucleotide upstream region. The statistical significance of this finding in the context of the nucleotide composition of Mimivirus genome was assessed as follows. We cut the Mimivirus genome into 15752 consecutive 150-nucleotide long segments (using both strands) and counted the occurrences of the above motif in each of them. Only 661 (4%) of these segments were found to contain the AAAATTGA motif at least once. Such a strong preferential occurrence of this motif in the 5'upstream region of Mimivirus genes is indeed highly significant (Fisher Exact $p < 10^{-194}$, for the table [(403, 911)(661, 15752)]. Overall, 60% of the occurrences of this motif are located in front of a gene. Moreover, the distribution of the motif within these 5' upstream regions is non random, with most of them being located in a narrow range, between 50 and 110 nucleotides from the translation start (Figure 1). A sequence logo [9] including the flanking regions of these AAAATTGA motifs shows that they are preceded by 2-3 AT-rich positions and followed by 8-9 AT-rich positions (Figure 2).

The same set of 150-nucleotide upstream regions was also analyzed using a more sophisticated Gibbs sampler approach, as implemented in MEME [7]. A position-weight matrix (PWM) that corresponds to a motif very similar to the previous AAAATTGA was identified (MEME motif; Figure S1). Using a score cutoff of 1000 (based on the bi-modal distribution of the PWM scores; supplementary Figure S2), 446 genes (49%) with a motif were detected. For comparison, only 464 (3%) of the above 15752 consecutive genomic segments exhibit the MEME motif (score \geq 1000). Most of them are thus found upstream of a gene coding region. (Fisher Exact p-value $< 10^{-280}$). Finally, it is worth to point out

that no other predominant motifs were found in the upstream regions of genes that do not contain a AAAATTGA motif.

The motif is less frequent in the more hypothetical URFs

The same analysis was performed with the 347 more unlikely genes (annotated as URFs) in the Mimivirus genome. Only 20 (6%) of them exhibited the AAAATTGA motif in their 150-nucleotide upstream region. Moreover, many of them did not fall within the [-110, -50] range, or were not flanked by AT rich positions. Consistently, only 11 (3%) of the URFs exhibited the previously defined MEME motif (score>1000). This motif distribution is not significantly different from the one observed in the 15752 consecutive 150-nucleotide long segments covering the whole genome.

The presence/absence of the motif does not correlate with the intergenic distance

We verified that the occurrences of the AAAATTGA motif were not trivially linked to the distance between successive ORFs. Figure 3 shows that the proportion of genes exhibiting the motif is fairly constant across the whole range of intergenic distances, except for the smallest ones (< 60 nt). It is worth to notice that a fraction of these short distances might be artifactual, corresponding to cases where the proximal ATG does not coincide with the actual translation start. The net result of this artifact could be to slightly underestimate the number of genes exhibiting the AAAATTGA motif.

The motif significantly correlates with genes transcribed from the leading strand

We then examined other possible correlations between the presence of the AAAATTGA motifs and the positions of the corresponding genes within the genome. Overall, the distribution of Mimivirus protein coding genes does not exhibit any significant strand bias: 450 genes are found on the positive strand (R-genes) and 461 on the negative strand (L-genes; Fisher Exact p-value = 0.8). The distribution of the AAAATTGA motif is similarly unbiased with 196 occurrence in front of the R-genes and 207 in front of the L-genes. The MEME motif exhibits a similar distribution with 217 in front of R-genes and 229 in front of L-genes (score>1000). There is thus no significant global strand preference for the occurrence of the upstream motif.(Fisher Exact p-value = 0.8).

However, a previous analysis [2] identified a putative origin of replication (OR) of the chromosome near position 380,000 (between genes L294 and L295). On the basis of this prediction, one can distinguish a “leading” strand with 578 genes transcribed away from the OR, and a “lagging” strand from which 333 genes are transcribed. Interestingly, the AAAATTGA motif occurs significantly more frequently (Fisher Exact p-value = 0.027) in front of the genes transcribed from the leading strand ($281/578 = 48.6\%$) than from the genes transcribed from the lagging strand ($122/333 = 36.6\%$). A similar asymmetry is found for the MEME motif ($313/578=54.2\%$ and $133/313 = 39.9\%$; Fisher Exact p-value = 0.015).

We also examined whether the presence of AAAATTGA motif correlates with certain categories of gene function (Table 1). 157 Mimivirus genes can be associated to one of the COG [10] functional classes. We found that most translation- and transcription-related genes exhibit the AAAATTGA motif. This was also true of genes related to nucleotide transport and metabolism. In contrast, the motif was absent from the upstream region of most genes related to DNA replication, recombination, and repair as well as from genes classified in the cell envelope biogenesis / outer membrane COGs. Overall, the upstream motif does not occur more frequently in front of genes associated with a functional annotations ($88/232 = 38\%$) compared to anonymous ones ($315/679 = 46\%$) (Tables S1).

NCLDV core genes and the AAAATTGA motif

Iyer et al. [6] identified a set of homologues genes which have been identified in all or most members of the four main NCLDV groups: Iridoviridae, Asfarviridae, Phycodnaviridae and Poxviridae. Some of these “core” genes are also found in baculoviridae and phages. These core genes are divided in 4 classes, from the most to the least conserved. Remarkably, we found that none of the 9 class I core genes found in Mimivirus have the exact octamer motif in their 5' upstream region, and only 2 exhibit the MEME motif (Table S2). In contrast, all but one of the six class II core genes have the octamer motif and the MEME motif. The motif distribution for the class III (7 /11) and Class IV (10/16) core genes is more balanced. However, none of the above distributions significantly differ from 446/911 ratio observed for all Mimivirus genes

(Fisher exact test p-value>0.1).

