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CYMIAZOLE, A SYSTEMIC ACARICIDE THAT CONTROLS ACARAPIS WOODI (RENNIE) INFESTING HONEY BEES. I. LABORATORY TESTS ¹

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ABSTRACT

Cymiazole (2-[2, 4-dimenthylphenyl-imino]-3-methyl-4-thiazoline-hydrochloride, Apitol[®], 17,5 % cymiazole granules, Ciba-Geigy Ltd.) was dissolved in 50 % sucrose syrup and fed to caged honey bees infested with *Acarapis woodi* (Rennie). Concentrations of 0.88 mg/ml and higher killed about 80 % of adult mites within one week. Lower concentrations took longer. After three weeks the LC_{50} was 0.03 mg/ml. A similar experiment involving cymiazole mixed in queen cage candy gave a comparable level of control (LC_{50} =0.15 mg/g). No adverse effects were observed in bees given effective doses of cymiazole. Lowered comsumption and shortened longevity were observed when concentrations 3-4 times that which gave adequate control were fed.

INTRODUCTION

For reasons that are not clear, Acarapis woodi has spread rapidly throughout Mexico, the United States, and recently localized infestations have been reported in Canada (GUZMAN-NOVOA and ZOZAYA-RUBIO, 1984; DELFINADO-BAKER, 1985; Anon. 1986b). In addition to becoming widespread, the prevalence of infested honey bee colonies (*Apis mellifera*) has increased greatly in some regions, especially Mexico (Anon. 1984; GUZMAN-NOVOA and ZOZAYA-RUBIO, 1984). Equally important is the high proportion of these colonies that have > 30 % of their bees parasitized (EISCHEN *et al.* 1988; EISCHEN *et al.*, unpublished). The economic consequences of infestations at this level are substantial (BAILEY, 1961; GUZMAN-NOVOA and ZOZAYA-RUBIO, 1984; EIS-CHEN, 1987; EISCHEN *et al.*, 1989).

Because registered therapeutic treatments are not currently available, legislation was enacted to provide protection for uninfested areas (eg., Anon. 1986a,c,d). However, the large numbers of colonies involved in a highly mobile industry make regulation difficult. Clearly, safe medication would help solve this situation. Recently cymiazole (Apitol[®], Ciba-Geigy), a systemic acaricide has showed promising results in controlling *Varroa jacobsoni* (RITTER, 1985; SCHMID, 1985). Though phylogenetically different, both *V. jacobsoni* and *A. woodi* utilize honey bee hemolymph for food. This suggests the possibility of similar physiology and response to an acaricide.

This report details laboratory experiments designed to evaluate the efficacy of cymiazole for controlling A. woodi infesting worker honey bees.

MATERIALS AND METHODS

Tests were conducted during the summer of 1986 in the facilities of Apicultura Cardoso, which is located in Buenavista Allende, Nuevo Leon, Mexico (lat. 100° 01' north, long. 25° 17' west). Bees of mixed Italian stock (*Apis mellifera ligustica* Spin.) were obtained locally from colonies heavily infested (70-100 %) with *A. woodi*. Severely infested bees were selected for testing as they are likely to be stressed by the parasite load. If medication exerts an additional stress then it is most easily detected in these bees. This serves to set the upper limits on safe dosages. Secondly, heavy parasite loads present a worst case. If medication is efficacious, then it is also likely to be so at lower levels.

All trials were conducted in laboratory test cages fitted with a small piece of comb (KULINČEVIĆ and ROTHENBUHLER, 1973). Thirty or 35 bees were placed in each cage, and one cage was set up for each treatment (i.e. concentration). Mite-infested bees were obtained by removing worker bees from the inner surface of the hive cover of a heavily infested colony. Young uninfested bees were obtained by placing combs of emerging adults in an incubator and collecting bees 24 hours later. Treatments consisted of 50 % sucrose syrup (vol./vol.) or solutions of cymiazole (2-[2, 4-dimethylphenyl-imino]-3-methyl-4-thiazoline-hydrochloride) in 50 % sucrose syrup. The commercial preparation of cymiazole is stable in syrup and honey (W.J. SCHMID, pers. commun.) Treatments and distilled water were provided *ad libitum* by means of inverted vials, which had small holes in their caps. All tests were conducted in a darkened incubator (32 ± 1 °C, 80 ± 5 % RH). Cages were examined daily and the dead bees removed and counted.

