



HAL
open science

Integrated control of trypanosomosis

Geoffrey Gimonneau, Jean-Baptiste Rayaisse, Jérémy Bouyer

► **To cite this version:**

Geoffrey Gimonneau, Jean-Baptiste Rayaisse, Jérémy Bouyer. Integrated control of trypanosomosis. Pests and vector-borne diseases in the livestock industry, pp.147 - 174, 2018, 10.3920/978-90-8686-863-6_6 . cirad-01877598

HAL Id: cirad-01877598

<https://hal.science/cirad-01877598v1>

Submitted on 28 Sep 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

6. Integrated control of trypanosomosis

Geoffrey Gimonneau^{1,2*}, Jean-Baptiste Rayaisse² and Jérémy Bouyer^{3,4,5,6}

¹CIRAD, UMR INTERTRYP, Département BIOS TA A-17/G Campus International de Baillarguet 34398 Montpellier Cedex 5, France; ²Centre International de Recherche-développement sur l'Élevage en Zone Subhumide, BP 454, Bobo-Dioulasso, Burkina Faso; ³Unité Mixte de Recherche ASTRE 'Animal, Santé, Territoires, Risques et Écosystèmes', Campus international de Baillarguet, Centre de Coopération Internationale en Recherche Agronomique pour le Développement (CIRAD), 34398 Montpellier, France; ⁴National Institute for Tsetse and Trypanosomosis Control and Eradication, Livestock Development Sector, Ministry of Agriculture, Addis Ababa, Ethiopia; ⁵Pan-African Tsetse and Trypanosomosis Eradication Campaign coordination office, Rural Economy and Agriculture Department, African Union Commission, P.O. Box 3243, Addis Ababa, Ethiopia; ⁶ASTRE, University of Montpellier, CIRAD, INRA, Montpellier, France; geoffrey.gimonneau@cirad.fr

Abstract

In many parts of Africa, tsetse eradication is impossible due to political, environmental or economic circumstances. In these situations, African animal trypanosomosis control relies on communities or farmer-based control, implemented at a local scale in accordance to the eco-epidemiological context and the cattle rearing system to be sustainable. Management of the African animal trypanosomosis requires integrated controls strategies that combine the use of more than one locally-based tool and where possible, needs to be assisted by veterinarians and other animal health professionals. Several tsetse control methods based on insecticide treated cattle (i.e. pour-on, manual spraying, community bath) and insecticide treated target (traps and screens impregnated with insecticides) are available and should be complemented with diagnostic tests and medication (active trypanocides with prophylactic and/or therapeutic action). However, their adoption is mainly dependent on the engagement of communities, farmers and herders. Indeed, the adoption of a locally-adapted control strategy will depend on farmers socio-technical networks, the cost-effectiveness of the control activities, as well as the time and cost for implementation. In general, insecticide treated cattle methods are the most suitable and acceptable for farmers, because they protect a private good i.e. cattle, whereas insecticide treated targets are generally considered to provide a public good. Nonetheless, selection of the most appropriate tools requires consideration of local disease epidemiology (including host-parasite coevolution), local environmental and socio-economic constraints. The active involvement of communities, farmers and herders is essential from the beginning of the conception of innovative control strategies, and the cost of local integrated pest management should be reduced as much as possible, to be adopted as an acceptable and sustainable animal production cost.

Keywords: African animal trypanosomosis, cattle rearing system, epidemiological cycle, *Glossina*, integrated management, vector control

Introduction

Tsetse flies are the major vectors of African animal trypanosomosis (AAT), a disease of economic importance to the livestock production in Africa (Bouyer *et al.* 2015; Itard *et al.* 2003). AAT is considered among the greatest constraints to livestock production in sub-Saharan Africa and its economic cost has been estimated at USD 4.75 billion per year (Van den Bossche *et al.* 2010). To date, vector borne diseases of cattle are mainly controlled through prophylactic and curative

drugs in livestock. This approach is no longer sustainable, because of the increasing development of drug resistance (Geerts *et al.* 2001).

In 2000, the African Heads of State and Government decided to increase efforts to address the tsetse and trypanosomiasis problem on the African continent and created the Pan-African Tsetse and Trypanosomiasis Eradication Campaign (Kabayo 2002). This initiative aims to encourage tsetse eradication projects throughout Africa, based on the area-wide integrated pest management principles (AW-IPM), see Vreysen *et al.* (2007) and (2013) for more details. The eradication of tsetse populations is considered the most cost-effective option when successful and sustainable (Kgori *et al.* 2006; Vreysen *et al.* 2000). However, successful examples of sustainable tsetse eradication cover less than 2% of the total infested area in Africa (estimated around 10 million km²). Several conditions must be met to achieve this goal: strong political support, feasibility studies, preoperational agreement (governmental and departmental agreements, ethical committee) and planning, mass rearing facilities (if the sterile insect technique is included in the IPM strategy) and a comprehensive eradication campaign (Vreysen *et al.* 2007). For more details on eradication, see Chapter 14, 'Genetic control of vectors' (Bouyer and Marois 2018). According to an extensive study of the published successes and failures (Bouyer *et al.* 2010; Vreysen *et al.* 2013), a decision diagram for tsetse control, taking into account the cost-efficiency of each technique has been proposed (Bouyer *et al.* 2013).

In most of cases (cases IA and IB in Figure 1), tsetse control will have to be conducted by the beneficiaries themselves in a sustainable way, since tsetse eradication at a regional scale would not be feasible. In this context, it is only necessary to achieve a reduction in the relative density of tsetse flies below the transmission threshold (Bouyer *et al.* 2013). This local integrated pest management (L-IPM) would be considered as a 'production cost' and therefore it would be essential to minimize it as much as possible. Several tools will be needed to implement effective local integrated control strategies for trypanosomiasis disease management by either communities or farmers and, in close association with veterinary services. It should include tsetse control methods such as insecticide-treated cattle (ITC; e.g. pour-on, spray and dip) and insecticide-treated targets (ITT; e.g. traps and screens impregnated with insecticides) (Vreysen *et al.* 2013), but also diagnostic tests for trypanosomes and medications (prophylactic and/or therapeutic trypanocides). Before any intervention, a baseline data collection should be performed in order to select the most appropriate tools, according to the local disease epidemiology, host-parasite coevolution factors and environmental and socio-economic constraints. For tsetse-transmitted AAT, the objective is to use vector control as the primary method to reduce the incidence of the most important trypanosome species for cattle, namely *Trypanosoma congolense* Broden and *Trypanosoma vivax* Ziemann (Trypanosomatidae), and thus to prevent or minimize establishment and spreading of strains resistant to trypanocides within the cattle population. Also, the use of insecticides needs to be carefully managed to reduce costs, and the risk of insecticide resistance in tsetse (although this has not been yet observed) but also in other vectors that could be exposed such as ticks in insecticide-treated cattle (Eisler *et al.* 2003). Therefore, a sustainable AAT control system requires careful over-sight to both minimize costs and avoid resistance development in both the vector and the parasite whilst ensuring effective disease control.

Moreover, the success of AAT control will be strongly dependent on the communities, farmers and herders implication. Indeed, the adoption of the proposed control strategy will depend on their socio-technical networks, and the cost-effectiveness derived from the control activities, as well as the time and cost it will require (Bouyer *et al.* 2011a; Hargrove 2003; Kamuanga *et al.* 2001a,b). The perception of the benefits will depend on the establishment and understanding of a sustainable

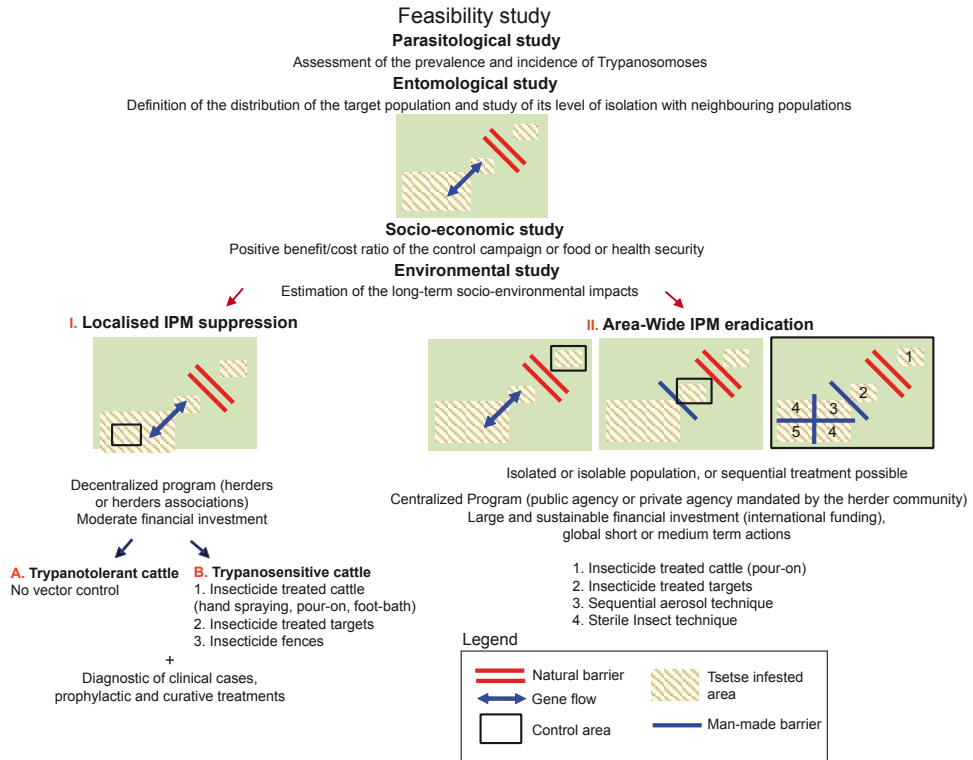


Figure 1. Decision diagram for tsetse control operation (Bouyer et al. 2013, with permission).

production plan (i.e. strategy including the management of health, nutritional and zootechnical constraints). In order to select the best L-IPM strategies in the different contexts, it is important to identify baseline indicators such as the eco-epidemiological setting, the rearing system and the cattle breed. According to these pieces of information, L-IPM implementation would be refined in association to the socio-technical networks involved (Bouyer *et al.* 2013).

This chapter will focus on the control of tsetse-transmitted African animal trypanosomosis (AAT) with a recommended framework for situations where farmer-based vector control is the only sustainable way of controlling the disease.

Vector control methods relevant to African animal trypanosomosis control in cattle

Insecticide treated cattle

This method is based on the treatment of cattle with insecticide formulations (mainly based on pyrethroids) using a wide range of techniques (pour-on, spraying, whole body dips / baths and sprays/ showers). The treated cattle then act as very attractive and lethal baits for tsetse and ticks due to their odour, movement and size.