The presence of such a highly conserved 5' upstream motif is unique to Mimivirus

For many years, the 5' upstream region of eukaryotic genes has been under intense scrutiny in many different organisms from different kingdom (e.g. fungi, plant and metazoan) in an attempt to decipher the sequence-based signal involved in the initiation and the regulation of the transcription process[11,12]. The sole common result that emerged from these numerous studies is that sequence conservation is the exception rather than the rule in the 5'upstream regions of eukaryotic genes [13,14]. Other recent studies have confirmed this lack of conservation in several lineage of parasitic protists [15]. However, large eukaryotic DNA viruses have been much less studied than their cellular counterparts. The analysis as applied to the Mimivirus genome was thus performed on the genomes of all fully sequenced NCLDVs of the Iridoviridae, Phycodnaviridae, and Poxviridae families, to establish the pattern of sequence conservation in the 5'upstream regions of their genes. Our results show that none of these other NCLDVs exhibit a pattern of conservation comparable to the one observed in Mimivirus (Table S3). For instance, only 30 out of 178 (17%) genes of the *Invertebrate iridescent virus 6* (CIV, Iridoviridae) have a conserved AAAATTGA motif within their 150 nt upstream region. 14 out of 231 (6%) *Fowlpox* (FOP, Poxviridae) genes, and 10 of 218 (4.5%) *Amsacta moorei entomopoxvirus* (AME, Poxviridae) genes also exhibit this motif. In all other NCLDVs less or no occurrences of this motif were detected. Note however, that 47 of the 218 (22%) AME genes display a conserved TTTTGAAA motif (Table S3). Finally, a similar analysis by Gibbs sampling of all viral genomes containing more than 100 annotated genes showed that the Mimivirus pattern of 5'upstream motif conservation is truly unique among large DNA viruses.

The conserved AAAATTGA motif is likely to be a main core promoter element

Analyses of the structure and expression of a number of genes from amoebal protists have shown that they are expressed in single transcription units, and that few of them have introns [15]. As the mechanisms of gene expression used by a virus and its host must be compatible, it is reasonable to propose that the short genome regions (157 nt average) separating two consecutive Mimivirus ORFs contain most of the promoter

sequence information. The eukaryotic core promoter includes DNA elements that can extend 35 bp upstream and/or downstream of the transcription initiation site. Most core promoter elements appear to interact directly with components of the basal transcription machinery. In metazoan, the most conserved and recognizable core promoter element is the TATA box, located 25 to 30 bp of the transcription start site, and present in about one third of the human genes [16]. The average position (-60 from the predicted initiator ATG) of the conserved octamer found in Mimivirus is consistent with it playing a role similar to the TATA box for the expression of the viral genes, provided the 3'untranslated regions (5'UTR) of viral mRNA are about 30 nucleotide-long in average. Such a compact promoter/3'UTR structure has been observed in amitochondriate protists such as *Giardia intestinalis* or *Entamoeba histolytica* [15]. The sequences of the TATA box-like element of these protozoa are also different from the 5'-TATATAAG-3' consensus identified in the other eukaryotic kingdoms. Accordingly we propose that the AAAATTGA motif found in 50% of Mimivirus intergenic regions is the virus TATA box-like motif.

Mimivirus TATA box-like motif is not prevalent in amoebal organisms

Interestingly, the Mimivirus TATA box-like motif does not bear a particular resemblance (if any) with the different TATA box-like consensus sequences that have been identified in various protozoan. For instance, *E. histolytica* TATA box-like consensus is TATTTAAA [15]. Using the available data from a genomic survey sequencing of *Acanthamoeba castellanii* (Anderson, I.J. and Loftus, B.J., Gene discovery in the *Acanthamoeba castellanii* genome, unpublished; ~19Mb sequence data available at <http://www.ncbi.nlm.nih.gov/>), we verified that the AAAATTGA motif is not particularly prevalent in the genome of the closest (partially sequenced) relative of the Mimivirus host *Acanthamoeba polyphaga*. In addition, no significant difference in codon usage between the sets of genes harboring or not harboring the AAAATTGA motif could be detected. Symmetrically, of the 29 Mimivirus genes most likely to have been acquired by lateral transfer (Ogata et al., submitted), 15 exhibits the AAAATTGA motif and 14 do not have it. These results argue against the hypothesis that this promoter element, together with the large proportion of associated Mimivirus genes, were simply acquired from its host. Interestingly clusters of paralogous genes that have been produced by

multiple round of duplications from an ancestral Mimivirus gene conserved the AAAATTGA motifs amidst the divergence of their respective promoter regions. The example of the large gene cluster L175-L185 is shown in Figure 4.

The two types of Mimivirus promoters may correspond to early vs. late functions

Homologues of the 3 main proteins involved in the formation of the transcription pre-initiation complex have been identified in Mimivirus genome: the two RNA polymerase II subunits (Rpb1 & Rpb2) and the TFIID (TATA-box binding) initiation factor. The corresponding amino acid sequences are extremely distant from their closest matches: *Candida albicans* Rpb1 (34% identical), *Dictyostelium discoideum* Rpb2 (36% identical) and *Plasmodium falciparum* TFIID (24% identical). We propose that Mimivirus unique TATA box-like sequence AAAATTGA may have co-evolved and be recognized by a pre-initiation complex including these divergent Mimivirus proteins. A second type of promoter – that has not yet been identified and may be highly degenerated – might then be recognized by the host pre-initiation complex, or involving a combination of Mimivirus and host encoded transcription factors. This hypothesis is consistent with the preferential occurrence of the AAAATTGA (type one promoter) in front of Mimivirus genes encoding functions required for the early phase of viral infection (transcription, nucleotide transport, and protein translation; Table 1). According to this scenario, the corresponding genes could be transcribed in the host cytoplasm. Conversely, the AAAATTGA motif is preferentially absent from the promoter of Mimivirus genes encoding “late” functions such as DNA replication and particle biogenesis and assembly (Table 1). These genes could be transcribed in the host nucleus.