Experiment 1. Young Bee Longevity

Thirty uninfested worker bees (0-24 hrs.) were placed in a laboratory test cage for each treatment (RINDERER and BAXTER, 1978). Treatments consisted of 0.0, 0.05, 0.11, 0.22, 0.44, 0.88, 1.75, 3.5, and 7.0 mg/ml of cymiazole in 50 % sucrose syrup (12 ml). After 14 days all cages received fresh sucrose syrup and distilled water only. Observations were stopped after 15 bees/cage died, i.e. 50 % of the bees (LT_{50}).

Experiment 2. Old Bee Longevity

Thirty bees from a heavily infested colony (70 %) were placed in a laboratory test cage for each treatment. Treatments consisted of 0.0, 0.11, 0.22, 0.44, 0.88, 1.75, 3.5, and 7.0 mg/ml of cymiazole in 50 % sucrose syrup (12 ml). Fresh syrups and distilled water were given on days 0, 7, and 14. An estimate of syrup hoarding was made by counting the number of comb cells containing syrup on the 14th day of treatment. Observations were stopped after 15 bees/cage had died (LT_{50}).

Experiment 3. LC₅₀ for A. woodi ; Cymiazole in syrup.

Thirty-five bees from a heavily infested colony (100 %) were placed in a laboratory test cage for each treatment. Treatments consisted of 0.0, 0.025, 0.05, 0.11, 0.22, 0.44, 0.88, and 1.75 mg/ml of

cymiazole in 50 % sucrose syrup (12 ml). Fresh syrups and distilled water were given on days 0, 7, and 14. Five bees/treatment were examined on days 7, 14, and 21. Dissections and mortality determinations of A. woodi were made according to EISCHEN *et al.* (1987). Both prothoracic tracheae were inspected and the most heavily infested trachea selected for scoring.

Experiment 4. LC₅₀ for A. woodi ; Cymiazole in queen cage candy.

Thirty-five bees from a heavily infested colony (100 %) were placed in a laboratory test cage for each treatment. Treatments consisted of 0.0, 0.75, 1.0, 1.25, 1.5, 2.0, and 3.0 mg/g of cymiazole in candy (confectioners' sugar and 50 % sucrose syrup, 3.7 :1, see LAIDLAW and ECKERT, 1962). Candy (5.5 g) was pressed into a vial cap and placed on the floor of the test cage. Fresh candies and distilled water were given on days 0 and 7. After 7 days, uneaten candy was weighed. Five bees/treatment were examined on days 7, 14, and 21.

Data were subjected to probit analysis using a SAS program (RAY, 1982) based on FINNEY (1971). Significant differences were determined by nonoverlap of the 95 % fiducial limits.

RESULTS

Experiment 1. Young Bee Longevity

The median life-span of young bees fed 0.05-1.75 mg/ml cymiazole treated syrups was similar to those fed sucrose syrup ($LT_{50} = 21$ days). Bees fed concentrations 3.5 mg/ml or higher had life-spans about one-half that of controls ($LT_{50} = 13$ days; t = 11.8, P < 0.01). In general, longevity was rather short even in control bees.

Experiment 2. Old Bee Longevity

The median life-span of bees fed, 0, 0.05, 0.11, 0.22, 0.44, 0.88, 1.75, 3.5, and 7.0 mg/ml of cymiazole in syrup was 33, 32, 21, 23, 27, 27, 14, 6, and 3 days, respectively. It is not clear why longevity was relatively short for bees fed the 0.11 and 0.22 mg/ml syrups. Bees fed cymiazole in concentrations > 0.88 mg/ml died significantly sooner than controls (t = 5.62, P < 0.01). The shorter life-spans observed at high concentrations may not have been caused totally by toxicity but also involved decreased palatability.