Pour-on formulations

The main advantages of pour-on formulations over other techniques is that no equipment and particularly no water are needed to treat cattle. Application is quick and easy, because the product is deposited on the back line of the animal and diffuses over the whole body (Figure 2A). However, pour-on formulation is more expensive than emulsifiable concentrate formulation of insecticides, due to large dose of active material per kg of animal weight. It is also more suitable for small rather than large herds, as treatment requires a lot of time. Moreover, pour-on formulations have a more important potential impact on the environment than spraying of insecticides (Vale *et al.* 2004).

Manual spraying

Manual spraying of emulsifiable concentrate formulations of pyrethroids is much cheaper (based on the cost of treatment per animal) than pour-on (lower amount of active ingredient required and emulsifiable concentrate cheaper than oil formulations), and presents a similar persistency. Insecticide solution is sprayed over the whole body of cattle (Figure 2B). The main hindrance of pour-on and manual spraying techniques is the treatment time (~5 min per animal) because the animal must be immobilized for treatment. For large herds (>100), the treatment time becomes



Figure 2. Insecticide treated cattle application methods. (A) Treatment of a zebu bull using a flumethrin pour on in Burkina Faso (photo by J. Bouyer). (B) Treatment of a zebu bull using a hand sprayer containing 0.005% alphacypermethrin in Burkina Faso (photo by J. Bouyer). (C) Treatment of zebu in community bath in South Africa (photo by J. Ntshangase). (D) Treatment of a zebu using a footbath containing 0.005% alphacypermethrin in Burkina Faso (photo by J. Bouyer).

very long. It can be reduced by handling the animals in a 'vaccination corridor' or cattle 'race' (with or without mechanical spray jet races) and/or by the simultaneous administration of other prophylactic or therapeutic disease controls. Moreover to be effective, the pour-on or manual spraying must be repeated every 2 weeks or monthly depending on the product used and tsetse species and density (Bouyer *et al.* 2007, 2008; Torr *et al.* 2007).

Community bath

Community baths (or dip tanks) are routinely used to treat cattle against ticks (vectors of East Coast fever) in some places like the Kwazulu-Natal region of South Africa. Cattle cross a community bath (generally more than 1.5 m deep) filled with an emulsifiable insecticide solution, resulting in the whole body impregnation in a short time (Figure 2C). The drawback of this method is that the dip tanks must be emptied when insecticides must be changed, with associated environmental hazards and high costs. Dip tanks are also used by individual owners in large and modern farms.

However, in general, dip tanks are very expensive to build and to use and are not appropriate for poor farmers. In addition, inappropriate management of residual insecticides can lead to hazards for individual and health risks for farmers.

Restricted spraying of insecticides

Restricting spraying reduces the quantity and cost of insecticides used and improves administration to only the lower parts of the cattle body (belly or lower abdomen, and legs) or by application using a footbath (Stachurski and Lancelot 2006), which is a cheap and fast way to treat cattle (Figure 2D).

This restricted treatment has the additional benefit of controlling the tick *Amblyomma variegatum* Fabricius (Acari: Ixodidae) for which it was first designed (Stachurski 2006; Stachurski and Lancelot 2006) as well as tsetse flies. Its principle is based on behavioural and ecological studies on *A. variegatum*, whose invasion process includes a temporary fixation in the inter-digital areas before they can reach their preferred sites on the body (Stachurski 2006). Similarly, this vector control method is very efficient against tsetse because their feeding behaviour mainly focuses on legs and belly of cattle (Torr and Hargrove 1998; Torr *et al.* 2007; Vale *et al.* 1999). Moreover, restricted application of insecticides reduces the environmental impact of ITC, particularly for dung fauna (Vale *et al.* 2004).

A very important parameter of restricting spraying is that 60 cattle can be treated within 8 min with a footbath versus 120 min for a full, hand-operated spray. For farmers, time savings are often more important than direct costs of these treatments, especially during the rainy season when they are very busy in cultivating their fields. The most important drawback of footbaths is that they are fixed installations and thus not appropriate for transhumant herds. Also, they require strong technical skills to correctly dose the insecticide and require community management (insecticide control, individual contribution) for sharing the use of the footbath that can be burdensome and problematic in some situations. A summary of ITC advantages and disadvantages is presented in Table 1.

Table 1. Summary of insecticide-treated cattle (ITC) advantages and disadvantages.

	Advantages	Disadvantages
ITT overall	cheap private good protection against ticks cost sharing of public facilities	requires the treatment of a large proportion of cattle to reduce tsetse densities no wild fauna partial protection (need trypanocides treatment)
Pour-on	no equipment, no water quick application	cost environmental impact
Manual spraying	cheap	treatment time need animal immobilisation low persistency (2-3 weeks) environmental impact
Community baths	treatment time	expensive to built management environmental impact
Restricted spraying	cheap reduced environmental impact	need animal immobilisation repeated weekly treatment time
Footbath	cheap very quick treatment time reduced environmental impact	low persistency (repeated application up to 10 times/month) fixed installation (not for pastoralists) community management constraints

Insecticide treated targets and traps

Tsetse traps and screens (Figure 3) impregnated with insecticides (known as insecticide treated targets), particularly pyrethroids, have been used as control methods since the 1970s, due to their user friendliness, low cost and efficacy (Cuisance *et al.* 1991; Laveissière *et al.* 1980). They can quickly decrease tsetse densities by up to 99% and therefore interrupt parasite transmission. Tsetse trapping has the additional advantage of being a good surveillance tool. Many models of traps and screens are available, and they must be selected specifically for the target species in order to function efficiently (Table 2; Figure 3).

Screens are simple devices (Figure 3B), originally constituted of 1 square meter of fabric impregnated with pyrethroids. They present the advantage to be environmentally friendly since insecticides are not dispersed in the environment, which limits their impact on non-targeted fauna, although a temporary impact on insectivorous birds densities and the disturbance of wild mammals have been observed during area-wide campaigns (De Garine-Wichatitsky *et al.* 2001). Their efficacy and persistence depends on the active ingredient, its formulation, and its concentration (200 to 400 mg/m² of cloth), and also on the nature of the cloth (density of fibres, presence of an anti-UV protective agent, type of fabric, thickness), and can last up to one year in certain conditions.

Table 2. Suitability of different trap models depending on the tsetse species targeted, based on trap efficiency (NA = not available, + low trapping rate for this species, ++ medium trapping rate, +++ good trapping rate) (Bouyer et al. 2005; Kappmeier 2000; Vale and Torr 2004).

Trap model	Target species									
	Tsetse			Savannah species			Forest species		Mechanical vectors	
	<i>Glossina palpalis</i>	<i>Glossina tachinoides</i>	<i>Glossina fuscipes</i>	<i>Glossina longipalpis</i>	<i>Glossina morsitans</i>	<i>Glossina pallidipes</i>	<i>Glossina austeni</i>	<i>Glossina brevipalpis</i>	Tabanids	Stomoxes
Biconical	+++	+++	+++	++	++	+	NA	+	+	++
Vavoua	+++	+++	+++	+++	++	+	NA	+	++	+++
Pyramidal	++	++	NA	+	+	NA	NA	NA	++	+
Nzi	+++	++	NA	++	++	NA	NA	NA	+++	+++
Tetra big	+++	+++	NA	NA	++	NA	NA	NA	+++	++
Tetra small	+++	+++	NA	NA	++	NA	NA	NA	+++	+++
X sticky	NA	NA	NA	NA	NA	NA	+++	+++	NA	NA
H3	NA	NA	NA	NA	NA	NA	+++	+++	NA	NA
F3	NA	NA	NA	NA	++	+++	NA	NA	NA	NA
Epsilon	NA	NA	NA	NA	++	+++	NA	NA	NA	NA



Figure 3. (A) Biconical trap set to monitor *Glossina palpalis gambiensis* Vanderplank (Diptera, Glossinidae) densities in Senegal (photo by G. Gimonneau). (B) Screen impregnated with deltamethrin, used to control *G. p. gambiensis* in Senegal (photo A.G. Mbaye).

Traps attractiveness can be boosted by olfactive attractants (octenol, acetone, metacresol, cow urine), especially against savanna species (Vale 1980), but results for riverine species are not so evident (Rayaisse *et al.* 2010). The use of olfactive attractant requires technical capacities and therefore is more adapted to AW-IPM programs than to farmer based control. Moreover, the cost effectiveness of attractants still needs to be proved.

The density of targets (trap or screen) placements to obtain the desired effect is context dependent and depends on the target tsetse species, and particularly the density of the vegetation. It is generally necessary to use higher densities of screens against riverine species in forest environments (density of 30/km of river or more) than against savanna species in open environments (1-5/km²).

The main drawbacks are the vulnerability of target material to fire, theft (especially for traps using metal frame) and floods. The insecticide persistence time may also be an issue, if it is too short and requires regular target replacements with associated cost. Moreover, there is a limited adoption of targets by farmers or communities due to a lack of sensitization on the use of these tools and also their availability on the market. A summary of ITT advantages and disadvantages is presented in Table 3.

Innovations and gaps

Tiny targets

The design of the target can have substantial effect on the cost-effectiveness in a vector control campaign and the optimal design is always dependent on the species targeted. Recently, smaller screens have been developed, that could lead to important implications for the costs of tsetse control programs (Esterhuizen *et al.* 2011). It was recently highlighted experimentally that reducing the target size to 1/16th of normal 1×1 m size (i.e. 25×25 cm), reduce catches of *Glossina fuscipes fuscipes* Newstead (Diptera, Glossinidae) only by half in average, suggesting a better cost-effectiveness (Lindh *et al.* 2009). Similar results were obtained for *Glossina palpalis gambiensis* Vanderplank and *Glossina tachinoides* Westwood reducing the size of the current 1×1 m black-blue-black target to horizontal design of around 50 cm and replacing black clothes by netting

Table 3. Summary of insecticide-treated targets (ITT) advantages and disadvantages.

	Advantages	Disadvantages
ITT overall	simple, easy to set up and efficient low environmental impact cheap	public good community management cost of deployment exposed to vandalism, theft could induce behavioural resistance
Screens	cheap easy and fast to set up	
Traps	strong sociological impact (visualisation of flies in the cage)	theft of iron pickets (not for all traps)
Small targets	cheaper	increased density low visibility in dense vegetation

will improve the cost effectiveness six-fold for *G. p. gambiensis* and *G. tachinoides* (Rayaisse *et al.* 2011). Table 4 gives the relative indexes (proportion of catches relative to black-blue-black 1×1 m standard targets) of small targets for *G. p. gambiensis* for different models and size.