CONCLUSION

Our bioinformatics and comparative genomics study revealed a unique feature of Mimivirus within the eukaryotic domain: the presence of a highly conserved AAAATTGA motif in the immediate 5' upstream region of close to 50% of its protein encoding genes. By analogy with the known promoter structures of unicellular eukaryotes, in particular amoebal organisms, we propose that this motif corresponds to a TATA box-like core promoter element. This element, and its conservation, appear to be

specific of the Mimivirus lineage, and might correspond to an ancestral promoter structure predating the radiation of the eukaryotic kingdoms [2]. Mimivirus genes exhibiting this type of promoter might be ancestral as well. Interestingly, this is the case of all translation apparatus- related genes for the first time identified in a virus (four amino-acyl tRNA synthetases, mRNA cap binding protein, translation factor eF-Tu, and tRNA methyltransferase), as well as Mimivirus Topoisomerase 1A (bacterial type). However, it is also possible, although less likely, that horizontally acquired ORFs could have been inserted downstream of a preexisting AAAATTGA motif.

More genomic data on amoebal species, other protozoa, and their associated viruses is now needed to better understand the evolutionary scenario through which the unique promoter structure of Mimivirus genes might have emerged. Our findings can now be used to guide the experimental characterization of Mimivirus transcription units, and identify the transcription factors involved in the recognition of its uniquely conserved promoter element.

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TABLES AND FIGURES

Table 1 : Number of ‘COGed’ Mimivirus genes with or without conserved AAAATTGA motif (only COG classes that contain at least 3 Mimivirus genes are shown). Numbers corresponding to genes with or without a MEME motif are given in parentheses.

Figure 1 : The distribution of the position of the AAAATTGA motif with respect to the predicted gene start shows the location of the conserved element around 50-110 nt..

Figure 2 : The sequence logo (based on 400 genes with a strictly conserved AAAATTGA motif ; 3 genes with a motif that is less than 20 nt from the predicted translation start were not included in the computation of this logo) shows the conserved AT-rich neighborhood of the exactly conserved AAAATTGA octamer (see Figure S1 for the corresponding logo of the MEME motif).

Figure 3 : No relation between the presence/absence of the AAAATTGA motif and the distance between two genes can be identified in the size distribution of Mimivirus intergenic regions (except for the virtual absence of the motif in intergenic regions that are too short in order to host it).

Figure 4 : The alignment of the intergenic region of the paralogous gene cluster L175-L185 shows the perfect conservation of the AAAATTGA motif within intergenic regions that have otherwise more diverged. Note that the AAAATTGA motif is part of the C-terminal region of gene L185 (indicated by white X’s).

SUPPLEMENTARY TABLES AND FIGURES

Table S1 : List of functionally annotated Mimivirus genes. Genes with an AAAATTGA and a MEME motif in the 150nt upstream region are marked by ‘X’; genes that display only a AAAATTGA motif by ‘A’; genes with only a MEME motif by ‘M’.

Table S2 : List of Mimivirus NCLDV core gene set members with and without the AAAATTGA motif; annotation of the presence or absence of a motif is as in Table S1.

Table S3: Occurrence AAAATTGA-related conserved 8-mers in other large DNA viruses.

Figure S1: MEME sequence logo for the 446 genes having a PWM score > 1000 .

Figure S2 : Distribution of the PWM scores (496 out of 911 genes have a score > 0 ; 446 have a score > 1000). The bi-modality of the distribution supports the choice of our score cutoff at 1000.

Table 1

with motif	without motif	Total	COG function
7 (9)	23 (21)	30	DNA replication, recombination, and repair
0 (0)	9 (9)	9	Cell envelope biogenesis, outer membrane
3 (3)	5 (5)	8	Amino acid transport and metabolism
5 (5)	11 (11)	16	Posttranslational modification, protein turnover, chaperones
1 (1)	2 (2)	3	Lipid metabolism
34 (38)	26 (22)	60	Function unknown or general function prediction only
2 (2)	1 (1)	3	Secondary metabolites biosynthesis, transport, and catabolism
5 (5)	0 (0)	5	Nucleotide transport and metabolism
7 (7)	2 (2)	9	Transcription
6 (5)	1 (2)	7	Translation, ribosomal structure and biogenesis