The estimate of hoarding revealed that bees treated with 0, 0.025, 0.05, 0.11, 0.22, 0.44, 0.88, and 1.75 mg/ml cymiazole had stored 50, 58, 77, 40, 42, 31, 43, and 0 cells of syrup, respectively.

Experiment 3. LC_{50} for A. woodi ; Cymiazole in syrup

Concentrations of 0.88 mg/ml or higher were effective after seven days (ca. 80 % adult mite mortality, Table 1). Lower concentrations, eg. 0.05 mg/ml, were not effective until the third week. The lowest concentration (0.025 mg/ml) appeared ineffective. The LC_{50} after three weeks was 0.03 mg/ml,

Treatment	Day	Adults		Larvae		_
		Alive (X ± SE)	Dead (X ± SE)	Alive (X + SE)	$\begin{array}{c} \text{Dead} \\ (\bar{X} + SE) \end{array}$	Eggs $(\bar{X} \pm SE)$
Sucrose control	7 14 21	$\begin{array}{c} 16.2 \pm 1.5 \\ 18.2 \pm 1.4 \\ 9.8 \pm 2.7 \end{array}$	$\begin{array}{l} 4.4 \pm 0.7 \\ 3.6 \pm 0.7 \\ 4.6 \pm 0.6 \end{array}$	8.2 ± 2.4 7.6 ± 2.7 3.4 ± 1.0	$\begin{array}{c} 0.0 \\ 0.2 \pm 0.2 \\ 0.4 \pm 0.2 \end{array}$	$\begin{array}{c} 8.8 \pm 1.5 \\ 6.2 \pm 2.6 \\ 5.4 \pm 3.1 \end{array}$
0.025 mg/ml	7 14 21	$17.4 \pm 2.1 \\ 21.2 \pm 4.4 \\ 21.4 \pm 6.7$	5.8 ± 0.9 7.2 ± 1.5 6.6 ± 1.9	6.8 ± 2.3 8.8 ± 3.1 7.8 ± 1.0	$0.0 \\ 0.0 \\ 0.6 \pm 0.4$	$\begin{array}{c} 6.2 \pm 3.3 \\ 8.2 \pm 2.2 \\ 6.2 \pm 0.9 \end{array}$
0.05 mg/ml	7 14 21	$22.2 \pm 4.5 \\ 32.2 \pm 4.2 \\ 16.8 \pm 3.1$	8.2 ± 2.1 10.6 ± 1.3 40.4 ± 6.2	$\begin{array}{c} 17.8 \pm 0.4 \\ 20.8 \pm 3.6 \\ 7.8 \pm 3.3 \end{array}$	$\begin{array}{c} 0.0 \\ 0.4 \pm 0.2 \\ 0.8 \pm 0.5 \end{array}$	$ \begin{array}{c} 10.0 \pm 3.1 \\ 16.2 \pm 2.3 \\ 2.8 \pm 0.8 \end{array} $
0.11 mg/ml	7 14 21	$\begin{array}{c} 12.8 \pm 2.7 \\ 10.6 \pm 2.3 \\ 13.0 \pm 5.4 \end{array}$	5.4 ± 1.0 10.8 ± 1.7 26.4 ± 3.8	8.0 ± 1.6 6.0 ± 1.0 4.4 ± 1.5	$\begin{array}{c} 0.4 \pm 0.2 \\ 0.4 \pm 0.4 \\ 0.8 \pm 0.4 \end{array}$	$5.4 \pm 2.3 \\ 8.0 \pm 1.1 \\ 3.2 \pm 1.2$
0.22 mg/ml	7 14 21	$17.0 \pm 4.2 \\ 14.6 \pm 4.9 \\ 7.6 \pm 2.1$	6.6 ± 2.2 9.6 ± 2.9 33.6 ± 6.0	$\begin{array}{c} 10.6 \pm 2.2 \\ 3.8 \pm 2.0 \\ 4.2 \pm 1.0 \end{array}$	$\begin{array}{c} 0.0 \\ 2.0 \pm 1.0 \\ 0.2 \pm 0.2 \end{array}$	$\begin{array}{c} 6.0 \pm 2.3 \\ 3.4 \pm 2.0 \\ 2.4 \pm 0.7 \end{array}$
0.44 mg/ml	7 14 21	$ \begin{array}{c} 11.4 \pm 2.0 \\ 8.2 \pm 3.8 \\ 8.4 \pm 3.2 \end{array} $	6.4 ± 1.4 11.2 ± 3.2 24.4 ± 4.8	$\begin{array}{r} 3.2 \pm 0.9 \\ 1.2 \pm 0.6 \\ 1.4 \pm 0.7 \end{array}$	$\begin{array}{c} 0.2 \pm 0.2 \\ 0.4 \pm 0.4 \\ 0.6 \pm 0.4 \end{array}$	$\begin{array}{c} 2.8 \pm 1.7 \\ 0.0 \\ 1.0 \pm 0.5 \end{array}$
0.88 mg/ml	7 14 21	$\begin{array}{c} 3.2 \pm 0.7 \\ 4.8 \pm 1.6 \\ 1.6 \pm 0.5 \end{array}$	$\begin{array}{c} 15.8 \pm 3.5 \\ 13.4 \pm 2.6 \\ 16.4 \pm 5.0 \end{array}$	$\begin{array}{c} 2.2 \pm 0.7 \\ 0.2 \pm 0.2 \\ 0.0 \end{array}$	3.2 ± 0.7 0.0 0.4 ± 0.4	$\begin{array}{c} 1.4 \pm 0.6 \\ 1.0 \pm 0.3 \\ 0.4 \pm 0.4 \end{array}$
1.75 mg/ml	7 14 21	$\begin{array}{c} 1.8 \pm 1.6 \\ 6.0 \pm 2.3 \\ 1.6 \pm 0.5 \end{array}$	16.0 ± 2.8 11.4 ± 3.2 18.2 ± 1.2	$\begin{array}{c} 4.4 \pm 0.9 \\ 0.2 \pm 0.2 \\ 0.0 \end{array}$	$\begin{array}{c} 0.0 \\ 0.4 \pm 0.2 \\ 0.2 \pm 0.2 \end{array}$	$\begin{array}{c} 0.2 \pm 0.2 \\ 0.8 \pm 0.6 \\ 1.6 \pm 0.2 \end{array}$