Although there is evidence that reducing the target size will reduce its cost, it is not so clear if in any situation, this tool will be more cost-effective than other methods. Since the index for smaller targets is below 1, their use requires an increase in their density by 1/index (Table 3). Therefore, their cost effectiveness must be carefully assessed before implementation in control programs

Table 4. Detransformed daily mean catches (transformed means in bracket) of male and female *Glossina palpalis gambiensis* in Folonzo, Burkina Faso (Rayaisse *et al.* 2011).^{1,2}

Size (m×m)	Material	Shape	Rep	Male	Index	Female	Index	Total	Index
0.5×0.75	NBIN	V	14	3.0 (0.60)	0.57	3.3 (0.63)	0.46	6.5 (0.88)	0.50
0.75×0.5	NBIN	H	14	4.3 (0.72)	0.82	5.6 (0.82)	0.78	10.3 (1.05)	0.79
0.25×0.5	BkBIN	V	14	0.6 (0.21)	0.12	0.9 (0.27)	0.12	1.3 (0.37)***	0.10
0.5×0.25	BkBIN	H	14	2.8 (0.58)	0.54	1.7 (0.43)	0.24	4.5 (0.74)	0.35
0.25×0.5	NBIN	V	14	0.3 (0.10)***	0.05	0.5 (0.16)***	0.06	0.7 (0.24)	0.06
0.5×0.25	NBIN	H	14	2.2 (0.51)	0.42	1.4 (0.37)	0.19	3.5 (0.65)	0.27
			sed	0.085		0.084		0.090	
0.25×0.5	BBkBlBk	V	24	0.9 (0.29)***	0.32	0.8 (0.25)***	0.25	1.8 (0.44)***	0.28
0.25×0.25	BkBlBk	V	24	0.3 (0.12)***	0.11	0.2 (0.08)***	0.07	0.5 (0.18)***	0.08
1×1	NBIN	V	24	3.2 (0.62)	1.09	4.6 (0.75)	1.47	7.9 (0.95)	1.25
0.25×0.5	NBIN	V	24	0.7 (0.24)***	0.26	0.6 (0.19)***	0.18	1.4 (0.37)***	0.21
0.25×0.25	NBIN	V	24	0.1 (0.05)***	0.04	0.1 (0.05)***	0.04	0.2 (0.09)***	0.04
			sed	0.068		0.066		0.075	

¹ Catches followed by *** differ from the control at 0.001 level. Catch index is the mean catch of a target expressed as a proportion of that of the standard, which is 1×1 BkBlBk.

² Bk = black; Bl = blue; H = horizontal; ; N = net; Rep. = replicates; sed = standard error deviation; V = vertical.

according to the environment and species targeted. For *G. fuscipes fuscipes*, small targets seems more interesting than classical one, since small targets have indexes of almost 1 (Lindh *et al.* 2009). A recent study evaluated the cost using small target against this species in Uganda and there was reduces by 48% from USD 179 to USD 85.4 per km² (Shaw *et al.* 2015). For some tsetse species such as flies of the *morsitans* group (Diptera, Glossinidae), small targets have been proven to be inefficient (Torr *et al.* 2011).

Although small target is an important innovation, the knowledge gap here, is a lack of a fine tuned cost-benefit study of small targets to determine if this technology can reduce the cost of ITT in different contexts. Also, when applied by the beneficiaries themselves, to what extent this might impact on the will of communities to take responsibility over targets.

Insecticide treated fences

Insecticide treated fences are very efficient in areas where dairy cows are confined in zero-grazing units (food and water supplied to them). The use of mosquito netting impregnated with a pyrethroid (Figure 4) can reduce significantly the incidence of trypanosomosis (Bauer *et al.* 2006).

This technique was first designed for tsetse control and the principle is to surround zero-grazing units by insecticide-treated mosquito netting at a height of 150 cm. Indeed, behavioural studies have highlighted that tsetse usually attack their hosts at a height of less than 100 cm above ground level to feed on the legs (Bauer *et al.* 2006). In some situations, insecticide treated fences can also have a significant effect on vector of human diseases. In Ghana, impregnated mosquito nets were used to protect cattle pens but also reduced significantly mosquito malaria vectors (*Anopheles gambiae* Giles and *Anopheles ziemanni* Grunberg; Diptera: Culicidae) (Maia *et al.* 2012). In Ghana, treated fences have been proved to be very efficient to protect pigs pen against tsetse (Bauer *et al.* 2011). It was also successfully used in Guinea in older non active human African trypanosomosis (HAT) foci to reduce vector densities and prevent HAT transmission (Kagbadouno *et al.* 2011). In some conditions, it can also offer a good protection against biting flies, *Stomoxys* spp. (Diptera: Muscidae) (Maia *et al.* 2010), which favours its adoption since animals were less disturbed by flies.



Figure 4. Protection of cattle with impregnated mosquito net in a modern farm in Senegal (photo by J. Bouyer).

However, in La Reunion Island, it failed to reduce the density of *Stomoxys* spp., probably because the cattle pens were not surrounded completely with mosquito netting (Bouyer *et al.* 2011b).

The residual gap for this technology is that most of the trials were organized by comparing fully small protected and small control pens, whereas in some farms the reality is quite different with a calculated total length of fence of 1.2 km. Therefore, according to the fence price (i.e. € 160 for a roll of 100 m length × 1 m height in Senegal), it would require € 1,920 to surround all the cattle pens, whereas in the same way, 5 impregnated targets would probably have similar impact on tsetse density. Further research is thus required to define the best areas to protect with either insecticidal fences or screens which will be between the tsetse resting sites in the vegetation and the cattle, in order to reduce the cost of fencing needed to protect a farm.

Combination of repellent and insecticides

Recently, several combinations of repellents and insecticides have been developed in order to improve the effect of formulations in the increasing context of insecticide resistance, especially against mosquitoes (Faulde and Nehring 2012). Today, although no insecticide resistance has been identified in tsetse, combination of different insecticides and molecules can significantly increase the formulation effect (Gimonneau *et al.* 2016). Application of repellents on cattle to reduce trypanosomosis incidence was tested, and it was concluded that the repellent technology was not sufficiently efficient under natural tsetse challenge to merit commercial development (Bett *et al.* 2010). Moreover, we think that pushing tsetse towards other animals or herds is not a good principle for tsetse control, even if the repellent technology is efficient. This is because the protection of an individual animal or herd should not increase the risk for other animals at a community level, especially since the poorer farmers in that community might not be able to afford the repellent product. Another option is to associate repellents with insecticides. A new pour-on containing two insecticides, a pyrethroid synergist and a repellent have recently been developed and compared to a standard pour-on (i.e. based on one pyrethroid) (Gimonneau *et al.* 2016). Results showed in the laboratory that the repellent and insecticides formulation was significantly more efficient than the classical formulation with a longer knockdown effect (37 and 28 days, respectively; Figure 5A). More importantly, it significantly increased the cattle protection against tsetse bites (Figure 5B). In the field, this combination has been proved to be very effective against ticks with a complete elimination three days after application and in the same time a significant reduction in trypanosomosis prevalence and increased in packed-cell volume (PCV).

This new pour-on formulation, combining insecticides and repellent, was highly effective against AAT and ticks, with a longer persistence than other pour-on products on the market. It offered immediate effect on ticks and low treatment frequency to maintain a low ticks infestation and trypanosomosis prevalence. Moreover, this new insecticide formulation represented the first one to provide a partial individual protection against tsetse bites and AAT (Hargrove *et al.* 2000). Although the efficacy will probably not be the same in various contexts and environments (Hargrove *et al.* 2003), it was therefore a great innovation for farmers for the control of ticks and AAT. Actually, providing individual protection of cattle against AAT and ticks might increase the adoption of the control technique by farmers, as opposed to collective protection (Bouyer *et al.* 2011a). In sites where AAT coexists with HAT, this tool might represent another weapon against tsetse within a one-health perspective, as it has been suggested earlier that treating cattle could help controlling HAT through a reduction of tsetse densities (Ndeledje *et al.* 2013).

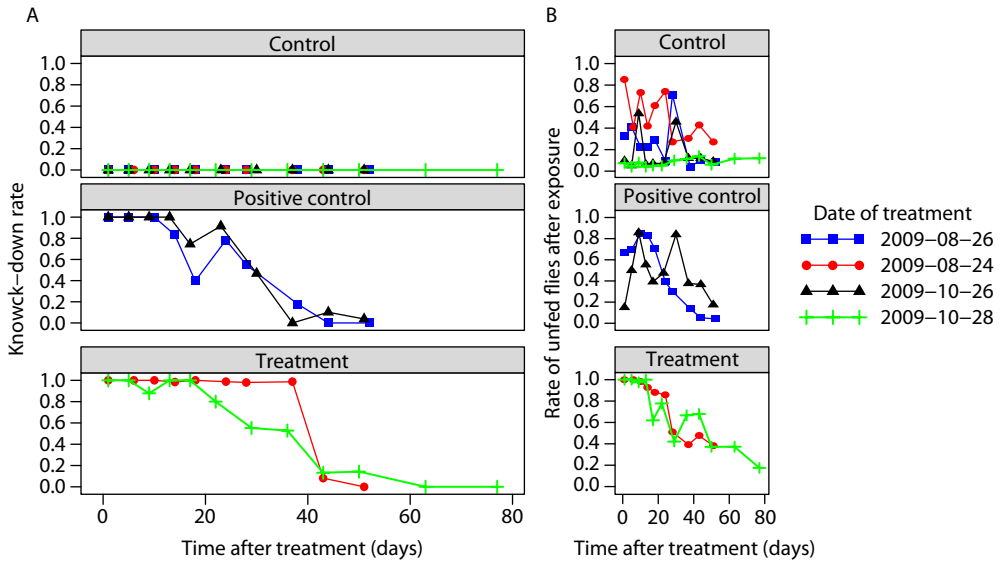


Figure 5. (A) Knock-down rate of *Glossina palpalis gambiensis* in function of the time after treatment. (B) Rate of unfed *G. palpalis gambiensis*, giving an estimation of the protective effect of treatment, in function of the time after treatment.

One of the main constraints for farmers could be the price but a formulation for water suspension has been developed, decreasing the total amount of insecticide used and the treatment price. However, its impact regarding knock-down rate and disease or biting protection will have to be tested. Another knowledge gap is that no data is available on the toxicity, residuals and ecotoxicity of this combination.

Restricted application of insecticides

Socio-economic studies on restricted application of insecticides to cattle have revealed that treatment time and usage by the community were the most important constraints for the adoption of partial spraying and footbaths respectively (Bouyer *et al.* 2011a; Selby 2010). Socio-economic studies identifying the types of setting in which corridors or footbaths (and the maximal distances between farms and communal corridors or footbaths) are acceptable for sustainable use of the technique are needed to further improve the adoption of these techniques. It would also be important to develop with the stakeholders, acceptable methods for sharing the costs between farmers. Actually, there is no recommendation on how to use this technique taking into consideration the herder constraints and their habits, particularly in the case of traditional farmers.