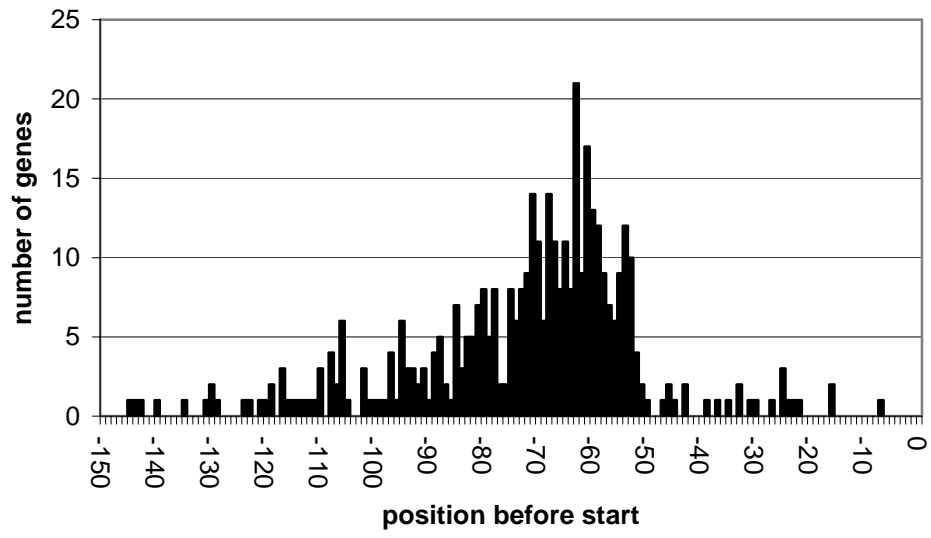


Figure 1

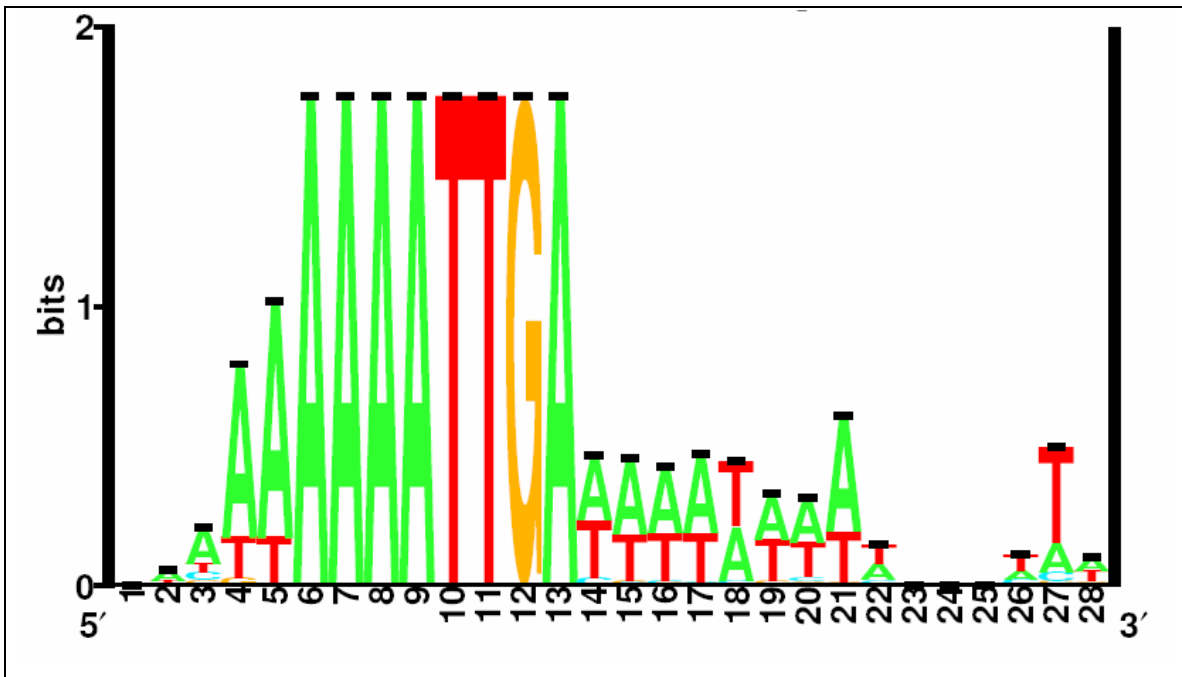


Figure 2

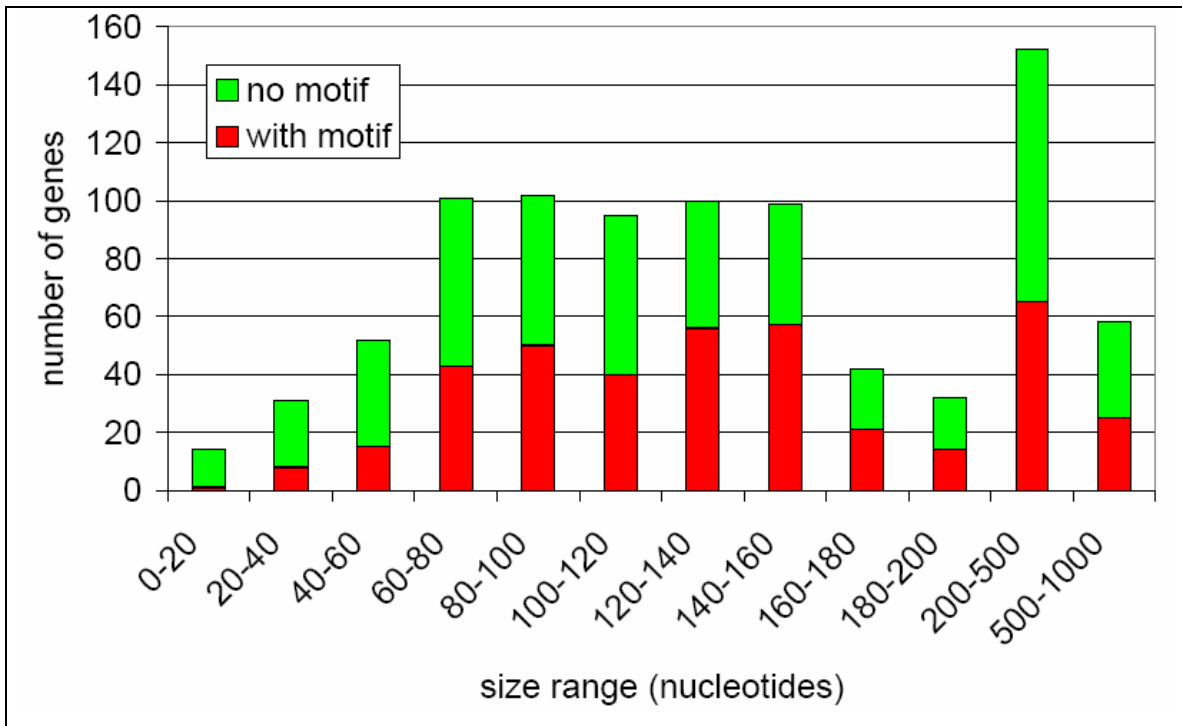


Figure 3

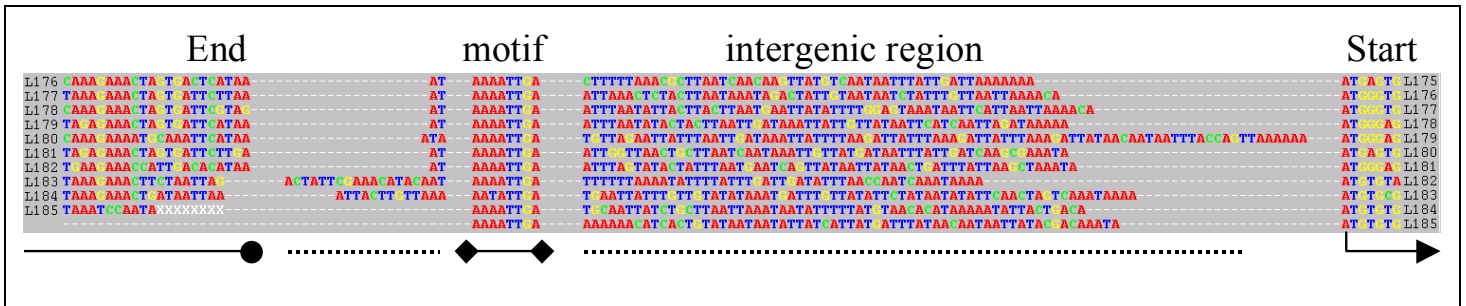


Figure 4

Table S1

Gene	Motif	Annotation
R1		Replication Origin binding protein
L2		Bro family N terminal domain containing protein
L4	X	KilA N-terminal Domain, N1R P28 DNA binding
R8		Helicase similar to Origin binding protein
L30	A	KilA N-terminal Domain, N1R P28 DNA binding
L32	X	KilA N-terminal Domain, N1R P28 DNA binding
L33	X	KilA N-terminal Domain, N1R P28 DNA binding
L37	X	BroA, KilA-N term
R61	X	WD-repeat family protein
L65		Virion-associated membrane protein
L71		Collagen triple helix repeat containing protein
L79		putative transposase
R80		putative resolvase
L84		WD-repeat family protein
L103		putative resolvase
R104		putative transposase
L108	X	Proliferating cell nuclear antigen (PCNA)
L124	X	Tyrosyl-tRNA synthetase
L128	X	Choline dehydrogenase
R135		Choline dehydrogenase or related protein
L136		Sugar transaminase dTDP-4-amino-4,6-dideoxyglucose biosynthesis
L137		contains glycosyl-transferase domain (PFAM)
L138		Contain glycosyl-transferase domain (PFAM)
R139		contains glycosyl-transferase domain (PFAM)
R141		GDP mannose 4,6-dehydratase (hydroxysteroid dehydrogenase?)
L142		contains hexapeptidase domain (PFAM)
L153		similar to A273L from Paramecium bursaria Chlorella virus
L164	X	Cysteinyl-tRNA synthetase
R186	X	putative transposase
L193		putative glycosyl-transferase
R194		Topoisomerase I (pox-like)
R195		Glutaredoxin (ESV128 type)
R196		Collagen triple helix repeat containing protein
R197		Probable cytidine deaminase
L205	X	Contains protein kinase domain (PFAM)
L206		Helicase III VV D5-type ATPase (C-term)