TABL. 1. — Survival of Acarapis woodi infesting honey bees fed cymiazole in syrup 1

1. Values are mites/trachea (N = 5 bees).

TABL. 2. - Response of adult Acarapis woodi infesting honey bees to cymiazole

Day evaluated	LC ₅₀ (mg/ml)	95 % Fiducial Limits	Slope ± SE	
Syrup				
7	0.29	0.09 -3.13	8.7 ± 0.28	
14	0.27	0.14 -0.77	5.3 ± 0.14	
21	0.03	0.007-0.09	7.2 ± 0.29	
Candy				
21	0.15	0.005-0.36	14.2 ± 0.42	

which was significantly smaller than evaluations made after 7 and 14 days (P = 0.05, Table 2). Additional information was gained by examining mite survival. By the third week, egg production had dropped in all treatments with the exception of the lowest concentration (0.025 mg/ml) and control (Table 1). Similar data for larvae were observed (Table 1). Reduced larval numbers in the higher concentrations may be caused by reduced egg production as well as mortality, though appreciable larval mortality was observed only at the 0.88 mg/ml concentration (59 %, Table 1).

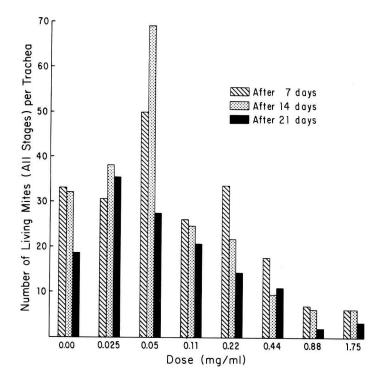


FIG. 1. - Survival of Acarapis woodi (all stages) infesting honey bees fed cymiazole in syrup.