Moreover, regarding footbaths, the dosage of the insecticide is a tricky issue for traditional farmers, when it is done using an abacus. A simpler system would be needed to reduce the technical problems associated with this dosage of insecticides.

Finally, because restricted application focused to the lower parts of the body, it would be very useful to quantify the insecticide residuals in milk.

Quality control and evaluation procedure for targets

There is a real need to develop a standard quality control procedure and evaluation for targets in order to improve the reliability, efficacy and persistency of the different batches of targets sent by a given manufacturer.

According to the literature, two tools have been developed to evaluate targets but with several drawbacks. The first method consists in the restriction of a fly in the head of a glass tube by a loose-fitting piston carrying a sample of insecticide (Kernaghan and Johnston 1962). The fly is handled only when first introduced into the tube; it then remains in the same tube during the holding period. This method is time-consuming because the test is performed on one fly at a time and several flies are needed to obtain robust statistics (i.e. minimum of 30 flies). Moreover, injuries could be inflicted to the flies, generating biases in the results. Also, the fly stays in the same tube during the observation period which represents a bias since insecticides could remain in the tube. The second apparatus is a 'T' shape flight tunnel where flies are attracted to the light where the netting is set (Torr 1985). The fly collides with the netting, set obliquely across the flight-path, and it then flies towards a second, stronger, light source. In this system, there is no possibility to control the time of exposure of the fly that is a very important criterion for insecticide bioassays. Moreover, the apparatus configuration allows some flies to reach the second light source without entering into contact with the net (Torr 1985). Anyway, these two tools seem not to be used by the researchers community as there is no mention of these in the literature, may be due to a lack of reproducibility.

In target manufactures, quality control procedures seem to be absent. Huge differences in persistency are observed in the field between batches that impact the success of vector control. A solution would be that the manufacturers should use exactly the same production process for a given trap or screen model, or that they should be able to test these different properties for each change in the production process and provide data, such as trapping indexes relative to a reference trap and persistency in the field.

Impact of farming systems and tick-borne diseases on the adoption of tsetse control methods

Even if socio-economic surveys have already been conducted to study the adoption of tsetse control techniques by farmers, it would be important to study accurately the perception and adoption rate of the different techniques available to control tsetse in different farming systems. Actually, farmers belonging to different farming systems (traditional/modern, sedentary/transhumant) have different perceptions of their cattle (production tool vs money savings) and socio-economic networks (Bouyer *et al.* 2015). Moreover, the risk level in case of an innovation is not the same (much higher for traditional farmers) and will probably impact the adoption process (Bouyer *et al.* 2011a).

It appears also important to study the impact of ticks and tick-borne diseases on the adoption of tsetse control techniques. Because ITC based on pyrethroids directly impact ticks and tick-borne diseases, the adoption process is facilitated. This aspect should not be overlooked because it can lead to situations where the farmers prefer to keep their usual control technique even if it is working only against ticks than to adopt a new one that is working both against tsetse and ticks

(see below the example of ITC in Uganda). It is thus very important to sensitize the farmers on the likely impact on ticks when a new strategy to control tsetse is proposed. These should be studied in each African region (at least western and eastern), since important local specificities might be observed.

AAT control strategies including vector control with preventive and/or curative treatment of cattle

Description of the different epidemiological settings

In this part, we consider only AAT or Nagana, caused by *T. congolense*, *T. vivax* and *Trypanosoma brucei brucei* (Plimmer & Bradford) transmitted to cattle by tsetse, and mechanically transmitted for *T. vivax* beyond the tsetse belt. Mechanically transmitted Animal trypanosomosis or Surra caused by *Trypanosoma evansi* (Steel) worldwide and by *T. vivax* mainly in South America is not considered. For Nagana, and with a special focus on *T. congolense* and *T. vivax*, three major eco-epidemiological cycles related to cyclical transmission by tsetse have been described, and one mechanical cycle occurring at the limit of the tsetse distribution area (Pagabeleguem *et al.* 2012; Van den Bossche *et al.* 2010).

Sylvatic trypanosomosis

In sylvatic trypanosomosis settings, trypanosomes are transmitted by tsetse to the trypanotolerant wild fauna. Cattle are supposed to be absent in these areas but sometimes enter this system for illicit grazing purposes (and/or transhumance) and are then infected with highly virulent strains.

Interface trypanosomosis

Interface trypanosomoses occur at the edge or interface of agro-pastoral areas and protected areas. Tsetse diversity and abundance is high at this interface (border effect), and virulent strains are transmitted from wild fauna to cattle, leading to acute infections in the latter, with a high mortality rate, or even epidemic situations.

Endemic trypanosomosis

In endemic trypanosomosis, no or very scarce wild fauna is involved and cattle are the main host for tsetse. Tsetse density, lifespan and diversity tends to reduce (Van den Bossche *et al.* 2010). The virulence of trypanosome strains is reduced by their circulation within the cattle compartment, although virulent strains can be imported from the former areas by transhumant herds. In this setting, cattle are used to 'live with the disease'.

Mechanical trypanosomosis

Mechanical trypanosomosis occurs at the limit of tsetse distribution, within the range of movement of transhumant herds. In West Africa, trypanosomes are imported by cattle transhuming into the tsetse belt and locally transmitted to resident herds by mechanical vectors (*T. vivax* only). In East Africa, *T. vivax* is well established in sedentary cattle whereas tsetse flies are absent, but numerous movements of cattle occur and might also allow the importation of trypanosomes (Ahmed *et al.* 2016). The relationship between mechanical trypanosomosis and cyclical trypanosomosis is not so clear and need further investigation (Tadesse *et al.* 2011).

Description of the different cattle rearing systems

In this section, 3 types of cattle rearing systems will be considered as important to take into consideration when selecting the best control options:

1. Transhumant rearing systems, where herders drive their cattle to remote grazing areas (up to several hundred kilometres), especially during the dry season. Animals are mainly local breeds, more or less trypanotolerant depending on AAT pressure. Inputs to improve productivity are very low. Animals may encounter tsetse eco-epidemiological contexts that are different from those in their area of origin. Therefore, they will probably move between tsetse free and infested area with different eco-epidemiological cycles, linking them and importing trypanosome strains from one to the other. Controlling this genetic flow is a very important challenge (see below).

The two other rearing systems can be considered as sedentary systems, which are generally exposed to one epidemiological cycle only:

2. Agro-pastoral system/mixed farming systems, where farmers have two main activities: agriculture and cattle rearing. Crop residues are generally used to rear cattle in association with the surrounding environment, i.e. natural pastures within a range of 5 to 10 km. Cattle are mainly local trypanotolerant breeds or more or less crossbred with trypanosusceptible breeds depending on AAT pressure. Inputs are low to medium. Different situations could be observed between farmers according to the time and efforts they spend in different activities, i.e. cropping or cattle rearing but they will be considered here together because the proposed AAT strategies will be the same.
3. Zero grazing units are modern farms, generally in peri-urban areas where animals are mainly fed with forage or agro industrial subproducts. Farmers rear improved local breeds (zebu), sometimes cross-breed with exotic cattle or even pure exotic cattle. Animals are considered as a production tool and generally have high economic value in comparison to previous systems. Inputs are generally high in order to maintain these breed under high parasitological pressure in combination to high productive pressure.

Proposal of a general framework for farmer-based AAT Control

Proposals presented in this section are mainly based on argument present in this chapter and other peer reviewed documents (Bouyer *et al.* 2010, 2013; Vale and Torr 2004; Van den Bossche and Delespaux 2011; Vreysen *et al.* 2013).

A general decision framework depending on the type of farming system and epidemiological cycles is presented in Figure 6 from Bouyer *et al.* 2013. The integrated packages are developed below.

Sylvatic AAT (AI and BI)

This epidemiological cycle is encountered in protected areas only, where cattle rearing should be discouraged. In this situation, ecotourism, ranching of wild fauna or game and hunting reserves should be encouraged instead.

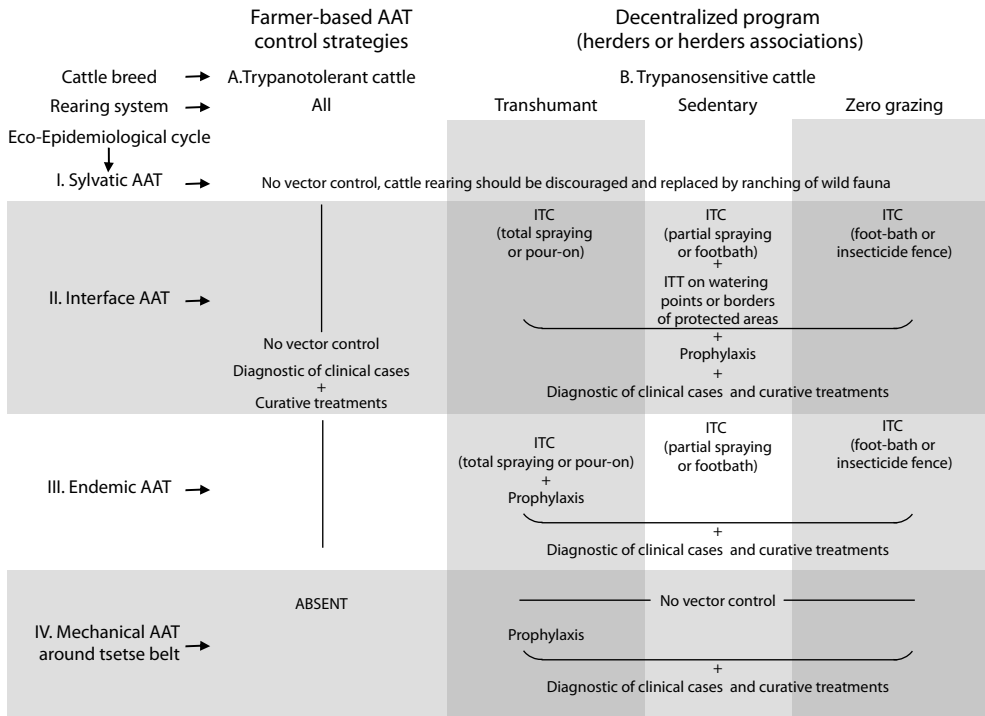


Figure 6. Decision framework proposing different strategies integrating vector control and the use of trypanocides, depending on the type of farming system (columns) and epidemiological cycles (rows) (ITC = insecticide treated cattle; ITT = insecticide treated targets; AAT = animal African trypanosomosis) (Bouyer et al. 2013, with permission).