L207	X	Helicase III VV D5-type ATPase (N-term)

R209	M	probable RNA polymerase subunit 6
R214		RAS family GTPase
L221	X	Topoisomerase I (bacterial type)
L230		Procollagen-lysine,2-oxoglutarate 5-dioxygenase
L232	M	Contains protein kinase domain (PFAM)
L233		putative Zn-dependent peptidase
L235	A	RNA polymerase subunit 5
R239		Collagen triple helix repeat containing protein
R240		Collagen triple helix repeat containing protein
R241		Collagen triple helix repeat containing protein
L244	X	Rpb2
L245		HNH endonuclease
L247	X	HNH endonuclease
L249	X	putative uracil-DNA glycosylase
L250	X	putative transcription initiation factor IIB
L251		Lon domain protease
L254		Heat shock 70Kd HSP
L258	X	Thymidine kinase
R259	X	Contains DUF549 domain (PFAM)
R260	X	Dnaj-like protein
R266		Dnaj-like protein
L268	X	Contains protein kinase domain (PFAM)
L275		Contains K ⁺ channel tetramerisation domain (PFAM)
L276	X	Putative oxoglutarate malate carrier protein
R296		Endonuclease IV
R299		contain Ribonuclease H domain (PFAM)
L300		contains Endo excinuclease amino terminal domain (PFAM)
R301		Uncharacterized protein (Chilo iridescent virus 380R)
R303		NAD-dependent DNA ligase
R307		Contains Protein phosphatase 2C domain (PFAM)
R311	X	BIR domain (Chilo iridescent virus 193R)
L312	X	Ribonucleotide reductase small subunit
R313	X	Ribonucleotide reductase large subunit
L315		Hydrolysis of DNA containing ring-opened N7 methylguanine
L316		Glucosamine 6-phosphate N-acetyltransferase
L318		DNA polymerase family X
R319	X	putative ubiquitin-specific protease
R322	M	DNA polymerase (B family)
L323		Myristoylated virion protein A
R325		Metal-dependent hydrolase (Chilo iridescent virus 136R)
R339	X	TFII-like transcription factor