A composite of all surviving life stages is shown in Fig. 1. A decrease in the number of living mites is evident as a factor of both increasing concentration and time. After three weeks, 18.6 mites/trachea were observed in the controls, while less than two living mites/trachea were observed at the 0.88 and 1.75 mg/ml concentrations (Table 1). The latter is likely to be below the economic threshold (EISCHEN, 1987; EISCHEN *et al.*, 1989). Bee mortality after 32 days was 0, 8.6, 0, 0, 14.3, 100, 20, 100, and 100 % when fed syrups containing 0.0, 0.025, 0.05, 0.11, 0.22, 0.44, 0.88, 1.75, and 3.5 mg/ml of cymiazole, respectively.

Treatment	Day	Adults		Larvae		E		
		$\begin{array}{c} \text{Alive} \\ (\bar{X} \pm SE) \end{array}$	$\begin{array}{c} \text{Dead} \\ (\bar{X} \pm \text{SE}) \end{array}$	$\begin{array}{c} \text{Alive} \\ (\hat{X} + SE) \end{array}$	$\begin{array}{c} \text{Dead} \\ (\bar{X} + SE) \end{array}$	$\begin{array}{c} \text{Eggs} \\ (\bar{X} \pm \text{SE}) \end{array}$		
Sucrose control	7 14 21	$\begin{array}{r} 33.0 \pm 7.0 \\ 25.2 \pm 3.0 \\ 23.4 \pm 1.3 \end{array}$	4.8 ± 0.4 14.2 ± 2.8 9.6 ± 2.7	3.4 ± 1.3 10.4 ± 3.4 9.2 ± 1.8	0.0 0.0 0.6 ± 0.2	3.6 ± 2.2 8.8 ± 2.4 4.4 ± 1.0		
0.75 mg/g	7 14 21	$22.2 \pm 4.2 \\ 6.6 \pm 1.9 \\ 4.8 \pm 1.1$	5.6 ± 1.4 10.4 ± 3.1 17.4 ± 3.2	4.6 ± 1.4 1.6 ± 0.8 2.4 ± 0.6	$0.0 \\ 2.0 \pm 0.7 \\ 1.4 \pm 0.2$	$\begin{array}{c} 0.4 \pm 0.4 \\ 1.2 \pm 0.8 \\ 2.2 \pm 0.7 \end{array}$		
1.0 mg/g	7 14 21	$\begin{array}{c} 16.6 \pm 1.6 \\ 9.8 \pm 2.6 \\ 2.0 \pm 1.0 \end{array}$	5.0 ± 0.4 15.8 ± 2.6 13.4 ± 1.8	$\begin{array}{c} 1.2 \pm 0.4 \\ 0.4 \pm 0.4 \\ 0.2 \pm 0.2 \end{array}$	$\begin{array}{c} 0.0 \\ 0.4 \pm 0.2 \\ 0.6 \pm 0.4 \end{array}$	$\begin{array}{c} 1.0 \pm 0.3 \\ 0.4 \pm 0.4 \\ 0.4 \pm 0.4 \end{array}$		
1.25 mg/g	7 14 21	$\begin{array}{c} 21.8 \pm 3.8 \\ 16.0 \pm 6.1 \\ 3.0 \pm 0.5 \end{array}$	5.0 ± 1.1 12.6 ± 2.1 17.6 ± 2.6	$\begin{array}{c} 3.0 \pm 1.0 \\ 0.2 \pm 0.2 \\ 0.0 \end{array}$	$\begin{array}{c} 0.0 \\ 0.2 \pm 0.2 \\ 0.2 \pm 0.2 \end{array}$	$\begin{array}{c} 2.0 \pm 1.3 \\ 0.6 \pm 0.4 \\ 1.2 \pm 0.6 \end{array}$		
1.5 mg/g	7 14 21	$\begin{array}{c} 13.6 \pm 5.3 \\ 9.0 \pm 3.7 \\ 4.4 \pm 1.5 \end{array}$	5.6 ± 1.0 24.0 ± 3.1 26.2 ± 7.5	$\begin{array}{c} 2.0 \pm 0.9 \\ 0.6 \pm 0.6 \\ 2.2 \pm 0.9 \end{array}$	$0.0 \\ 0.0 \\ 0.4 \pm 0.4$	$\begin{array}{c} 1.8 \pm 0.4 \\ 0.8 \pm 0.6 \\ 1.0 \pm 0.5 \end{array}$		
2.0 mg/g	7 14 21	$\begin{array}{c} 24.4 \pm 8.7 \\ 4.6 \pm 2.6 \\ 3.0 \pm 0.5 \end{array}$	7.8 ± 1.2 14.4 ± 3.7 31.4 ± 1.9	$\begin{array}{c} 1.4 \pm 0.9 \\ 0.0 \\ 0.2 \pm 0.2 \end{array}$	$\begin{array}{c} 0.0 \\ 0.4 \pm 0.4 \\ 0.6 \pm 0.2 \end{array}$	$\begin{array}{c} 0.6 \pm 0.4 \\ 0.4 \pm 0.4 \\ 0.0 \end{array}$		
3.0 mg/g	7 14 21	8.0 ± 0.9 11.4 ± 3.1 high bee mo	24.4 ± 5.0 32.2 ± 2.9 rtality	2.6 ± 0.7 1.4 ± 0.6	$1.2 \pm 0.2 \\ 0.4 \pm 0.2$	2.0 ± 0.3 0.6 ± 0.2		
1. Values are mites/trachea (N = 5 bees).								