Trypanotolerant cattle (All and AIII)

In systems involving trypanotolerant cattle, vector control is not cost-effective. Trypanotolerance is defined here as the ability to limit parasitaemia and anaemia and remain productive in enzootic areas (Dayo et al. 2012). Therefore, clinical cases are rare and generally caused by a loss of immunity (other disease or insufficient food). These clinical cases should be diagnosed and treated using trypanocide drug treatments but prophylactic treatments should be avoided in order to reduce the development of drug-resistant strains of trypanosomes. Moreover, it might prevent the establishment of a protective immunity in young cattle that must be infected at an early age to ensure maintenance of tolerance. The relationship between trypanosomes and trypanotolerant cattle is more or less at equilibrium, and interventions should in general be limited to avoid breaking this equilibrium. This strategy has the additional advantage to allow the long term conservation of trypanotolerant breeds.

\$ [protocol]://www.wageningenacademic.com/doi/pdf/10.3920/978-90-8686-863-6_6 - Wednesday, September 19, 2018 5:04:34 AM - IP Address: 197.239.81.1

*Trypanosensitive cattle (BII to BIV)***Interface AAT (BII)**

Interface AAT is probably the case where the economic impact of AAT on cattle rearing systems is the highest, because virulent strains are transmitted from wild fauna to trypanosensitive cattle on a regular basis. Here, the use of different vector control strategies and the use of trypanocides (prophylactic and curative trypanocides) are necessary and cost-effective, in order to reduce the incidence of AAT, the morbidity and mortality of cattle, and the development of resistant strains of trypanosomes.

ITC is the method of choice because it protects an individual good instead of a public one and it is also the most cost-effective (Shaw *et al.* 2013). The type of ITC must be differentiated depending on the rearing systems:

For transhumant herds, fixed structures like footbaths are not suitable, and water is difficult to transport. Therefore, it should be recommended to reduce the treatment frequency as much as possible by the use of pour-on or total spraying (if water is available) of the cattle.

For sedentary herds, fixed structures are convenient. Restricted application of insecticides using partial spraying (every two weeks) or footbaths (weekly) should be recommended: the treatment corridor or the footbath respectively can be easily built at the exit of the cattle' pen, with a door allowing to pass cattle through it when desired, in order to reduce treatment time as much as possible.

For zero-grazing herds, insecticide fences can be used around the cattle pens, or footbaths placed between the pen and the grazing area in order to facilitate treatment.

However, ITC is generally not sufficient to protect cattle totally and it should be completed by ITT especially at watering points when riverine tsetse species are involved. The use of impregnated targets or traps set every 100 m along 1 km of the river on each side of the water points will then be enough to suppress the tsetse population at this site (Knols *et al.* 1993; Willemse 1991). Based on several tsetse control programs, target barriers will never be maintained by the farmers themselves; therefore authorities managing game or hunting reserves should be sensitized to use part of the benefits generated by tourism to maintain these barriers.

Endemic AAT (BIII)

In this system, the same ITC strategy as described for interface AAT (BII) should be recommended but ITT is no more required. Indeed, the strains circulating here are less pathogenic. Moreover, due to environmental degradation, tsetse distribution is much more fragmented and densities are generally low. Because the tsetse feed mainly on domestic animals due to the absence of wild fauna, ITC is likely to have a more important impact on tsetse.

In endemic AAT, the relationship between trypanosomes and trypanotolerant cattle is more or less at equilibrium, so prophylactic treatments should be limited. However, importation of virulent strains from the sylvatic and interface cycles is possible because of transhumant herds. Clinical cases should be quickly diagnosed and treated immediately using curative trypanocides.

Mechanical AAT (BIV)

In mechanical AAT, tsetse are absent and vector control approaches are generally not effective. In this system, AAT occurs as epidemics so prophylactic treatments may not be cost-effective, except in the case of transhumant herds. If animals come from a tsetse free area, it is recommended to use prophylactic drugs and curative ones just before their return.

This regimen is intended to avoid losing animals during transhumance through areas with other epidemiological cycles, and to avoid importing trypanosomes into this tsetse-free area and thus prevent the establishment of mechanical transmission.

Case studies of vector control approaches

Restricted application of insecticides using footbaths around Bobo Dioulasso, Burkina Faso

This case study is based on three peer reviewed articles (Bouyer *et al.* 2007, 2009, 2011a). The epidemiological cycle is the endemic AAT cycle and the type of cattle rearing system is an agro-pastoral rearing system (situation BIII).

In Dafinso (Burkina Faso, 15 km north of Bobo-Dioulasso), 68 cattle of a herd of 96 cattle (i.e. 71%) were treated with a footbath containing a pyrethroid formulation. The effect of this restricted insecticide treatment of cattle was observed on released cohorts of reared, irradiated and marked tsetse flies (Figure 7A).

Footbath treatment significantly reduced the mean lifespan of released flies (*G. p. gambiensis* and *G. tachinoides*) from 4.7 days (95% CI 3.4-7.5) to 1.7 days (95% CI 1.3-2.4). The apparent densities of wild flies at the water point frequented by the footbath-treated cattle herds reduced quickly in Dafinso (Bouyer *et al.* 2007), with an estimated daily mortality rate related to this exposure of 0.39 (95%CI 0.19-0.54), which was much higher than needed to reduce a wild tsetse population (0.03) (Hargrove 2003).

An one-year survey (May 2005-June 2006) was conducted in the same site in order to compare the incidence of AAT in a footbath-treated cattle herd (48 footbath-treated animals with a suspension of pyrethroids administered every 2 days by farmer itself with a control one (69 cattle not treated), watered at another river section in the same area (Bouyer *et al.* 2009). From farmers' viewpoint, the main justification of the footbath treatment was to prevent infestation by *A. variegatum* so, they stopped using the footbath when tick infestation decreased, after the end of the rainy season, i.e. the 7 October 2005. This scenario was monitored and data used to assess whether tsetse-control measure implemented during the rainy season only were sufficient to limit tsetse populations and trypanosome transmission during the dry season, when riparian flies are usually still active and numerous enough to have a great epidemiological importance. During the survey, cattle were sampled for identification of the three trypanosome species occurring in the area using PCR (*T. vivax*, *T. brucei sensu lato* and *T. congolense* savannah type). The study can thus be considered as a comparison of an integrated AAT control strategy combining vector control and curative treatment of clinical cases with the control group only receiving curative treatment of clinical cases but no vector control.

The initial prevalence of trypanosomosis was 6.6% (n=91), i.e. 0.0% for *T. brucei*, 4.4% for *T. congolense* and 2.2% for *T. vivax* (no difference between the two herds, Fisher's exact test, $P=0.69$).

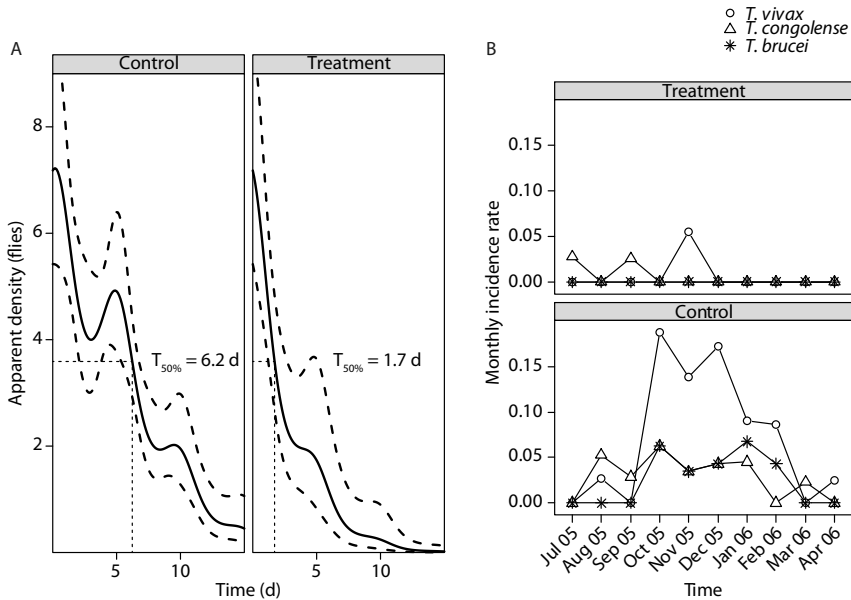


Figure 7. (A) Effect of the exposition to footbath-treated cattle on the time corresponding to a 50% reduction ($T_{50\%}$) of the initial apparent density of released, irradiated tsetse flies in Dafinso (Burkina Faso) during the early rainy season 2005 (points: observed data; the solid line was estimated from a negative-binomial model fitted to the points; dashed lines: 95% confidence limits) (Bouyer et al. 2007, with permission). (B) Observed incidence rate of cattle trypanosomosis in Dafinso (Burkina Faso) according to *Trypanosoma* species and treatment category (Bouyer et al. 2009, with permission).

The overall PCV was 28.3% with no significant difference between the 2 groups (Welch 2-sample t-test, $P=0.18$). Pyrethroid administration with the footbath resulted in a significant reduction of the monthly trypanosomosis incidence rate ($P=0.003$). This effect lasted during the dry season, after the treatment period, and no new AAT infection was recorded in the treated herd (Figure 7B). The overall incidence of cattle trypanosomosis was reduced ($P=0.01$) from 0.76 (control group) to 0.11 (footbath treated group). A positive effect of the footbath treatment on packed-cell volume was also observed ($P<0.001$, see Bouyer et al. 2009) which proved a significant improvement of animal health (Bouyer et al. 2009).

These field studies demonstrated that restricted application of insecticides using footbaths allowed a significant reduction of tsetse populations, leading to a breakdown of trypanosome transmission cycle. Moreover, it reduced treatment cost and time by more than 90% compared with other techniques such as spraying of the full body of cattle. These results were also attributed to environmental conditions favourable to tsetse control in this area. Indeed, the scarcity of wild hosts but also the landscape fragmentation has favoured a fast reduction of tsetse sub-populations by reducing the speed of re-invasion of the cleared water points.

Unfortunately, these good results were not sustained. A socio-economic survey conducted in the same study site in 2008 at the beginning of the rainy season revealed that farmers have

abandoned the use of footbaths (Bouyer *et al.* 2011a). The main reason given was technical problems encountered to treat animals (whereas they did not during the experimental period), i.e. cattle were reluctant to pass in the footbaths. Later and in other sites, other farmers built more traditional cattle pens (with wood) instead of using metallic wire which greatly improved the treatment conditions. Moreover, footbaths manager were young educated people that have to count cattle during each treatment in order to calculate the price to pay for each herd (in order to buy new insecticides) and it conferred to them a strategic position which changed the former social relationships that farmers could not accept.

Therefore, the experience of a communal use of footbaths in this site was a failure despite the clear benefits associated to the technique, and an one year follow up by technicians offering long-term technical advice to the farmers. The main adoption factor was related to the economic dynamism of farmer associations. Actually, in another group of farmers using a similar farming system than the one studied here, but with more active farmer associations, a large proportion of the farmers adopted the technique (Bouyer *et al.* 2011a).