R343	X	RNAse III (dsRNA binding motif)
L344	X	WD-repeat family protein
R350		putative transcription termination factor, VV D6R helicase
R354		Lambda-type exonuclease
L359		DNA mismatch repair ATPase MutS
R362		contain Thioredoxin domain (PFAM)
R363		Contains glycosyl-transferase domain (PFAM)
L364	X	SW1 SNF2 helicase (MSV224)
R366		Contains helicase domaine (PFAM)
R368		putative thiol oxidoreductase
L373		putative glycosyltransferase
L377		putative NTPase I
R380		Contains Leucine Rich Repeat domain (PFAM)
R382	X	mRNA Capping Enzyme
L386	X	putative endonuclease of the XPG family
L388	X	contains Endonuclease Exonuclease phosphatase domain (PFAM)
L393		Bos Taurus HSP70
R395	X	similar to EsV-1-87 [Ectocarpus siliculosus virus]
L396	M	VV A18 helicase
R398		Contains Calcineurin-like phosphoesterase domain (PFAM)
R400		S T protein kinase, similar to Paramecium bursaria chlorella virus 1 A617R
R405	X	tRNA (Uracil-5-)-methyltransferase
R406	X	Alkylated DNA repair
R407	X	tRNA (uracil-5-)-methyltransferase
L410		Similar to poxvirus P4B major core protein
R418	X	NDK synthesis of nucleoside triphosphates
R419		putative dehydrogenase
L425		Capsid protein D13L (4 paralogs)
R429		PBCV1-A494R-like, 9 paralogs
R431	X	Contains 3'-5' exonuclease domain (PFAM)
L432		Predicted HD phosphohydrolase
R436	X	Putative serine threonine protein kinase
L437		VV A32 virion packaging ATPase
R439		Capsid protein
R440		Capsid protein
R441		Capsid protein
R443		Contains thioredoxin domain (PFAM)
L444		Contains ADP-ribosylglycohydrolase domain (PFAM)
R445		Contains Dnaj domain (PFAM)
L446		Patatin-like phospholipase (463L)
R449		Uncharacterized protein (Paramecium bursaria)

R450		All transcription factor
R453	X	TATA-box binding protein (TBP)
R458	X	putative translation initiation factor
L460	X	ubiquitin-conjugating enzyme
R464		Translation initiation factor SUI1
L469	M	Polynucleotide phosphatase kinase
R470	M	DNA directed RNA polymerase subunit L
R475	X	Asparagine synthase (glutamine hydrolysing)
R476	X	Contains ATPase domain (PFAM)
L477		Cathepsin B, Cystein protease
L478	X	putative replication factor C
R480	X	Topoisomerase II
R486		Contains 2 PAN domains (PFAM)
R493	X	Proliferating Cell Nuclear Antigen
L496	A	Translation initiation factor 4E, (mRNA cap binding)
R497	X	Thymidylate synthase
L498		Zn-dependent alcohol dehydrogenase
L499	X	putative replication factor C
R501	X	Rpb1
R510	X	putative replication factor C subunit
L511	A	Contains FtsJ-like methyltransferase domain (PFAM)
R512	X	Deoxynucleotide monophosphate kinase
R517		Contains protein kinase domain (PFAM)
R518		Contains protein kinase domain (PFAM)
R519		putative peptidase, contains peptidase M13 domain (PFAM)
L524	X	MuT-like NTP pyrophosphohydrolase
R526		putative triacylglycerol lipase
R530	M	putative DNA helicase
L532	X	Contains cytochrome p450 domain (PFAM)
L538		Helicase conserved C-terminal domain (PFAM)
L540		VVI8 helicase
R541	X	K ⁺ channel tetramerisation domain
L543	X	ADP-ribosyltransferase (DraT)
L544		Transcription initiation factor TFIIB
R548		Contains thioredoxin domain (PFAM)
R555		putative DNA repair protein
R563		Contains helicase conserved C-terminal domain (PFAM)
R565		Glutamine synthetase (Glutamate-amonia ligase)
R568	X	Contains UvrD REP helicase domain (PFAM)
R569		Contains exonuclease domain (PFAM)
R571		Patatin-like phospholipase (similar to Chilo iridescent virus 463L)

L572		Contains ATPase domain (PFAM)
R592	X	Contains helicase conserved C-terminal domain (PFAM)
L593		prolyl 4-hydroxylase
R595		putative phospholipase carboxylesterase family protein
R596		Thiol oxidoreductase E10R
L605		peptidylprolyl isomerase
R610		proline rich protein
L612		Mannose-6P isomerase
R614	X	Contains SPFH domain Band 7 family (PFAM)
L615		Contains FYVE zinc finger domain (PFAM) and
L619		putative glucosamine--fructose-6-phosphate aminotransferase
L620		Contains Patatin-like phospholipase domain (PFAM)
L621		putative N-myristoyltransferase
R622		Dual specificity S Y phosphatase
R624	X	GTP binding elongation factor eF-Tu
L628		Cyt-b5, Cytochrome b5-like Heme Steroid binding domain
L630		putative Ubiquitin-conjugating enzyme E2
R632		putative N-acylsphingosine amidohydrolase
R639	X	Methionyl-tRNA synthetase
R644		putative Phosphatidylethanolamine-binding protein (PFAM)
R654		Contains glycosyltransferase domain (PFAM)
R655		Contain Glycosyltransferase domain (PFAM)
R663	X	Arginyl-tRNA synthetase
R665	X	putative oxidoreductase
L668		Collagen triple helix repeat containing protein
L669		Collagen triple helix repeat containing protein
L670		Contains Protein kinase domain (PFAM) and Cyclin, N-terminal domain (PFAM)
L673	X	Contains protein kinase domain (PFAM) and Cyclin, N-terminal domain (PFAM)
L687		Endonuclease for the repair of UV-irradiated DNA
R689		N-acetylglucosamine-1-phosphate uridylyltransferase
R693	X	Methylated-DNA-protein-cysteinemethyltransferase
R700		Contains Serpin (serine protease inhibitor) domain (PFAM)
R707		P13-like protein
R708		Fructose-2,6-bisphosphatase
L709	X	Contains Ubiquitin-conjugating enzyme domain (PFAM)
L716		GMP synthase (Glutamine-hydrolyzing)
L720		Hydrolysis of DNA containing ring-opened N7 methylguanine
R721		Similar to CheD, Chemotaxis protein
R726		Peptide chain release factor eRF1
R730		Contains ATPase domain (PFAM)
L733	X	ABC transporter ATP-binding domain