TABL. 3. — Survival of Acarapis woodi infesting honey bees fed cymiazole in queen cage candy 1

Experiment 4. LC_{50} for A. woodi ; Cymiazole in queen cage candy.

Initially, only the highest concentration gave control, but after three weeks all concentrations showed activity (Table 3). The LC_{50} for cymiazole after 21 days was 0.15 mg/g (Table 2). Mite survival (all stages) ranged from 40 mites/ trachea in controls to 2.6 in bees given the 1.0 mg/g diet (Fig. 2 and Table 3). Candy consumption differed considerably. After one week, bees fed the 0, 0.75, 1.0, 1.25, 1.5, and 2.0 mg/g treatments consumed 98.2, 61.8, 61.8, 56.4, 52.7, and 41.8 % of the candy, respectively. This indicates that even the lowest concentration, 0.75 mg/g, reduced palatability. Bee mortality after 21 days on these same diets was 5.7, 0, 0, 2.8, 17.1, and 22.9 % respectively.

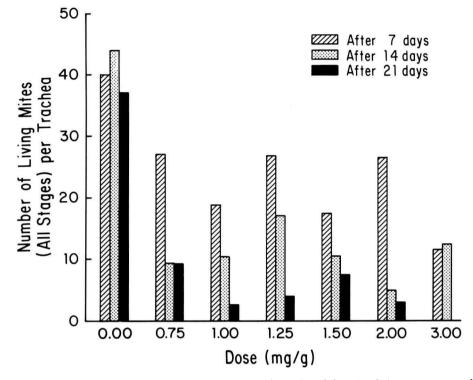


FIG. 2. — Survival of Acarapis woodi (all stages) infesting honey bees fed cymiazole in queen cage candy.

DISCUSSION

These laboratory tests have shown that, when dissolved in sucrose solutions, cymiazole can control *A. woodi* infesting worker honey bees. Total control was generally not achieved, but 80 % mortality was regularly observed. Mite response to cymiazole was comparatively slow, though this is not likely to be an important factor when treating colonies. The poor control of immature mites is not surprising as they probably feed less than adults (see MORISON, 1932, ÖRÖSI-PÁL, 1934). This has been observed previously with other effective adult acaricides (VECCHI and GIORDANI, 1968; EISCHEN *et al.*, 1987).