ITC in Uganda: the SOS program

This case study is based on several peer reviewed articles but two main data sources focusing on vector control and animal trypanosomosis were used (Morton 2010; Selby 2010). The Stamp Out Sleeping Sickness (SOS) program was a human health program, initiated mainly to fight sleeping sickness, which is not the priority of this work but it is a good case study for three reasons:

1. it combined mass treatment of cattle with curative drugs and restricted insecticide treatment of cattle (partial spraying);
2. the size of the initiative in terms of the number of cattle treated (>180,000);
3. the inclusion of the cattle farmers as one of the partners, together with researchers, Ugandan public authorities and the private sector (Ceva Santé Animale and IKARE).

In October 2006, an intervention was initiated to arrest the northerly advance through Uganda of the zoonotic parasite *Trypanosoma brucei rhodesiense* (Stephens & Fantham). The main goal of the SOS campaign was to target the cattle reservoir of *T. brucei rhodesiense* (agent of the acute form of sleeping sickness), in an areas of approximately 8,000 km² by treating >180,000 head of cattle (Selby 2010).

The field operations were conducted during 4 years between 2006 and 2010, and Morton (2010) reported the treatment of 'almost 180,000 cattle – belonging to around 50,000 households – with trypanocidal drugs and insecticidal sprays over five districts'. In this context, we will focus on the involvement of farmers in treating their cattle and the sustainability of the results and also in establishing whether or not this mass treatment of cattle has improved animal health, by controlling *T. congolense* and *T. vivax*, which were also present.

The control operations were subdivided in four rounds of treatments of cattle over 5 districts with trypanocides (isometamidium chloride and diminazene aceturate) and pyrethroid spray. Objectives of the first control session (November 2006) were to treat 86% of the estimated cattle population and to sensitise the farmers to pursue treatments on their own with support from District Veterinary Offices. In one month, 178,000 head of cattle were treated, corresponding to more than 100% of the initially estimated cattle population, but more probably to ~55% of the actual one (Selby 2010). The second round of control operations presented a lower coverage and dropped to approximately 30% of animals treated with strong variations of coverage observed

within and between districts despite provision of pyrethroid for two more rounds during the first round. Therefore, the third round (April/May 2007) was supervised by people from the Makerere University and was more successful (~80%). This low coverage was attributed to lack of communication about the insecticide efficacy because herders used other products, and distrust of the programme objectives (fear of taxation of cattle). Finally, a fourth round was implemented in a specific area where substantial cases of *T. brucei rhodesiense* were found both in cattle and human populations. Only 30,000 cattle were treated. In the other 5 districts, a new control strategy was implemented by five veterinarians sent to the field and in charge of sensitizing farmers and distributing products. Because of different problems lowering the farmers' investment capacities, this situation was not sustainable for veterinarian earning not enough money. Therefore, they started to sell different products, especially amitraz which is not efficient against tsetse. This product was considered more efficient against ticks by farmers due to its strong and visual knock-down effect on ticks whereas this effect was not visible with pyrethroids (the ticks become dry but remain on cattle; F. Stachurski, personal communication). Moreover, the initial price of the insecticide during the interventions was 0.02 US\$/treatment due to reduced or subsidized prices by the private partners, whereas the actual cost of the wholesale drugs was closer to 0.06 US\$/treatment (Shaw *et al.* 2013). This data on farmers' awareness and practice of regular spraying have been judged inconclusive. Indeed, by June 2009, 45% of cattle keepers had sprayed their animals in the last month' which was not dissimilar to the 40% who reported spraying their animals monthly in 2006 before the SOS project (Morton 2010).

Prior to the SOS project, the prevalence of *T. b. brucei* within the cattle reservoir was estimated by PCR at 15.57% (*T. brucei rhodesiense* as 0.81%) (Selby 2010). This prevalence was then monitored in 23 locations at three, nine and 18 months after the beginning of SOS program. A consistent number of cattle was sampled in the 23 sites during the survey (>1700 heads of cattle). The prevalence of *T. brucei. s.l.* dropped from 15 to 3% three months post intervention, but then returned at nine month and exceeded the baseline at 18 months (Figure 8).

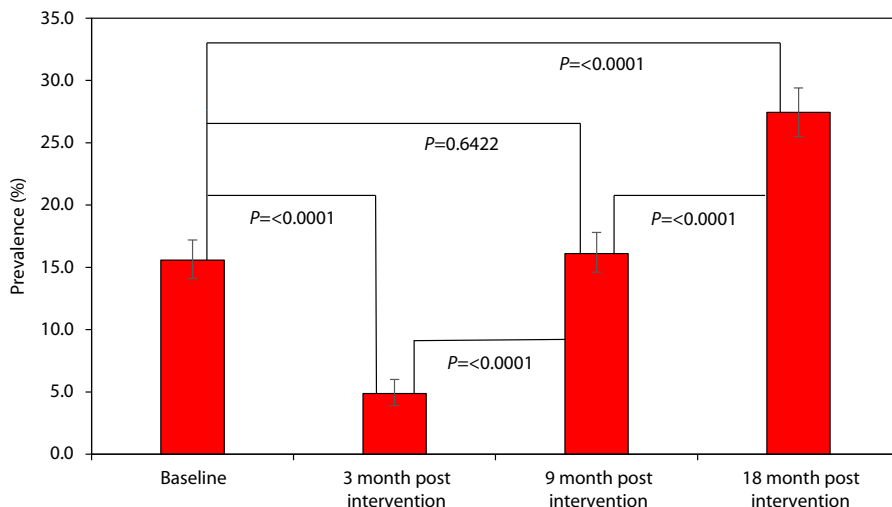


Figure 8. Prevalence of *Trypanosoma brucei. s.l.* during the SOS operation program (Selby 2010, with permission).

A similar pattern was observed for *T. brucei rhodesiense*. This increase in prevalence was attributed to the absence of sustained vector control. Although *T. congolense* and *T. vivax* were not directly studied during this program, some data was available before the beginning of the program and highlight that *T. congolense* was the most prevalent trypanosome species with 43% (of the animals tested; 15/35), followed by *T. brucei* and *T. vivax* with 34% (12/35) and 23% (8/35) respectively (Cox et al. 2010). This high prevalence indicated that animal trypanosomosis was a serious problem in this country. Therefore, the SOS program has probably lead to similar reduction in prevalence level in *T. congolense* and *T. vivax* as suggested by the increase of PCV in cattle.

Interview of the personnel implicated in the SOS program point out several reasons for the disappointing results such as poor community involvement; absence of crush in treatment sites (technical problem) and transport of animals to and from the treatment site (technical problem).

Therefore, the main conclusions of this program are that:

- Treating cattle with a partial spray of pyrethroid associated with an injection of diminazene did not succeed to control the transmission of *T. brucei. s.l.* in a sustainable way. This result is probably the consequence of a failure in the suppression of *G. fuscipes fuscipes*, the exclusive vector of this trypanosome species in the area. Because resistance to trypanocides has been previously highlighted (Matovu et al. 1997), it could partially explain the result. However, the temporary significant reduction of its prevalence 3 months after mass-treatment shows that *T. brucei* was at least partially susceptible to diminazene.
- The reduction in pyrethroid treatment cost due to the restricted application and subvention did not ensure good adoption by farmers.
- The technical problems related to the treatment sites and the associated time loss for farmers is probably responsible for the lack of commitment. It proves that even for traditional farmers, 'time is money'. Therefore the technical approaches could significantly be improved.
- The potential for improved control of human sleeping sickness was not considered important enough by the farmers to change their habits, and the apparent efficiency of the insecticide treatment against ticks seemed more important (since both molecules were probably as efficient but amitraz effect was more visible).

ITT in the Loos Islands of Guinea

This case study is based on two peer reviewed articles (Kagbadouno et al. 2009, 2011). In Guinea, the ministry of health has launched a control program against *G. palpalis gambiensis* (Diptera, Glossinidae) on three islands: Fotoba, Room and Kassa islands. Before the start of control operations, baseline entomological data were collected and revealed that tsetse populations from the Loos Islands presented a low rate of genetics exchange with flies from the mainland and also between islands (Camara et al. 2006; Solano et al. 2009). Therefore, these apparent isolated populations could be targeted using an area-wide integrated strategy with very low risk of reinvasion.

Different control operations were implemented on each island:

- in Fotoba island, impregnated targets were used at a density of 30 and then 60 targets/km²;
- in Room island, ground spraying and impregnated targets at a density of 30 targets/km² and then 60 targets/km² were used;
- in Kassa island, ground spraying, pour-on treatment of pigs (the major species bred on Kassa), fencing of pig pens with impregnated nets and targets were used at a density of 30 and then 60 targets/km².

Before control operation, average tsetse apparent densities were 10.33, 3 and 1.16 flies/trap/day in Kassa, Room and Fotoba, respectively. Three years after the beginning of control operations, tsetse apparent reduction was 100%, indicating that populations have been suppressed. This objective was achieved more or less faster according to measures implemented. The fully integrated strategy on Kassa allowed a very fast reduction of tsetse densities (>98% within 6 months) and similar results were obtained in Room island (>97% within 6 months; Figure 9).

The surrounding of pig pens with insecticide mosquito net proved particularly efficient on Kassa. However, on Fotoba using only blue targets set at the density of 30/km² was not sufficient to reduce tsetse densities below 50% and target density was therefore increased to 60/km² after one year of control, which allowed a further reduction of densities to undetectable levels within 6 months. The success of control operation was also due to strong community sensitization and participation. The farmers appeared cooperative and the targets and insecticide mosquito nets were well maintained. However, all these techniques were provided free of charge by the national program, and there was no transfer of the control operations to the farmers. Apart from ground spraying, all other techniques used during this control campaign would be suitable for farmer-based vector control. The main limitation was that targets were considered as public goods, and thus pour-on and insecticide mosquito nets would probably have better chances to be adopted. On these islands, the programme was not sustainable and tsetse densities recovered within a few years after the end of the intervention of national authorities.

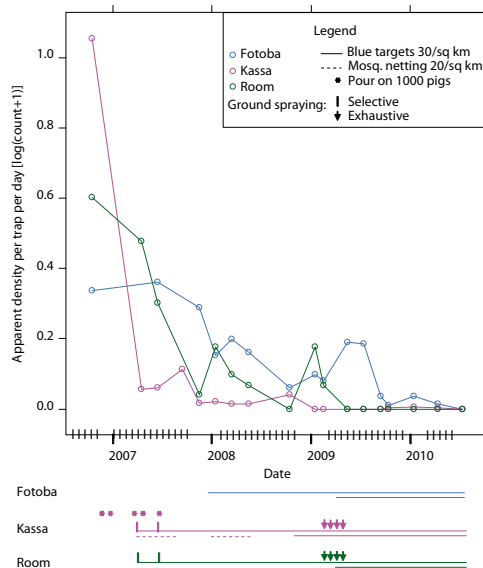


Figure 9. Evolution of tsetse apparent density in Loos Islands according to different control operations (Kagbadouno et al. 2011).