R739	X	WD-repeat family protein
R749	X	Contains homeobox domain (PFAM)
R756	X	similar to Predicted Fe-S-cluster redox enzyme (COG)
L770		putative transposase
R771		putative resolvase
L775		Contains 2 PAN domains (PFAM)
L780		dTDP-4-dehydrorhamnose reductase
L783	A	WD-repeat family protein
R795	X	Contains ubiquitin-conjugating enzyme domain (PFAM)
L805		MACRO domain (splicing related)
R807	M	7-dehydrocholesterol reductase
L808	X	Lanosterol 14-alpha-demethylase
R818		Contains 2 protein kinase domains (PFAM)
L823	X	Proliferating Cell Nuclear Antigen
R824	X	Putative 5'(3')-deoxyribonucleotidase
R826	X	Contains 2 protein kinase domains (PFAM)
R831	X	Contains 2 protein kinase domains (PFAM)
R832	X	putative alcohol dehydrogenase (N-term)
R833		putative alcohol dehydrogenase (C-term)
R854		putative transposase
R856	X	Contains 6 TPR domains (PFAM)
R877		putative outer membrane lipoprotein
R878	X	KilA N-terminal Domain, N1R P28 DNA binding
R879	M	KilA N-terminal Domain, N1R P28 DNA binding
R892	X	putative oxidoreductase (C-term)
L893		putative oxidoreductase (C-term)
L894		putative oxidoreductase (N-term)
R904	X	KilA N-terminal Domain, N1R P28 DNA binding
L905		putative methyl-transferase
L906	X	Acetylcholinesterase

Table S2

Class	Gene	Motif	Annotation
I	L206		Helicase III VV D5-type ATPase (C-term)
I	L396	M	VV A18 helicase
I	L425		Capsid protein D13L (4 paralogs)
I	L437		VV A32 virion packaging ATPase
I	R322	M	DNA polymerase (B family)
I	R350		putative transcription termination factor, VV D6R helicase
I	R400		S T protein kinase, similar to Paramecium bursaria chlorella virus 1 A617R
I	R450		A1L transcription factor
I	R596		Thiol oxidoreductase E10R
II	L312	X	Ribonucleotide reductase small subunit
II	L323		Myristoylated virion protein A
II	L524	X	MuT-like NTP pyrophosphohydrolase
II	R313	X	Ribonucleotide reductase large subunit
II	R339	X	TFII-like transcription factor
II	R493	X	Proliferating Cell Nuclear Antigen
III	L244	X	Rpb2
III	L364	X	SW1 SNF2 helicase (MSV224)
III	L37	X	BroA, KiA-N term
III	L65		Virion-associated membrane protein
III	R195		Glutaredoxin (ESV128 type)
III	R311	X	BIR domain (Chilo iridescent virus 193R)
III	R382	X	mRNA Capping Enzyme
III	R429		PBCV1-A494R-like, 9 paralogs
III	R480	X	Topoisomerase II
III	R501	X	Rpb1
III	R622		Dual specificity S Y phosphatase
IV	L235	A	RNA polymerase subunit 5
IV	L258	X	Thymidine kinase
IV	L32	X	KiA N-terminal Domain, N1R P28 DNA binding
IV	L33	X	KiA N-terminal Domain, N1R P28 DNA binding
IV	L37	X	BroA, KiA-N term
IV	L4	X	KiA N-terminal Domain, N1R P28 DNA binding
IV	L446		Patatin-like phospholipase (463L)
IV	L477		Cathepsin B, Cystein protease
IV	L540		VVI8 helicase
IV	L805		MACRO domain (splicing related)
IV	R141		GDP mannose 4,6-dehydratase (hydroxysteoid dehydrogenase?)
IV	R301		Uncharacterized protein (Chilo iridescent virus 380R)
IV	R303		NAD-dependent DNA ligase
IV	R325		Metal-dependent hydrolase (Chilo iridescent virus 136R)
IV	R343	X	RNAse III (dsRNA binding motif)
IV	R354		Lambda-type exonuclease
IV	R449		Uncharacterized protein (Paramecium bursaria)
IV	R497	X	Thymidylate synthase
IV	R571		Patatin-like phospholipase (similar to Chilo iridescent virus 463L)
IV	R878	X	KiA N-terminal Domain, N1R P28 DNA binding
IV	R879	M	KiA N-terminal Domain, N1R P28 DNA binding
IV	R904	X	KiA N-terminal Domain, N1R P28 DNA binding