Why incomplete adult control was observed is not clear. Lowered palatability at high concentrations may have caused bees to limit the dose by consuming less medicated syrup. Perhaps low chronic doses are worth further examination. Alternatively, these observations were made on caged, queenless, and broodless bees. Consumption rates and physiologies might be quite different under more natural conditions. It is encouraging though, that cymiazole was able to effect acceptable control at levels that did not appear to harm the bees.

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RÉSUMÉ

LE CYMIAZOLE, UN ACARICIDE SYSTÉMIQUE CONTRE ACARAPIS WOODI (RENNIE), PARASITE DE L'ABEILLE . I. TESTS EN LABORATOIRE

Le cymaziole (hydrochlorure de 2-[2, 4-dimethylphenyl-imino]-3-methyl-4-thiazoline ; spécialité Apitol[®] de Ciba-Geigy Ltd. sous forme de granules dosés à 17,5 % de cymiazole) a été dissout dans un sirop de saccharose à 50 % (vol./vol.) et donné à des abeilles encagées, parasitées par *Acarapis woodi*. Les sirops contenaient 0,0, 0,025, 0,05, 0,11, 0,22, 0,44, 0,88 et 1,75 mg de cymiazole par ml. Les deux plus fortes concentrations ont tué environ 80 % des acariens adultes en une semaine. Au bout de 3 semaines on a observé des résultats plutôt bons dans tous les groupes traités, excepté chez ceux qui avaient reçu le sirop à 0,025 mg de cymiazole par ml. La CL₅₀ à 3 semaines a été de 0,03 mg/ml. Les abeilles nourries avec les sirops à 0,025-0,88 mg de cymiazole par ml ont eu un comportement d'amassement et une longévité à peu près semblables à ceux des témoins (sirop de sucre pur). Aux concentrations plus élevées, on a observé une consommation plus faible et une longévité réduite. Dans une expérience semblable avec du cymiazole mélangé à du candi dans une cage à reine, on a obtenu un niveau comparable d'efficacité. (CL = 0,15 mg/g).

ZUSAMMENFASSUNG

CYMIAZOL (APITOL[®]), EIN SYSTEMISCHES AKARAZID ZUR BEKÄMPFUNG VON ACARAPIS WOODI (RENNIE) BEI DER HONIGBIENE. I. LABORTESTS

Cymiazol (2 - [2,4 - Dimethylphenyl - iminol - 3 - methyl - 4 - thiazolin - hydrochloride, Apitol[®], 17.5 % Cymiazol Granulat, Ciba-Geigy Ltd.) wurde in 50 % igem Saccharosesirup (Vol./Vol.) gelöst und an gekäfigte Honigbienen, die mit *Acarapis woodi* (Rennie) infiziert waren, verfüttert. Es wurden Sirupe der Konzentrationen 0.0, 0.025, 0.05, 0.11, 0.22, 0.44, 0.88 und 1.75 mg/ml verglichen. Die beiden höchsten Konzentrationen töteten innerhalb einer Woche ca. 80 % der adulten Milben. Nach drei Wochen war die Bekämpfung in allen behandelten Gruppen hinreichend gut außer bei der Konzentration 0.025 mg/ml. Die LC₅₀ war nach drei Wochen 0.03 mg/ml. Das Futtereintragen und die Lebensdauer der Bienen, die mit Cymiazol der Konzentrationen 0.025-0.88 mg/ml gefüttert wurden, waren in etwa gleich mit dem von mit Saccharosesirup gefütterten Bienen. Bei höheren Konzentrationen wurde eine reduzierte Nahrungsaufnahme und verkürzte Lebensdauer der Bienen festgestellt. Ein ähnliches Experiment mit Cymiazol in Futterteig ergab einen vergleichbaren Wirkungsgrad in der Bekämpfung (LC₅₀ = 0.15 mg/g).

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