Use of fencing to protect zero grazing units in western Kenya

This case study is based on the peer reviewed article from Bauer *et al.* (2006). The epidemiological cycle is the endemic AAT cycle and the type of cattle rearing system is zero grazing units (situation BIII).

In Kenya, 80% of 3 million dairy cows are the property of small-scale farmers, of which a large part are confined to zero grazing units with all food and water provided to the animals. The farmers use trypanocidal drugs (diminazene aceturate for therapy or isometamidium chloride for prophylaxis (Bauer *et al.* 2006) but they still experience trypanosomosis cases (adults mortalities, abortions, stillbirths and a reduction of milk production). Four tsetse species occur in western Kenya: *G. fuscipes fuscipes*, *G. pallidipes*, *Glossina brevipalpis* Newstead and *Glossina swynertonii* Austen at generally low densities (<1 fly per trap per day) (case IIIB of Figure 9), except in Busia District along fringing vegetation, which borders the shores of Lake Victoria, where the densities can be up to 30 per trap per day and in protected areas (Cecchi *et al.* 2015) (case IIB of Figure 9).

In this context, Bauer *et al.* (2006) tested the impact of surrounding the zero-grazing units with insecticide-treated mosquito netting to a height of 150 cm to protect the cows against tsetse and AAT. Fifty-seven randomly selected dairy units were protected with netting and another 42 randomly selected units served as controls. The dominant cattle breeds in the trial were Holstein-Friesian and Ayrshire, which are highly trypano sensitive breeds. The authors used black mosquito netting, impregnated with pyrethroid. At the start of the experiment, all cattle (except those in late-stage pregnancy) were given a therapeutic dose of diminazene aceturate (3.5 mg/kg) and also during the trial in case of positive diagnosis of AAT or hematocrit below 25%.

The insecticide-treated netting significantly reduced the risk of trypanosome infection in cattle and significantly increased the mean haematocrit (from 27.6 ± 0.6 to 29.7 ± 0.4 ; $P < 0.05$) (Bauer *et al.* 2006).

Just after setting the net, no more flies were detected in the protected pens and cows remained calm during milking. Farmers also reported a reduction in the number of biting flies, mosquitoes and house flies in their homesteads. However, no differences in milk production between the experimental and control groups were observed despite immediate increases in milk production in individual cows, probably because of other constraints (possibly food).

The authors observed that the netting did not last longer than 2 months due to destruction by strong wind or animal movements. Also many farmers let their cattle graze freely outside the units during the day, despite technical advice, resulting in exposure of animals to habitats suitable for tsetse. Although these problems led to increased AAT incidence in comparison to animals under continuous protection, it still offered an advantage in comparison to a system with no protection.

In conclusion, fencing farms with insecticidal netting can provide efficient and cost effective protection of zero-grazed cattle but its adoption by farmers is not yet warranted. Indeed, it requires modification of the rearing system, especially for free-grazing system and only a minority of the farmers accepted to change their habits. In this case, the benefit was probably lower because the cattle will be only partially protected. Moreover, in this trial, the pens were totally surrounded with insecticide-treated netting, which is sometime not possible in modern farms (see above). Fencing cattle pens will benefit farmers as they will spend less money for trypanocidal drugs and in the same time, it will limit the emergence of drug resistance of the exposed trypanosome populations.

Conclusions

The aim of this review was to present the different vector control methods available for farmers in the fight against AAT. To be sustainable, integrated control strategies of trypanosomosis must be context dependent, mainly based on the environment, cattle breed and rearing system. More importantly, the success of farmer based control operation is highly dependent on the cost-effectiveness and user friendliness of tools proposed, which will increase their acceptability in the community. As we have seen in different study cases, sensitisation and implication of farmers is one of the key to succeed in control operations. Moreover, new vector control technologies such as tiny targets, impregnated fences or restricted treatments must be more widely adopted by stakeholders and in the same time important effort must be done by veterinarian offices to train community farmer in their use. The availability of new pour-on treatment offering individual protection against AAT is a new promising development. However, this is not a warranty for success and the different tools proposed should be adapted to each particular situation and rearing system in collaboration with the farmers (co-building of innovative control strategy) (see chapter on vector control acceptability in this book). In the context of increasing resistance of trypanosomes to both to diminazene and isometamidium there is a urgent need to implement a control strategy including vector control, particularly in the sites where these resistances have a high prevalence.

References

- Ahmed SK, Rahman AH, Hassan MA, Salih SEM, Paone M and Cecchi G (2016) An atlas of tsetse and bovine trypanosomosis in Sudan. *Parasite Vectors* 9: 194.
- Bauer B, Gitau D, Oloo FP and Karanja SM (2006) Evaluation of a preliminary trial to protect zero-grazed dairy cattle with insecticide-treated mosquito netting in western Kenya. *Trop Anim Health Prod* 38: 29-34.
- Bauer B, Holzgrefe B, Mahama CI, Baumann MP, Mehlitz D and Clausen PH (2011) Managing tsetse transmitted trypanosomosis by insecticide treated nets – an affordable and sustainable method for resource poor pig farmers in Ghana. *PLoS Negl Trop Dis* 5: e1343.
- Bett B, Randolph TF, Irungu P, Nyamwaro SO, Kitale P, Gathuma J, Grace D, Vale G, Hargrove J and McDermott J (2010) Field trial of a synthetic tsetse-repellent technology developed for the control of bovine trypanosomosis in Kenya. *Prev Vet Med* 97: 220-227.
- Bouyer F, Bouyer J, Seck MT, Sall B, Dicko AH, Lancelot R and Chia E (2015) Importance of vector-borne infections in different production systems: bovine trypanosomosis and the innovation dynamics of livestock producers in Senegal. *Rev Sci Tech Off Int Epiz* 34: 213-225.
- Bouyer F, Hamadou S, Adakal H, Lancelot R, Stachurski F, Belem AMG and Bouyer J (2011a) Restricted application of insecticides: a promising tsetse control technique, but what do the farmers think of it? *PLoS Negl Trop Dis* 5: e1276.
- Bouyer J and Marois E (2018) Genetic control of vectors. In: Garros C, Bouyer J, Takken W and Smallegange RC (eds.) *Pests and vector-borne diseases in the livestock industry. Ecology and Control of Vector-borne diseases. Vol. 5.* Wageningen Academic Publishers, Wageningen, the Netherlands, pp. 435-451.
- Bouyer J, Bouyer F, Donadeu M, Rowan T and Napier G (2013) Community- and farmer-based management of animal African trypanosomosis in cattle. *Trends Parasitol* 29: 519-522.
- Bouyer J, Grimaud Y, Pannequin M, Esnault O and Desquesnes M (2011b) Importance épidémiologique et contrôle des stomoxes à la Réunion. *Bull Epidémiol* 43: 53-58.
- Bouyer J, Solano P, Cuisance D, Itard J, Frézil J-L and Authié E (2010) Trypanosomosis: control methods. In: Lefèvre P-C, Blancou J, Chermette R and Uilenberg G (eds.) *Infectious and parasitic diseases of livestock. Éditions Lavoisier (Tec & Doc), Paris, France, pp. 1936-1943.*
- Bouyer J, Stachurski F, Gouro AS and Lancelot R (2008) On-station cattle insecticide treatment against tsetse flies using a footbath. *Rev Elev Méd Vét Pays Trop* 61: 153-160.

- Bouyer J, Stachurski F, Gouro AS and Lancelot R (2009) Control of bovine trypanosomosis by restricted application of insecticides to cattle using footbaths. *Vet Parasitol* 161: 187-193.
- Bouyer J, Stachurski F, Kaboré I, Bauer B and Lancelot R (2007) Tsetse control in cattle from pyrethroid footbaths. *Prev Vet Med* 78: 223-238.
- Camara M, Caro-Riaño H, Ravel S, Dujardin J-P, Hervouet J-P, De Meeüs T, Kagbadouno MS, Bouyer J and Solano P (2006) Genetic and morphometric evidence for population isolation of *Glossina palpalis gambiensis* (Diptera: Glossinidae) on the Loos Islands, Guinea. *J Med Entomol* 43: 853-860.
- Cecchi G, Paone M, Argilés Herrero R, Vreysen MJB and Mattioli RC (2015) Developing a continental atlas of the distribution and trypanosomal infection of tsetse flies (*Glossina* species). *Parasite Vectors* 8: 284.
- Cox AP, Tosas O, Tilley A, Picozzi K, Coleman P, Hide G and Welburn SC (2010) Constraints to estimating the prevalence of trypanosome infections in East African zebu cattle. *Parasite Vectors* 3: 82.
- Cuisance D, Cailton P, Kota Guinza A, Ndokoué F, Pounekrozou E and Demba D (1991) Lutte contre *Glossina fuscipes fuscipes* par piégeage chez les éleveurs Mbororo de République Centrafricaine. *Rev Elev Méd Vét Pays Trop* 44: 81-89.
- Dayo GK, Gautier M, Berthier D, Poivey JP, Sidibe I, Bengaly Z, Eggen A, Boichard D and Thevenon S (2012) Association studies in QTL regions linked to bovine trypanotolerance in a West African crossbred population. *Anim Genet* 43: 123-132.
- De Garine-Wichatitsky M, Cheke RA and Lazaro D (2001) Effects of tsetse targets on mammals and birds in Kasungu National Park, Malawi. *Biodivers Conserv* 10: 869-891.
- Eisler MC, Torr SJ, Coleman PG, Machila N and Morton JF (2003) Integrated control of vector-borne diseases of livestock – pyrethroids: panacea or poison? *Trends Parasitol* 19: 341-345.
- Esterhuizen J, Rayaisse JB, Tirados I, Mpiana S, Solano P, Vale GA, Lehane MJ and Torr SJ (2011) Improving the cost-effectiveness of visual devices for the control of riverine tsetse flies, the major vectors of human African Trypanosomiasis. *PLoS Negl Trop Dis* 5: e1257.
- Faulde MK and Nehring O (2012) Synergistic insecticidal and repellent effects of combined pyrethroid and repellent-impregnated bed nets using a novel long-lasting polymer-coating multi-layer technique. *Parasitol Res* 111: 755-765.
- Geerts S, Holmes PH, Eisler MC and Diall O (2001) African bovine trypanosomiasis: the problem of drug resistance. *Trends Parasitol* 17: 25-28.
- Gimonneau G, Alioum Y, Abdoulmoumini M, Zoli A, Cene B, Adakal H and Bouyer J (2016) Insecticide and repellent mixture pour-on protects cattle against animal trypanosomosis. *PLoS Negl Trop Dis* 10: e0005248.
- Hargrove JW (2003) Optimized simulation of the control of tsetse flies *Glossina pallidipes* and *G-m. morsitans* (Diptera : Glossinidae) using odour-baited targets in Zimbabwe. *Bul Entomol Res* 93: 19-29.
- Hargrove JW, Omolo S, Msalilwa JSI and Fox B (2000) Insecticide-treated cattle for tsetse control: the power and the problems. *Med Vet Ent* 14: 123-130.
- Hargrove JW, Torr SJ and Kindness HM (2003) Insecticide-treated cattle against tsetse (Diptera: Glossinidae): what governs success? *Bull Entomol Res* 93: 203-217.
- Itard J, Cuisance D and Tacher G (2003) Trypanosomoses: historique – répartition géographique. In: Lefèvre P-C, Blancou J and Chermette R (eds.) Principales maladies infectieuses et parasitaires du bétail. Europe et Régions Chaudes, Lavoisier, Paris, France, pp. 1607-1615.
- Kabayo JP (2002) Aiming to eliminate tsetse from Africa. *Trends Parasitol* 18: 473-475.
- Kagbadouno M, Camara M, Bouyer J, Courtin F, Onikoyamou M, Schofield C and Solano P (2011) Progress towards the eradication of tsetse from the Loos Islands, Guinea. *Parasite Vectors* 4: 18.
- Kagbadouno M, Camara M, Bouyer J, Hervouet JP, Courtin F, Jamonneau V, Morifaso O, Kaba D and Solano P (2009) Tsetse elimination: its interest and feasibility in the historical sleeping sickness focus of Loos Islands, Guinea. *Parasite* 16: 29-35.
- Kamuanga M, Sigue H, Swallow B, Bauer B and d'Ieteren G (2001a) Farmers' perceptions of the impacts of tsetse and trypanosomosis control on livestock production: evidence from southern Burkina Faso. *Trop Anim Health Prod* 33: 141-153.
- Kamuanga M, Swallow BM, Sigué H and Bauer B (2001b) Evaluating contingent and actual contributions to a local public good: tsetse control in the Yale agro-pastoral zone, Burkina Faso. *Ecological Economics* 39: 115-130.