Table S3

	<i>Acanthamoeba polyphaga Mimivirus</i>	<i>Invertebrate iridescent virus 6</i>	<i>Fowlpox virus</i>	<i>Amsacta moorei entomopox virus</i>
annotated genes	911	178	231	218
with AAAATTGA motif	403	30	14	10
chunks (150 nt; both strands)	15754	2834	3848	3100
chunks with motif	664	61	22	56
AAAATTGA (best Mimi)	403	30	14	10
AAATTGAA (best CIV)	206	46	12	14
TTTTGAAA (best AME)	28	18	3	47

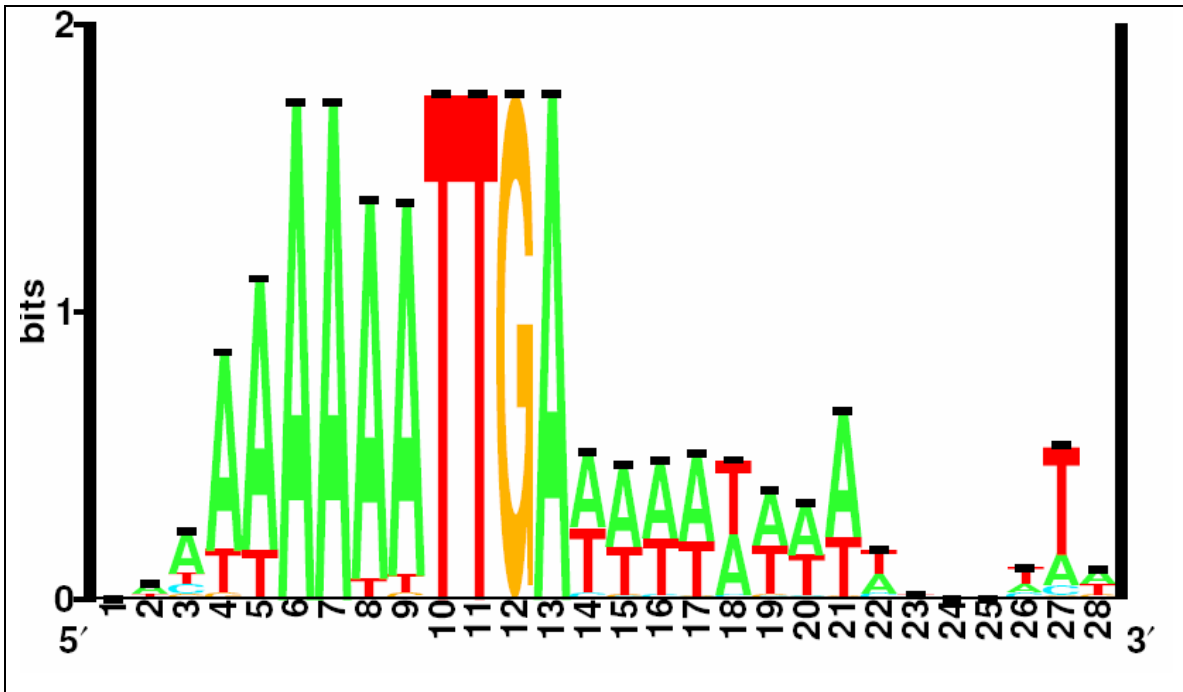


Figure S1

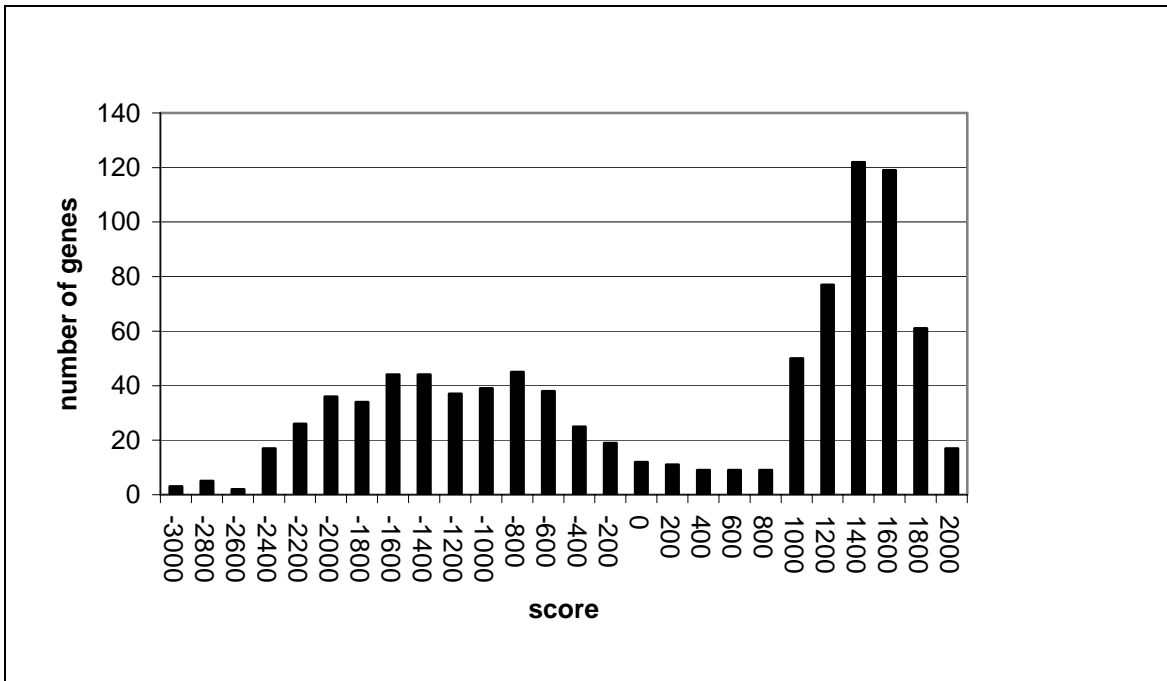


Figure S2