- Kernaghan RJ and Johnston MRL (1962) A method of determining insecticide persistence in tsetse fly control operations. *Bul World Health Org* 26: 139-141.
- Kgori PM, Modo S and Torr SJ (2006) The use of aerial spraying to eliminate tsetse from the Okavango Delta of Botswana. *Acta Trop* 99: 184-199.
- Knols BGJ, Willemse L, Flint S, and Mate A (1993). A trial to control the tsetse fly, *Glossina morsitans centralis*, with low densities of odour-baited targets in west Zambia. *Med Vet Entomol* 7: 161-169.
- Laveissière C, Couret D and Kiéno J-P (1980) Lutte contre les glossines riveraines à l'aide de pièges biconiques imprégnés d'insecticides, en zone de savane humide: 1. Description du milieu, du matériel et de la méthode. *Cahiers ORSTOM Série Entomol Méd Parasitol* 18: 201-207.
- Lindh JM, Torr SJ, Vale GA and Lehane MJ (2009) Improving the cost-effectiveness of artificial visual baits for controlling the tsetse fly *Glossina fuscipes fuscipes*. *PLoS Negl Trop Dis* 3: e474.
- Maia M, Clausen PH, Mehlitz D, Garms R and Bauer B (2010) Protection of confined cattle against biting and nuisance flies (Muscidae: Diptera) with insecticide-treated nets in the Ghanaian forest zone at Kumasi. *Parasitol Res* 106: 1307-1313.
- Maia MF, Abonuuusum A, Lorenz LM, Clausen PH, Bauer B, Garms R and Kruppa T (2012) The effect of deltamethrin-treated net fencing around cattle enclosures on outdoor-biting mosquitoes in Kumasi, Ghana. *PLoS ONE* 7: e45794.
- Matovu E, Iten M, Enyaru JC, Schmid C, Lubega GW, Brun R and Kaminsky R (1997) Susceptibility of Ugandan *Trypanosoma brucei rhodesiense* isolated from man and animal reservoirs to diminazene, isometamidium and melarsoprol. *Trop Med & Intern Health* 2: 13-18.
- Morton J (2010) The innovation trajectory of sleeping sickness control in Uganda. Research knowledge in its context. Research Into Use Programme, UK Department for International Development, London, UK, 43 pp. Available at: <https://tinyurl.com/y74lhd6j>.
- Ndeledje N, Bouyer J, Stachurski F, Grimaud P, Belem AMG, Mbaïndingatoloum FM, Bengaly Z, Cecchi G and Lancelot R (2013) Treating cattle to protect people? Impact of footbath insecticide treatment on tsetse density in Chad. *PLoS ONE* 8: e67580.
- Pagabeleguem S, Sangaré M, Bengaly Z, Akoudjin M, Belem AMG and Bouyer J (2012) Climate, cattle rearing systems and African animal trypanosomosis risk in Burkina Faso. *PLoS ONE* 7: e49762.
- Rayaisse JB, Esterhuizen J, Tirados I, Kaba D, Salou E, Diarrassouba A, Vale GA, Lehane MJ, Torr SJ and Solano P (2011) Towards an optimal design of target for tsetse control: comparisons of novel targets for the control of *palpalis* group tsetse in West Africa. *PLoS Negl Trop Dis* 5: e1332.
- Rayaisse JB, Tirados I, Kaba D, Dewhirst SY, Logan JG, Diarrassouba A, Salou E, Omolo MO, Solano P, Lehane MJ, Pickett JA, Vale GA, Torr SJ and Esterhuizen J (2010) Prospects for the development of odour baits to control the tsetse flies *Glossina tachinoides* and *G. palpalis s.l.* *PLoS Negl Trop Dis* 4: e632.
- Selby R (2010) Limiting the northerly advance of *Trypanosoma brucei rhodesiense* in post conflict Uganda. PhD thesis, University of Edinburgh, Edinburgh, UK.
- Shaw APM, Tirados I, Mangwiro CTN, Esterhuizen J, Lehane MJ, Torr SJ and Kovacic V (2015) Costs of using 'Tiny Targets' to control *Glossina fuscipes fuscipes*, a vector of *Gambiense* sleeping sickness in Arua district of Uganda. *PLoS Negl Trop Dis* 9: e0003624.
- Shaw APM, Torr SJ, Waiswa C, Cecchi G, Wint GRW, Mattioli RC and Robinson TP (2013) Estimating the costs of tsetse control options: an example for Uganda. *Prev Vet Med* 110: 290-303.
- Solano P, Ravel S, Bouyer J, Camara M, Kagbadouno MS, Dyer N, Gardes L, Herauld D, Donnelly MJ and De Meeûs T (2009) The population structure of *Glossina palpalis gambiense* from island and continental locations in coastal Guinea. *PLoS Negl Trop Dis* 3: e392.
- Stachurski F (2006) Attachment kinetics of the adult tick *Amblyomma variegatum* to cattle. *Med Vet Entomol* 20: 317-324.
- Stachurski F and Lancelot R (2006) Footbath acaricide treatment to control cattle infestation by the tick *Amblyomma variegatum*. *Med Vet Entomol* 20: 402-412.
- Tadesse A, Hadgu E, Mekbib B, Abebe R and Mekuria S (2011) Mechanically transmitted bovine Trypanosomosis in Tselemti Woreda, western Tigray, northern Ethiopia. *Agri J* 6: 10-13.

- Torr SJ (1985) The susceptibility of *Glossina pallidipes* Austen (Diptera, Glossinidae) to insecticide deposits on targets. *Bul Entomol Res* 75: 451-458.
- Torr SJ, Chamisa A, Vale GA, Lehane MJ and Lindh JM (2011) Responses of tsetse flies, *Glossina morsitans morsitans* and *Glossina pallidipes*, to baits of various size. *Med Vet Entomol* 25: 365-369.
- Torr SJ and Hargrove JW (1998) Factors affecting the landing and feeding responses of the tsetse fly *Glossina pallidipes* to a stationary ox. *Med Vet Entomol* 12: 196-207.
- Torr SJ, Maudlin I and Vale GA (2007) Less is more: restricted application of insecticide to cattle to improve the cost and efficacy of tsetse control. *Med Vet Entomol* 21: 53-64.
- Vale G and Torr S (2004) Development of bait technology to control tsetse. In: Maudlin I, Holmes P and Miles M (eds.) *The Trypanosomiasis*. CABI Publishing, Wallingford, pp. 509-523.
- Vale GA (1980) Field studies of the responses of tsetse flies (Glossinidae) and other diptera to carbon-dioxide, acetone and other chemicals. *Bul Entomol Res* 70: 563-570.
- Vale GA, Grant IF, Dewhurst CF and Aigreau D (2004) Biological and chemical assays of pyrethroids in cattle dung. *Bul Entomol Res* 94: 273-282.
- Vale GA, Mutika G and Lovemore DF (1999) Insecticide-treated cattle for controlling tsetse flies (Diptera: Glossinidae): some questions answered, many posed. *Bul Entomol Res* 89: 569-578.
- Van den Bossche P, De la Rocque S, Hendrickx G and Bouyer J (2010) A changing environment and the epidemiology of tsetse-transmitted livestock trypanosomiasis. *Trends Parasitol* 26: 236-243.
- Van den Bossche P and Delespaulx V (2011) Options for the control of tsetse-transmitted livestock trypanosomiasis. An epidemiological perspective. *Vet Parasitol* 181: 37-42.
- Vreysen MJB, Robinson AS and Hendrichs J (2007) *Area-wide control of insect pests. From research to field implementation*. Springer, Dordrecht, the Netherlands.
- Vreysen MJB, Saleh KM, Ali MY, Abdulla AM, Zhu ZR, Juma KG, Dyck VA, Msangi AR, Mkonyi PA and Feldmann HU (2000) *Glossina austeni* (Diptera: Glossinidae) eradicated on the island of Unguja, Zanzibar, using the sterile insect technique. *J Econ Entomol* 93: 123-135.
- Vreysen MJB, Seck MT, Sall B and Bouyer J (2013) Tsetse flies: their biology and control using area-wide integrated pest management approaches. *J Invertebr Pathol* 112, Suppl. 1: S15-S25.
- Willemsse LPM (1991) A trial of odour baited targets to control the tsetse fly, *Glossina morsitans centralis* (Diptera: Glossinidae) in west Zambia. *Bull Entomol Res* 81: 351-357.